



Gold bipyramids as a promising alternative to gold nanorods for analytical and biomedical applications

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Abstract

Gold nanobipyramids and nanorods of comparable dimensions were synthesized to evaluate and contrast their plasmonic, analytical, and photothermal performance. The bipyramids exhibited a markedly higher absorption spectral quality factor than the nanorods. For surface-enhanced Raman scattering comparison, both nanoparticle types were functionalized with thiolated nitrobenzene molecules, revealing an approximately fourfold higher enhancement factor for the bipyramids. Their analytical sensitivity was further assessed by monitoring the plasmon resonance wavelength shift in response to changes in the external refractive index, as well as variations in the thickness and refractive index of a nanoscale coating. In all cases, the bipyramids outperformed the nanorods. Additionally, both experimental and theoretical analyses confirmed the superior efficiency of bipyramids as photothermal agents for converting laser irradiation into heat. Effective photothermal inactivation of *Escherichia coli* was achieved under resonant laser excitation using either particle type. Given the recent advances in their synthesis, gold bipyramids emerge as highly promising alternatives to nanorods for a broad range of analytical and biomedical applications.

Introduction

Anisotropic metal nanoparticles [1-5] are actively researched and utilized due to their flexible tunability of the plasmon resonance (PR) across a broad UV–vis–NIR range [6], significantly

larger scattering and absorption cross sections [7], and enhanced local field effects [8] compared to plasmonic nanospheres. Modern methods of colloidal chemical synthesis [9]

enable the production of a wide variety of anisotropic nanoparticles, including gold–silver nanocages [10], silver nanocubes [11], gold nanostars [12], triangular nanoprisms [13], flat particles [14], gold–silver nanorods [15], as well as particles with field enhancement in gaps (gap-enhanced Raman tags) [16]. However, the most well-known and widely used are conventional gold nanorods [17] with approximately hemispherical ends and a cylindrical or pentagonal cross section. To date, the synthesis and application protocols for gold nanorods are well established [18–21], which explains the consistently high annual number of publications on these nanoparticles (according to SCOPUS, over 180 articles in 2025 alone have titles containing the phrase “gold nanorods”).

Compared to nanorods, pentagonal gold nanobipyramids (BPs) are far less studied [22], despite having similar plasmon resonance tunability [23] (with an aspect ratio change from 2 to 6, the plasmon resonance wavelength shifts from approximately 600 to 1100 nm) and thermoplasmonic properties [24–26]. Regarding the differences, BPs possess five uniformly distributed twinning planes at the vertices and sharp tips [27], whereas nanorods are single crystals with roughly hemispherical ends. These morphological differences result in distinct local plasmonic properties, higher resonance sensitivity to the dielectric environment [28–30], and higher quality factors of the plasmonic absorption and scattering peaks in BPs. Despite these apparent advantages, BPs remain, in our opinion, undeservedly less popular due to their non-trivial synthesis [23,31–35] and functionalization [36]. Furthermore, simulating the optical properties of BPs (especially when coated with functional molecules) requires numerical methods [37,38] as until very recently there were no simple and sufficiently accurate analytical models for plasmonic nanoparticles with a dielectric coating [39].

In this work, we investigate how differences in the morphology of gold nanorods and bipyramids lead to vast differences in local field distribution and, through these differences, to multimodal enhancements in extinction quality spectra, surface-enhanced Raman scattering (SERS), sensing, and photothermal response. Specifically, we present a comparative theoretical and experimental study of UV–vis extinction spectra, SERS, refractive index sensitivity, thermoplasmonic properties of gold BPs and nanorods with similar PR wavelengths, and plasmonic photothermal killing of bacteria in water suspensions. In contrast to published similar studies (see, e.g., [40,41]), we compared the normalized quantities that are proportional to the SERS intensity per adsorbed molecule. For comparison of thermoplasmonic experiments, we introduced the normalized absorption efficiency, which is proportional to the absorbed power per unit volume at a constant mass/volume concentration of particles.

Finally, our analysis of analytical sensitivity includes two different options, that is, (1) the usual sensitivity to the refractive index of the surrounding medium and (2) an original analysis of the analytical response to an adsorbed layer of analytes with different nanometer-scale thicknesses.

Using the chemical etching method [42], we synthesized a set of nanorod samples with PR ranging from 600 to 950 nm, including a sample with a PR close to that of the BP sample. To calculate the plasmonic properties of the nanoparticles, we used a previously described 2.5D finite element method [43,44] implemented in the commercial software package COMSOL Multiphysics 5.1 (Wave Optics module), applicable to axially symmetric particles. Also, we used our recently developed analytical method [39].

We demonstrate that BP properties, including the quality factor of extinction spectra, SERS signal enhancement, the analytical sensitivity of the PR wavelength to the local dielectric environment, and the efficiency of light-to-heat conversion, surpass those of gold nanorods with comparable dimensions and PR wavelengths. Experiments also demonstrate effective photothermal killing of *Escherichia coli* DH5 α bacteria under laser irradiation at a wavelength near the PR.

Methods

Theoretical methods

In the 2.5D formalism [43,44], the incident and scattered fields are expanded in cylindrical harmonics, and the response of each harmonic is computed separately, leading to significant savings in computational resources in terms of memory and processing time [45]. Compared to conventional 3D modeling, the 2.5D formalism substantially reduces the computational time per spectral data point and enables averaging cross-sectional spectra over random particle orientations with acceptable computational costs. The dielectric function of gold was calculated using a spline interpolation [46] of the data from [47]. In all calculations presented hereafter, the correction of the bulk dielectric function for the finite particle size was performed using the volume-equivalent sphere radius as the effective electron surface scattering length and the scattering constant $A_s = 0.33$ [48,49]. The refractive index of the shell, $n_2 = \sqrt{\epsilon_2} = 1.5$, was chosen to be close to that of many biopolymers and typical stabilizers, such as cetyltrimethylammonium bromide (CTAB) or cetyltrimethylammonium chloride (CTAC). In all calculations, the dielectric function of water [44] $\epsilon_m(\lambda) = n_m^2(\lambda)$ was used as the external medium dielectric function. The thickness of the CTAB stabilizer shell was set to 3 nm based on the data from [50]. The employed analytical method MEM+DEM [39] is based on a combination of the modal expansion method (MEM) [51] and the dipole equivalence method (DEM) [52]. Hereafter,

for brevity, we will refer to the MEM+DEM method simply as MEM. The primary theoretical models considered were pentagonal bipyramids with tip rounding radii specified in [51] and cylindrical nanorods with hemispherical ends. We also employed an inscribed bicone as a bipyramid model, following the approach in [38].

Experimental methods

The following reagents were used in this study: hydrogen tetrachloroaurate(III) trihydrate ($\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$), cetyltrimethylammonium bromide (CTAB, 98%), cetyltrimethylammonium chloride (CTAC, 25% solution in water), sodium borohydride (NaBH_4 , 99%), silver nitrate (AgNO_3 , 99%), hydrochloric acid (concentrated HCl), L-ascorbic acid (AA, 99.9%), citric acid (CA, 99%), 4-nitrothiophenol (NBT, 97%), and hydroquinone (HQ, 99%). Ultrapure deionized water obtained using a Milli-Q Integral 5 system was used in all experimental syntheses.

Extinction spectra were measured using a Specord 300 spectrophotometer (Analytik Jena, Germany). The Raman spectra of composite labels were recorded using a combined setup comprising a Leica MD 2500 microscope, a SeekerPro785 spectrometer from Ocean Optics (USA), and a 785 nm laser operating at 30 mW. The surface potentials (charges) of the nanoparticles were measured using a Zetasizer NanoSeries NT dynamic light scattering spectrometer (Malvern Instruments, UK). Nanoparticle images were obtained with a Libra-120 transmission electron microscope (TEM) (Carl Zeiss, Jena, Germany) at the “Symbiosis” Center for the Collective Use of Research Equipment (IBPPM RAS). A diode laser with a wavelength of 795–810 nm, fiber optic output, and a maximum continuous output power of 2 W (Opto Power Corp., USA) was used to heat nanoparticle suspensions. The temperature in the cuvette was measured using a Guide PC210 thermal imager (China).

Gold nanobipyramids were obtained by overgrowth of polycrystalline seeds in a CTAB medium [32]. In the first, preliminary stage, gold thermal seeds were synthesized. For this, 10 mL of 0.25 mM $\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$ were reduced with 0.25 mL of 25 mM sodium borohydride under vigorous stirring in a 20 mL flask, in an aqueous solution containing 50 mM CTAC and 5 mM citric acid. The reaction mixture changed color from yellow to brown. After 2 min, the solution was transferred to a silicone bath at 80 °C and stirred for 90 min. During the reaction, the solution turned deep red. In the second stage, the obtained seeds, cooled to room temperature, were added under vigorous stirring to a solution containing 50 mL of 100 mM CTAB, 2.5 mL of 10 mM $\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$, 500 μL of 10 mM silver nitrate, 1 mL of 1 M hydrochloric acid, and 400 μL of 100 mM ascorbic acid. The mixture was kept at 30 °C for about

2 h. The resulting bipyramids were centrifuged several times (10,000 rpm) and redissolved in 1 mM CTAC. The synthesis yielded gold bipyramids with an average longitudinal size of 75 ± 3 nm, a transverse size of 25 ± 2 nm, and a plasmon resonance maximum around 753 nm.

Gold nanorods were obtained using a modified protocol [53] that enabled the synthesis of thin, long rods. Gold seeds were prepared by adding an aqueous sodium borohydride solution (10 mM, 0.6 mL) to an aqueous solution containing CTAB (0.1 M, 10 mL) and HAuCl_4 (10 mM, 0.25 mL). Silver nitrate (3.5 mL, 0.1 M) was added to a solution of HAuCl_4 (500 mL, 0.5 mM) in 0.1 M CTAB, followed by the addition of an aqueous hydroquinone solution (25 mL, 0.1 M). Note that silver nitrate plays multiple, intertwined roles in the seed-mediated synthesis of single-crystalline gold nanorods [54,55]. Its functions span three critical areas, namely, (1) symmetry breaking and nucleation kinetics, (2) anisotropic growth directionality and aspect ratio control, and (3) shape and facet regulation via surface passivation.

The resulting mixture was stirred manually until it became clear. Then, 8 mL of the seeds were added, and the mixture was stirred and left overnight at 30 °C without stirring. A significant drawback of the protocol [53] is the high impurity particle content (10–15%). In this work, we used a modified version [56] with a purification step: the colloid was centrifuged at 10,000 rpm for 20 min, and the pellet was redissolved in 30 mL of 200 mM CTAC and left undisturbed for 1 h. The nanorods aggregated at the bottom and the walls of the tube, forming a brown film. The supernatant, containing particles of other shapes, was removed. The nanorods were redissolved in 50 mM CTAB to a concentration corresponding to an optical density of 2 at the longitudinal resonance at 945 nm, measured in a 2 mm path length cuvette. To completely remove impurity particles, the purification procedure was repeated twice. TEM analysis of the samples revealed the presence of thin nanorods with an average thickness of about 20 nm and a length of about 95 nm. A distinctive feature of our protocol is the practically zero percentage of non-target particles (0.5%) and the high quality of the extinction spectrum, as measured by the extinction ratio at the longitudinal and transverse PR wavelengths (>6).

To obtain AuNR samples with different longitudinal plasmon resonance (LPR) wavelengths, 50 mL of AuNR colloid in 50 mM CTAB was titrated with a 2 mM HAuCl_4 solution (adding 20 μL aliquots every 10 min). Before each addition, the extinction spectrum was measured, and if the desired LPR wavelength was reached, the required amount of sample (typically 3 mL) was taken from the colloid.

For the functionalization of gold nanorods and bipyramids with 4-nitrothiophenol (NBT) molecules, 10 μL of a 2 mM ethanolic NBT solution was added to 1.5 mL of the colloid. The mixture was incubated for 30 min. The nanoparticles were then centrifuged at 10,000 rpm for 5 min and resuspended in 2 mL of 10 mM CTAC.

For photothermal experiments, a 24-hour bacterial culture of *Escherichia coli* DH5 α (from the collection of IBFPM RAS) with an optical density $A_6 = 1$ in a 1 cm cuvette and gold nanorod and nanobipyramid suspensions with an optical density of $A_{80} = 5$ were used. A 0.01 M PBS buffer with pH 7.5 was used as the dilution medium. For the Alamar Blue assay [57], a resazurin solution at 0.55 $\text{mg}\cdot\text{mL}^{-1}$ was used. Fluorescence spectra were recorded on a Cary Eclipse spectrofluorimeter (Agilent) over 590–610 nm, with a slit width of 5 nm and an excitation wavelength of 530 nm.

Results and Discussion

1 Extinction spectra and geometrical parameters of nanoparticles

In Figure 1A, solid lines represent the extinction spectra of four nanorod samples with LSPR at 945, 844, 735, and 644 nm (curves 1–4), as well as the spectrum of gold bipyramids (5); the inset shows a TEM image of the bipyramids. Additional and statistical data can be found in Supporting

Information File 1 (Figures S1–S5). Dashed lines show the spectra of the same samples after functionalization with the Raman reporter molecule NBT. Figure 1B–E shows TEM images of the initial 944 nm nanorods and of samples obtained through chemical length etching with virtually no change in thickness. The geometric parameters of the particles, based on statistical analysis of TEM images, are given below in Table 1.

As shown in Table 1, during the etching process, the length of the nanorods decreases from 95 to 45 nm, and the aspect ratio decreases on average from approximately 4.85 to 2.4, while the nanorod diameter remains practically constant at 19 ± 0.8 nm. Accordingly, the plasmon resonance wavelength shifts from the infrared (945 nm) to the visible (640 nm) region. The spectra in Figure 1 were measured at approximately equal molar concentrations of gold (0.1 mM), as determined using the method described in [58].

In Table 1, the theoretical AR_p values were calculated by the linear equation

$$\lambda_{\text{PR}} (\text{nm}) = 111 \times \text{AR} + 393, \quad (1)$$

which was obtained as the average of T-matrix simulations for bare AuNRs and COMSOL simulations [39] for AuNRs

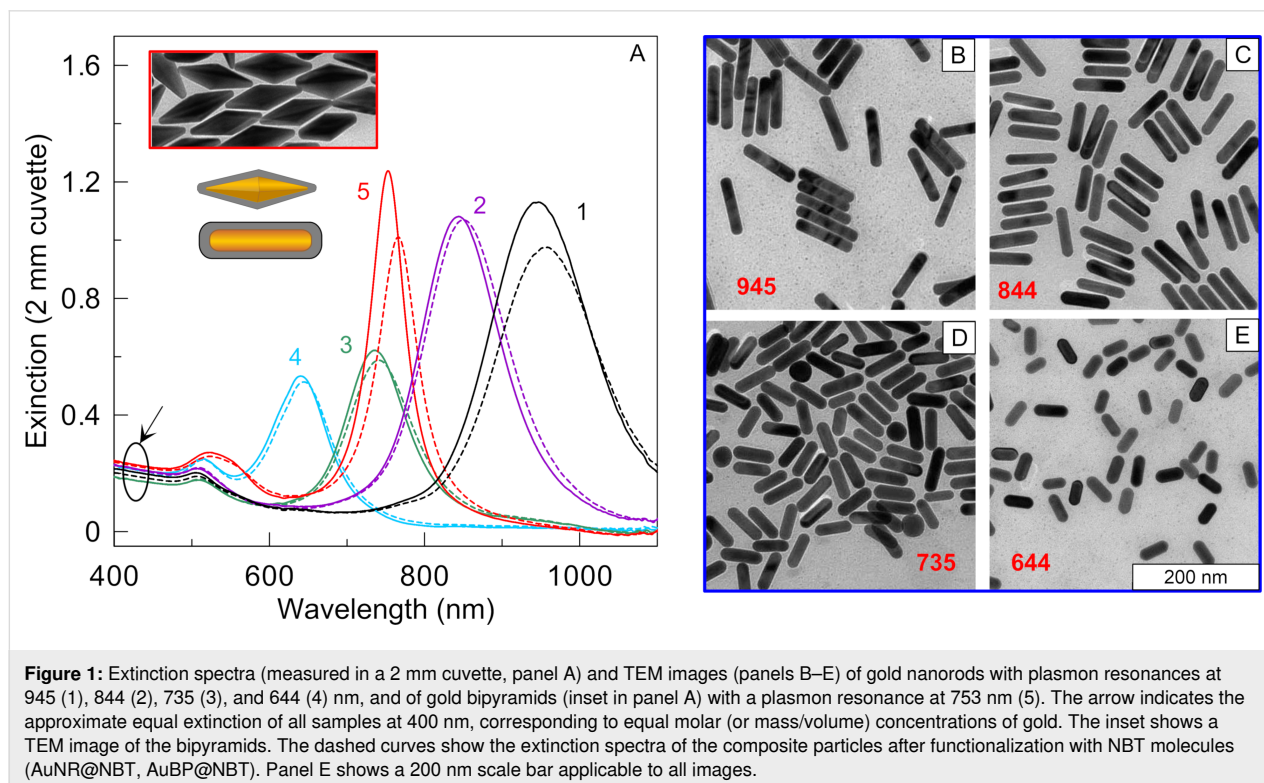


Table 1: Geometric and optical parameters of the synthesized nanoparticles (length, diameter, aspect ratio, surface area to volume ratio (S/V)), the normalized plasmon resonance bandwidth ($\text{FWHM}/\lambda_{\text{PR}}$), shift of the plasmon resonance after functionalization with NBT molecules (PR shift), measured SERS signal intensity (I_{1347}), and the value proportional to the intensity per molecule ($I_{1347}/(S/V)$). The ratio of the absorption efficiency factor to the radius of the equivalent volume sphere ($Q_{\text{abs}}/R_{\text{ev}}$) determines the photothermal efficiency of the particles at the plasmon resonance wavelength. The average length and aspect ratio were determined from TEM data (top numbers) and from the plasmon resonance wavelength, assuming a constant average diameter of 19 nm (bottom numbers). The aspect ratios in parentheses are calculated according to [59].

Sample	L_{TEM} L_{PR} (nm) ^a	d_{TEM} (nm) ^a	AR_{TEM} AR_{PR} (a.u.) ^a	S/V (nm ⁻¹)	$\text{FWHM}/\lambda_{\text{PR}}$ (a.u.) ^b	PR shift (nm) ^b	I_{1347} (a.u.) ^b	$I_{1347}/(S/V) \times 10^{-3}$ (a.u.) ^b	$Q_{\text{abs}}/R_{\text{ev}}$ (nm ⁻¹)
NR-644	44 ± 5.8 44 ± 4.0	19.0 ± 1.2	2.30 ± 0.16 2.26 ± 0.20 (2.14)	0.246	0.14 ± 0.01	3 ± 0.5	415 ± 72	1.7 ± 0.30	0.337
NR-735	62.2 ± 6.1 60 ± 4.9	19.0 ± 1.4	3.27 ± 0.09 3.14 ± 0.13 (3.08)	0.234	0.13 ± 0.01	4 ± 0.5	570 ± 108	2.4 ± 0.45	0.547
NR-844	82.3 ± 7.0 78 ± 5.2	18.9 ± 1.4	4.35 ± 0.07 4.10 ± 0.2 (4.21)	0.229	0.14 ± 0.01	6 ± 0.7	526 ± 85	2.3 ± 0.37	0.683
NR-945	90.2 ± 8.7 94 ± 6.1	18.7 ± 1.4	4.85 ± 0.13 4.97 ± 0.12 (5.25)	0.229	0.16 ± 0.02	7 ± 0.7	265 ± 59	1.2 ± 0.27	0.778
BP-753	75 ± 2.5	26.3 ± 1.2	2.85 ± 0.04	0.222	0.066 ± 0.008	13 ± 1.2	2150 ± 234	9.7 ± 0.90	0.996

^aSD values from statistical analysis of 200 particles; ^bSD values from three independent measurements.

covered by a typical 3 nm CTAB layer [50]. The corresponding length L_{P} was obtained by multiplying AR_{PR} by the average rod diameter of 19 nm. There is a close agreement between the nanorods aspect ratios derived from TEM images and from PR wavelength through Equation 1. Recently, Engel et al. [59] reported a similar linear relation between the PR wavelength and the nanorods' aspect ratio $\lambda_{\text{PR}}(\text{nm}) = 97 \times \text{AR} + 436$. The corresponding AR values are indicated in parentheses of the fourth column. We note that Equation 1 agrees somewhat better with TEM data.

The average plasmon resonance bandwidth of the nanorods, expressed in terms of the standard parameter full width at half maximum (FWHM), remains approximately constant (around 0.14) for nanorods samples. When comparing with the extinc-

tion spectrum of bipyramids (curve 5 in Figure 1A), the remarkably high quality of their spectrum, with an FWHM parameter of 0.066, half that of all nanorods values, immediately stands out. This observation agrees with a small SD value for the aspect ratio length/width = 2.85 ± 0.04 derived from TEM measurements.

Further confirmation of the high quality of the bipyramids' extinction spectra comes from comparing experimental and theoretical extinction spectra (Figure 2). For randomly oriented gold nanorods, extinction spectra were calculated using the T-matrix method. For the bipyramids, the longitudinal excitation spectrum is presented, calculated using MEM (practically coinciding with the COMSOL results). From Figure 2, it follows that the experimental nanorod spectra are significantly

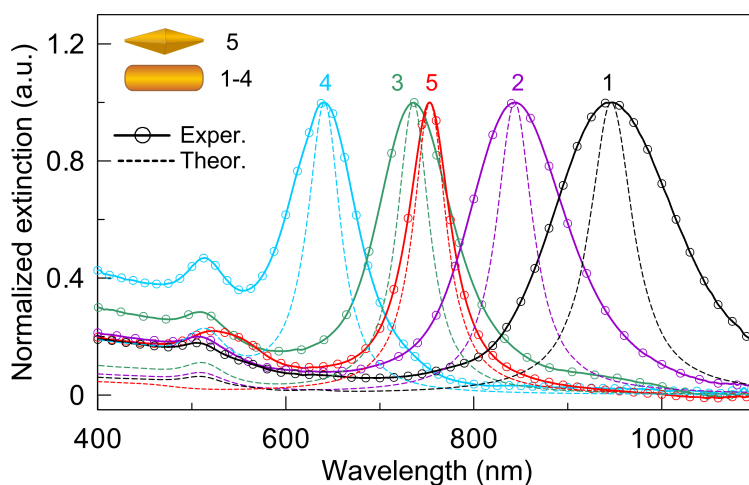


Figure 2: Experimental and theoretical (dashed lines) normalized extinction spectra for the five samples listed in Table 1. The theoretical particle aspect ratios AR_{P} were used as fitting parameters to match the calculated and measured plasmon resonance wavelength.

broadener than the theoretical ones, as the latter do not account for particle size (aspect ratio) distribution. Quantitatively, this spectral broadening, expressed as the ratio of the experimental to the theoretical FWHM value, is 2.5 for the nanorods. In contrast, for the bipyramids, the same ratio is half of that, approximately 1.25, even though the theoretical model also did not account for particle size distribution.

After functionalization with the Raman reporter molecule NBT, the plasmon resonance maximum of all particles decreases in magnitude and shifts, as is typical, to the red region. It is clear that these changes are most pronounced in the bipyramids compared to the nanorods. Quantitatively, the plasmon resonance shift after functionalization for bipyramids is 2–4 times greater than that for nanorods. This is related to the higher analytical sensitivity of the bipyramids' plasmon resonance to the dielectric environment, which will be discussed below.

In addition to the usual geometric particle parameters L and AR , Table 1 also provides the average surface area-to-volume ratio (S/V). For nanorods with hemispherical ends, $S/V = 12AR/[d(3AR - 1)]$, and for bipyramids modeled as inscribed bicones, $S/V = 6\sqrt{AR^2 + 1}/(d \cdot AR)$. For rounded bicones, the numerical S/V value is slightly less than the analytical one. As shown in Table 1, this parameter varies within a narrow range of 0.222–0.246. In the next section, we will use this parameter to estimate the SERS signal intensity per NBT reporter molecule.

The last column of Table 1 presents the theoretical value of $Q_{\text{abs}}/R_{\text{ev}}$, which is the ratio of the absorption efficiency factor Q_{abs} to the radius of the equivalent volume sphere R_{ev} . This parameter determines the photothermal efficiency of plasmonic nanoparticles per unit volume at a constant concentration c ($\text{g} \cdot \text{mL}^{-1}$), or, in other words, the photothermal efficiency per unit mass of nanoparticles. Indeed, the absorbed power per unit volume equals the product of the incident intensity I ($\text{W} \cdot \text{cm}^{-2}$) and the numerical concentration N (cm^{-3}) multiplied by the absorption cross section $C_{\text{abs}} (\text{cm}^2) = \pi R_{\text{ev}}^2 Q_{\text{abs}}$:

$$\begin{aligned} P_{\text{abs}} \left(\text{W} / \text{cm}^3 \right) &= I \times N \times C_{\text{abs}} = I \times \frac{c}{\frac{4}{3} \pi R_{\text{ev}}^3 \rho} \times \pi R_{\text{ev}}^2 Q_{\text{abs}} \\ &= I \frac{3c}{4\rho} \times \frac{Q_{\text{abs}}}{R_{\text{ev}}}, \end{aligned} \quad (2)$$

where ρ is the metal density. From Equation 2, it follows that the absorbed power per unit volume is proportional to the ratio $Q_{\text{abs}}/R_{\text{ev}}$ at a constant mass/volume concentration of particles c ($\text{g} \cdot \text{mL}^{-1}$).

2 Comparison of SERS spectra for nanorods and nanobipyramids

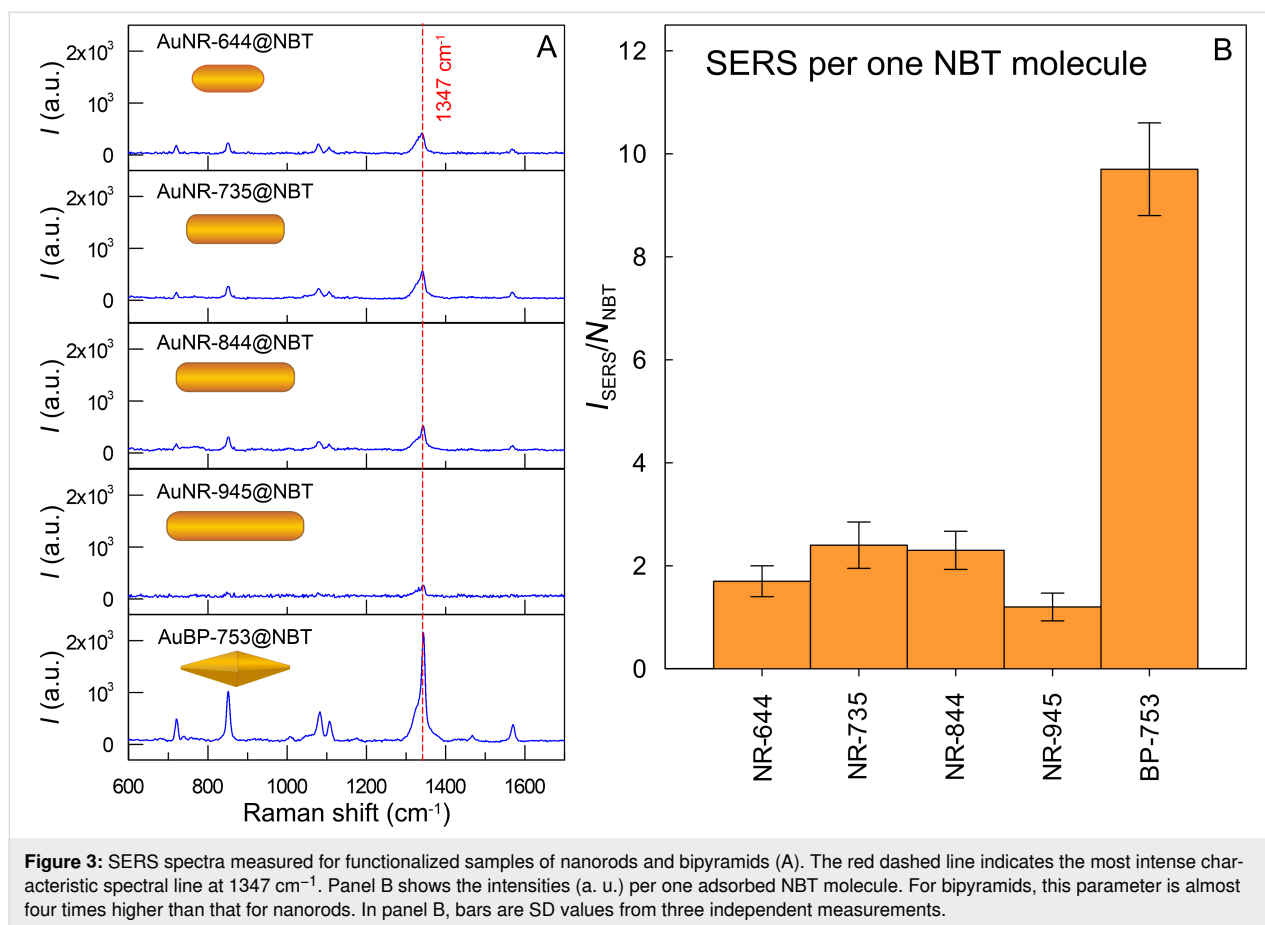
Figure 3 shows the SERS spectra of functionalized gold nanorods and bipyramids. The most intense line at 1347 cm^{-1} was used as a reference. Comparison of the spectra reveals a more intense peak for the functionalized bipyramids. Even for NR-735 nanorods, which are close to resonant excitation, the peak intensity is almost four times lower, even though the extinction peaks are comparable in magnitude and the mass/volume concentration of nanoparticles was the same.

To give some insight into the data of Figure 3B, we use a simple estimate based on experimental extinction spectra, SERS spectra, and particle geometric parameters from TEM data. As mentioned earlier, the extinction and SERS spectra were recorded at approximately the same molar concentration of 0.1 mM or, correspondingly, at a mass/volume concentration of gold $19.7 \times 10^{-6} \text{ g} \cdot \text{mL}^{-1}$. For numerical estimates, we will assume that the number of molecules per particle is proportional to their surface area. Since functionalization was carried out with an excess of NBT, this assumption implies monolayer adsorption of SERS-active molecules, and the number of molecules on a single particle is proportional to the ratio of the particle's surface area to the effective footprint area of one molecule $N_1 = S/S_1$. The total number of particles in 1 mL of colloid equals the ratio of the mass/volume concentration of gold to the mass of a single particle $N = m/m_1 = c/m_1$. The gold concentration was constant in all experiments ($0.1 \text{ mM} = 19.7 \times 10^{-6} \text{ g} \cdot \text{mL}^{-1}$), therefore, the number of SERS-active molecules in 1 mL of colloid is

$$N_{\text{R}} = N_1 \times N = \frac{S}{S_1} \frac{c}{m_1} = \frac{S}{S_1} \frac{c}{\rho V} = \left(\frac{c}{S_1 \rho} \right) \frac{S}{V} = K \frac{S}{V}, \quad (3)$$

where, for our comparative estimates, the K constant can be set to 1. Thus, under our experimental conditions, the number of molecules per particle is proportional to the ratio of its surface area to its volume.

It should be emphasized that our final Equation 3 involves only one experimental parameter, the particle surface-to-volume ratio (S/V), whereas other adsorption parameters and the number concentration of particles are not considered. This simple and important result is based on several simplifying assumptions. Our main assumption is that the number of SERS-active NBT molecules N_{R} is roughly proportional to the particle surface area. This assumption does not necessarily imply a monolayer or any specific arrangement of the adsorbed layer. The only requirement is that the total number of adsorbed SERS-active mole-



cles be proportional to the particle surface area. In other words, the number of SERS-active molecules should be distinguished from the total number of adsorbed molecules. Indeed, as the local field decreases strongly with increasing distance from the particle surface and the SERS intensity is roughly proportional to the fourth power of the exciting field [60], only surface-adsorbed molecules make the main contribution to the detected SERS signal. This means that other adsorption parameters such as accessible surface area and adsorption affinity probably play a minor role. Moreover, an accurate experimental assessment of these parameters is not straightforward and would require a separate investigation. To the best of our knowledge, no studies of this type have been reported for gold nanobipyramids. In summary, we believe that our Equation 3 provides a reasonable estimate of the number of SERS-active molecules in comparative experiments with a fixed mass/volume concentration of particles having different sizes, shapes, and morphologies.

Figure 3B presents the normalized SERS intensity $I/(S/V)$ values, which, as explained above, are proportional to the average SERS intensity from a single adsorbed NBT molecule. In this sense, this parameter provides a comprehensive, averaged

characteristic of the efficiency of a plasmonic nanoparticle as an SERS platform. As shown in the figure, the SERS efficiency of bipyramids is approximately four times higher than that of nanorods.

3 Analytical sensitivity of plasmon resonance to the dielectric environment

The sensitivity of PR to the dielectric environment is utilized for various analytical tasks aimed at determining target substances based on PR shifts [61]. Sensors based on this principle are somewhat similar to total internal reflection sensors, although the physical mechanism is entirely different in this case. In recent years, the most popular trend has been the development of fiber-optic sensors with a plasmonic nanostructure at the tip [29]. Theoretically, there are two approaches to estimating the analytical sensitivity of a PR sensor. In the most straightforward approach, the plasmonic response (e.g., the PR wavelength) is typically analyzed in a homogeneous dielectric medium with a varying refractive index (usually from 1.334 (water) to 1.6), and the sensitivity of different platforms is compared in terms of the derivative of the response with respect to the refractive index. For example, in the case of the PR wavelength, parameters such as $S_{\text{PR}} = \Delta\lambda_{\text{PR}}/\Delta n_{\text{m}}$ [62,63] are com-

pared. Within the dipole approximation, the polarizability of a particle of any shape and structure can be written as [52,64]:

$$\alpha = \frac{3V}{4\pi} \frac{\epsilon_{av} - \epsilon_m}{\epsilon_{av} + \varphi\epsilon_m} = a_0^3 \frac{\epsilon_{av} - \epsilon_m}{\epsilon_{av} + \varphi\epsilon_m}, \quad (4)$$

where ϵ_{av} is the average dielectric permittivity of particles with a complex structure, calculated using the DEM. The function φ also depends on the shape and structure, and ϵ_m is the dielectric permittivity of a homogeneous medium. Here are three simple examples: (1) a metal sphere with the dielectric permittivity $\epsilon_{metal} = \epsilon(\omega)$ in a homogeneous medium with dielectric permittivity $\epsilon_m(\omega)$: $\epsilon_{av} = \epsilon(\omega)$, $\varphi = 2$; (2) a metal ellipsoid with the dielectric permittivity $\epsilon_{metal} = \epsilon(\omega)$ in a homogeneous medium $\epsilon_m(\omega)$: $\epsilon_{av} = \epsilon(\omega)$, $\varphi = L_{a,b,c}^{-1} - 1$, where $L_{a,b,c}$ are the geometrical depolarization factors along the axes a , b , and c , $\sum_{i=a,b,c} L_i = 1$. For a sphere $L_i = L = 1/3$; and (3) a two-layered sphere with a metal core $\epsilon_{metal} = \epsilon_1(\omega)$ covered by a dielectric shell of thickness s with a dielectric permittivity $\epsilon_s = \epsilon_2(\omega)$ in a homogeneous medium $\epsilon_m(\omega)$:

$$\epsilon_{av} = \epsilon_2 \frac{1 + 2f_{12}\alpha_{12}}{1 - f_{12}\alpha_{12}}, \quad \varphi = 2, \quad (5)$$

where $\alpha_{12} = (\epsilon_1 - \epsilon_2)/(\epsilon_1 + 2\epsilon_2)$, $f_{12} = a_1^3/a_2^3 = a_1^3/(a_1 + s)^3$. It follows from Equation 4 that the PR resonance condition reads:

$$\text{Re}(\epsilon_{av}) = -\varphi\epsilon_m. \quad (6)$$

In the Drude approximation, the metal dielectric function is given by

$$\epsilon(\omega) = \epsilon_{ib}(\omega) - \frac{\omega_p^2}{\omega(\omega + i\gamma_b)}, \quad (7)$$

where $\epsilon_{ib}(\omega)$ accounts for the contribution of interband transitions, ω_p is the bulk metal plasma frequency (for gold, $\hbar\omega_p \approx 9$ eV), and the damping constant γ_b corresponds to a bulk sample. It does not account for additional damping mechanisms [64]. Combining Equation 6 and Equation 7, we find the plasmon resonance wavelength [52,64]:

$$\lambda_{PR} = \lambda_p [\epsilon_{ib} + \epsilon_m\varphi]^{1/2}. \quad (8)$$

Using Equation 8, one can obtain a theoretical estimate of the sensitivity of PR to changes in the refractive index of the external medium in the form of universal relations:

$$S_{PR} = \frac{\Delta\lambda_{PR}}{\Delta n_m} = \varphi \frac{\lambda_p n_m}{\sqrt{\epsilon_{ib} + \varphi n_m^2}}, \quad (9)$$

$$\frac{\Delta\lambda_{PR}}{\lambda_{PR}} = \frac{\Delta n_m}{n_m} \left(1 - \frac{\lambda_p^2}{\lambda_{PR}^2} \epsilon_{ib} \right), \quad (10)$$

where $\lambda_p = 2\pi\omega_p/c$ is the plasma wavelength (for gold, $\lambda_p \approx 130$ nm), and c is the velocity of light in vacuum.

Equation 9 explains the theoretically obtained [62,63] and experimentally observed linear dependencies $\lambda_{PR} = f(n_m)$ with a constant coefficient S_{PR} , since n_m usually changes within a narrow range of 1.3 to 1.5 (for water–glycerol mixtures). In this case, according to Equation 10, the relative change $\Delta\lambda_{PR}/\lambda_{PR}$ turns out to be proportional to the relative change $\lambda_{PR} = f(n_m)$ with an accuracy of approximately a constant coefficient (for gold in water, it is approximately 0.5 [65]). Equations 9 and 10 also explain that the structure and shape of a particle decisively influence the angular slope of the dependence $\lambda_{PR} = f(n_m)$ through the parameter φ in Equation 9.

For the theoretical evaluation of the analytical sensitivity of a plasmonic sensor, one can also calculate the change in the plasmonic response resulting from the formation of a dielectric layer on the particle surface, for example, due to antigen–antibody interactions [66]. This formulation of the problem seems more justified from the perspective of modeling real experimental situations, where the process of analyte molecule adsorption on the surface of a plasmonic nanosensor is detected by the shift in plasmon resonance. This modeling process itself can be carried out in two variants, namely, (1) analysis of the PR shift as a function of shell thickness with a constant refractive index and (2) analysis of the PR shift as a function of the shell's refractive index at its constant nanometer-scale thickness. The real situation is most likely a combination of these two variants since, as the thickness of the adsorbed biopolymer layer increases, the volume fraction of the aqueous buffer (or other solvent) will decrease. Below, we present comparative data for nanorods and BPs in a homogeneous, infinite medium and in a shell of variable thickness with a constant refractive index.

Figure 4 shows the dependence of the plasmon resonance wavelength shift $\Delta\lambda_{PR}$ on the refractive index of the external medium n_m for gold BPs and nanorods with a diameter of 15 nm and an aspect ratio of 3. For the bipyramids, calculations were performed using the 3D FEM COMSOL method under longitudinal excitation of the particles by an electric field. For the

nanorods, the calculation was performed on randomly oriented nanoparticles using the T-matrix method. We also performed an approximate averaging over the orientations of the bipyramids and confirmed that the result differs only slightly from that presented in Figure 4.

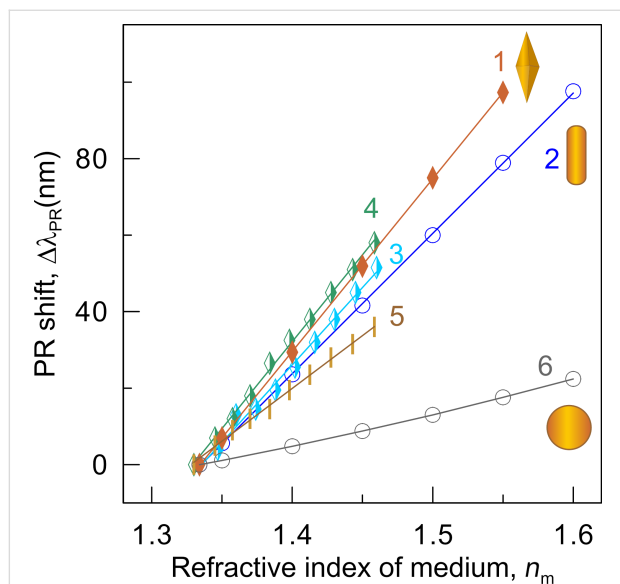


Figure 4: Dependence of the plasmon resonance wavelength shift $\Delta\lambda_{PR}$ on the refractive index of the external medium (n_m). Calculation for gold BPs (1, COMSOL) and nanorods (2, T-matrix method) with a diameter of 15 nm and an aspect ratio of 3. Experimental points 3 and 4 for gold BPs are reproduced from works by Chen et al. [28] (AR = 3.8) and by Fang et al. [30] (AR = 3.3), respectively. Experimental points (5) for gold nanorods with $AR_{TEM} \approx 4.8$ ($AR_{PR} = 4.2$) are reproduced from Fang et al. [30]. For comparison, the linear dependence (6) according to Mie theory for spheres with an equivalent diameter of 24 nm is also shown.

From Figure 4, it follows that for nanoparticles with identical geometric parameters, the analytical sensitivity of bipyramids, measured by the plasmon resonance shift, is higher than that of nanorods. It should be noted that both the FEM (COMSOL) and T-matrix methods yield identical results for axially symmetric particles such as nanorods (see, e.g., Figure S6 in Supporting Information File 1). Therefore, the difference in the slopes of lines 1 and 2 in Figure 3 arises from the distinct plasmonic sensitivities of the two particle types, rather than from any computational artifacts.

Additional confirmation of the strong influence of the particle is provided by line (6), obtained from Mie theory for spheres of equivalent diameter. For qualitative comparison with experimental data, we have reproduced in Figure 4 data from the work of Chen et al. [28] for bipyramids with an aspect ratio of AR = 3.8 and from Fang et al. [30] for bipyramids with an aspect ratio of AR = 3.3. The last data are in close agreement with our simulations. It should also be noted that the particle

sizes and tip rounding radii in the Chen et al. [28] samples were significantly larger than those used in our calculation for dependence (1); therefore, this comparison should be considered qualitative. This is precisely why the experimental line (4) for bipyramids with an aspect ratio of 3.8 lies below the theoretical line for an aspect ratio of 3 and the experimental line (4) by Fang et al. [30] for an aspect ratio of 3.3. The experimental data [30] for nanorods confirm a lower angular slope than that for BPs. In general, we conclude that the theoretical estimates of the angular slope S_{PR} for BPs and nanorods are in qualitative agreement with the experimental data.

Now, let us discuss the data for the case where the analyte forms a shell on the surface of a plasmonic nanoparticle. Figure 5A shows the dependence of the plasmon resonance wavelength shift $\Delta\lambda_{PR}$ on the shell thickness s (curves 1, 2) or its volume fraction $g = V_s/(V_s + V_{Au})$ (curves 3, 4) for bipyramids, modeled as an inscribed bicone [38] with an aspect ratio of 3. For comparison, two particle diameters of 15 and 30 nm were chosen, which are close to the experimental conditions [38], with a typical refractive index of 1.5 for many biopolymers or silicon dioxide [67].

From plots 1 and 2, it can be seen that the main sharp plasmon resonance shift occurs when the shell thickness changes from 0 to 5–10 nm, at which point the shell volume fraction approaches 75–80%. Such a sharp dependence of PR shift on the shell thickness is closely related to the strong localization of plasmonic field as explained below in Section 4. Figure 5B shows the dependencies of the plasmon resonance wavelength shift on the refractive index of the shell for two particle diameters, 15 and 30 nm, at shell thicknesses of 5 and 10 nm, which also differ by a factor of two. As shown, lines 1 and 2 are practically indistinguishable, which is explained by the approximately equal shell volume fractions in both nanocomposites. As mentioned above, increasing the shell thickness to that of an infinite homogeneous dielectric medium somewhat increases the angular slope of dependence (3). Still, it does not change the fundamental essence of the matter. Thus, the formation of dielectric analyte shells with a thickness of about 10 nm is essentially equivalent to the plasmon resonance shift when placed in an infinite analyte medium.

4 Physical mechanisms underlying the superior SERS and analytical responses of bipyramids over nanorods

The data presented in Sections 2 and 3, together with available literature data, indicate the key influence of particle shape and morphology (all other things being equal) on their SERS and analytical responses. In particular, the well-known decrease in the SERS signal from spherical particles of equal volume com-

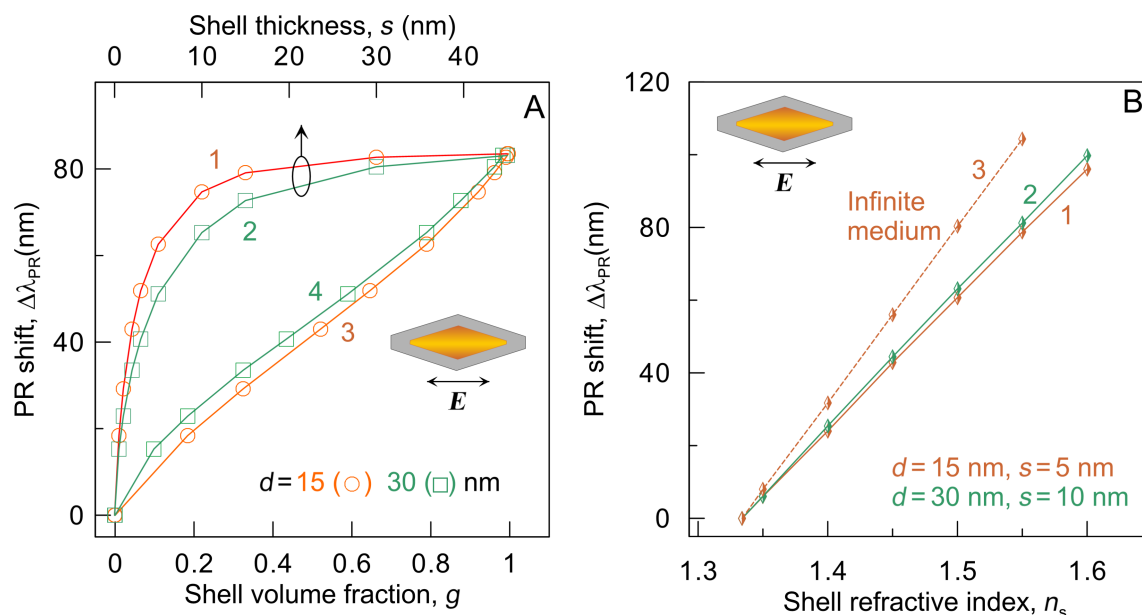


Figure 5: (A) Dependence of the plasmon resonance wavelength shift $\Delta\lambda_{PR}$ on shell thickness (1, 2) or its volume fraction (3, 4). Calculation based on the analytical MEM theory for composite nanoparticles consisting of gold bicones with diameters of 15 nm (1, 3) and 30 nm (2, 4), an aspect ratio $AR = 3$, and a dielectric shell thickness s ranging from zero to 45 nm. The refractive index of the shell is 1.5. (B) Dependence of the plasmon resonance wavelength shift $\Delta\lambda_{PR}$ of gold bicones on the refractive index of a dielectric shell with thicknesses of 5 nm (1) and 10 nm (2). For comparison, the dashed line (3) shows the dependence of the plasmon shift (1) for a particle in an infinite homogeneous medium. Particle diameters are 15 nm (1) and 30 nm (2), and the axial ratio is 3.

pared with nanorods [2], as well as the small plasmon resonance shift for equal-volume spheres (curve 6 in Figure 4), suggest that the underlying physical mechanism of the observed differences in SERS and analytical sensitivity is related to the features of particle shape and morphology, which lead to different local field distributions near the nanoparticle surface where the reporter molecules are located [2] (Figure 6).

To quantitatively assess the SERS efficiency of particles with different morphologies, theoretical estimates of the average fourth power of the local field amplitude near the particles $\langle |E(\omega)|^4 \rangle$ are often used, or a slightly more rigorous estimate $\langle |E(\omega_L)|^2 |E(\omega_R)|^2 \rangle$ that accounts for the difference between the laser, ω_L , and Raman, ω_R , frequencies [68]. Here, we define two average quantities

$$\langle |E(\lambda_{PR})| \rangle = \frac{1}{S} \int_S |E(\lambda_{PR})/E_0| dS, \quad (11)$$

$$EF = \langle |E(\lambda_{PR})|^4 \rangle = \frac{1}{S} \int_S |E(\lambda_{PR})/E_0|^4 dS, \quad (12)$$

where the integral is taken over the outer particle surface. The first integral provides a straightforward averaged estimate of the

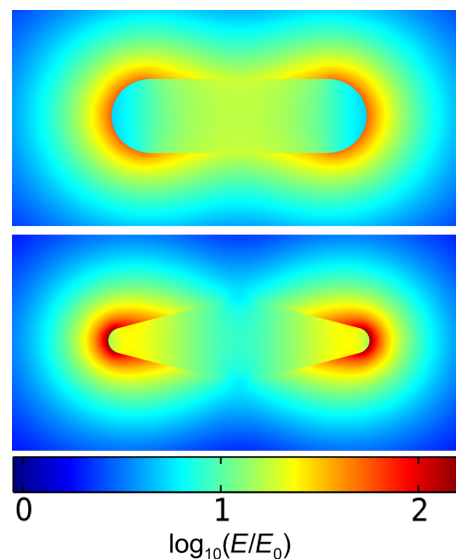


Figure 6: Calculated distribution of the logarithm of the field modulus ($\log_{10}|E|/|E_0|$) around the particle in water at the plasmon resonance wavelength for nanorods (top) and nano-bipyramids BP (bottom). The particle parameters correspond to the experimental ones (Table 1). The numbers on the color scale correspond to an order-of-magnitude change in the local field amplitude.

field on the surface near the particle, while the second integral gives an approximate average estimate of the plasmonic enhancement factor (EF) for the SERS signal. A more accurate

estimate of EF would also need to include the surface density of SERS-active molecules (with normalization $(1/S)\int_S n(S)dS = 1$) and the difference between the Raman and laser frequencies:

$$EF = \frac{1}{S} \int_S \left(|E(\omega_L)|^2 |E(\omega_R)|^2 \right) n(S) dS. \quad (13)$$

A detailed analysis of the theoretical SERS enhancement factor is given in [69]. Table 2 summarizes the average local field parameters for nanorods and bipyramids with sizes and shapes close to our experimental samples particles NR-735 and BP-753. The calculation details are given in Supporting Information File 1. Note the excellent agreement between calculated and measured PR wavelength.

Table 2: The local field parameters of nanorods NR-735 and bipyramids BP-753 at longitudinal excitation. Calculations by COMSOL for the normalized incident field at theoretical plasmon resonance wavelengths λ_{PR} . The angle bracket designate the averaging over the particle surface.

Sample	λ_{PR} (nm)	S/V (nm ⁻¹)	$ E _{max}$	$\langle E \rangle_S$	$\langle E ^4 \rangle_S \times 10^{-6}$
NR-735	740	0.235	50.2	25.9	1.91
BP-753	756	0.222	157	31.4	19.8

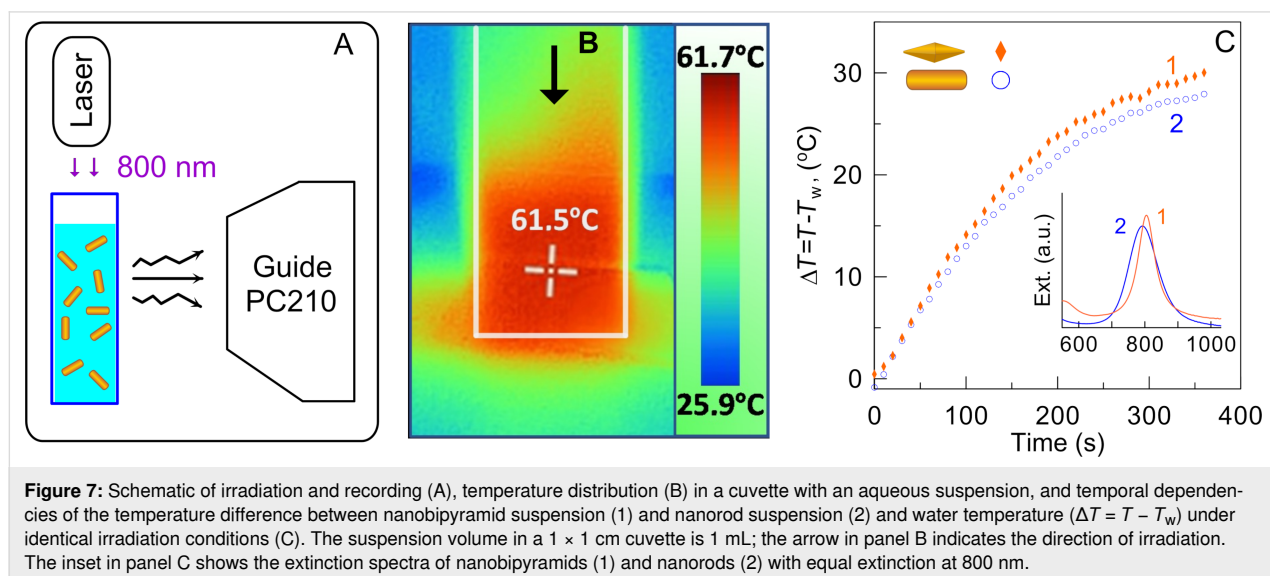
According to Table 2, the nanorods and bipyramid surfaces are similar, but the SERS enhancement factor of bipyramids is ten times higher. Experimentally (Figure 3), we obtained a much smaller enhancement ratio of about 4. As it has been explained in Section 2 and during derivation of Equation 3, this difference originates from a simplified estimation by Equation 13, which does not take into account many factors including the

adsorption accessibility, affinity, and resulting surface density distributions of Raman NBT molecules. Obviously, the purely electromagnetic EF (Equation 12) overestimates the expected experimental parameter. Nevertheless, the simplified electromagnetic calculations by Equation 12 confirm the superior SERS efficiency of bipyramids compared to that of nanorods.

From Figure 4A, it follows that for nanoparticles with identical geometric parameters, the analytical sensitivity of bipyramids, measured by the plasmon resonance shift, is higher than that of nanorods. From a physical standpoint, this difference can be explained by the strong field localization near the vertices of the bipyramid (Figure 4B) and the resulting more substantial change in the dipole moment upon variation of the external refractive index compared to nanorods (Figure 4C). Qualitatively, the modulus of the local field near the bipyramid vertices is approximately an order of magnitude greater than the field near the ends of the nanorods. In accordance with the field distribution shown in Figure 6, this shell region precisely falls within the “hottest” area of the local electric field. Further increase in shell thickness has a smaller effect on the plasmon shift, which essentially becomes close to the PR shift in an infinite dielectric medium with the shell’s refractive index (cf. dependencies 1 and 3 in Figure 5B). Interestingly, the average values of the local field modulus at the particle surface are similar for both particle types. This implies that the plasmonic shift induced by thin shell formation is not linearly dependent on the near field.

5 Comparison of thermoplasmonic properties for gold nanorods and nanobipyramids

To compare the efficiency of converting laser radiation into heat, an experiment was conducted (Figure 7A) comparing the



heating of gold BPs and nanorods with equal extinction at 800 nm, a wavelength close to the plasmon resonance wavelength of both nanoparticle types. The concentration of nanorods was 0.062 mM ($12 \mu\text{g}\cdot\text{mL}^{-1}$), determined by absorbance at 400 nm. A 2 W laser with a wavelength of 800 nm was used for irradiation; the intensity of the expanded beam at the cuvette entrance was approximately $290 \text{ mW}\cdot\text{cm}^{-2}$. Irradiation was performed from above into a $1 \times 1 \text{ cm}$ cuvette containing 1 mL of suspension, so the optical path length from the top to the bottom of the cuvette was 1 cm. The temperature distribution (Figure 7B) was recorded from the side using a thermal imager (Figure 7A). Irradiation and temperature measurement lasted about 6 min. Figure 7C shows the dependencies of the temperature increment (relative to the water temperature under the same irradiation conditions) on the irradiation time. It can be seen that after 5–6 min of irradiation, the temperature increment reaches an approximately steady-state value of around 30°C . Although the heating kinetic curves are generally similar, we observe slightly higher efficiency for gold bipyramids, in agreement with the theoretical estimate of the $Q_{\text{abs}}/R_{\text{ev}}$ ratio in the last column of Table 1. Thus, theoretical and experimental data show that, despite the smaller metal volume per particle, the photothermal efficiency of bipyramids is of the same order (or even slightly higher) as that of widely used nanorods.

6 Comparison of the photothermal killing of bacteria

For plasmonic photothermal therapy experiments with bacteria and nanoparticles, cells were mixed with nanoparticles in a 1:1 ratio, incubated for 15 min, and irradiated with a laser for 10 min. Four controls were used in the experiments, namely, (1) native bacteria without any treatment (B Cont), (2) bacteria without particles irradiated with the laser (B-Part+L), (3) bacteria with nanorods without laser irradiation (B+NR-L), and (4) bacteria with nanobipyramids without laser irradiation (B+BP-L). In the two positive experiments, that is, (5) (B+NR+L) and (6) (B+BP+L), bacteria were incubated with nanoparticles for 15 min, then irradiated with a laser intensity of $290 \text{ mW}\cdot\text{cm}^{-2}$ for 10 min. All experiments were conducted using a 96-well plate. Cell viability was assessed using the standard Alamar Blue test [57]. For this, $5 \mu\text{L}$ of resazurin was added to each $200 \mu\text{L}$ sample, and the mixture was incubated in a thermal chamber at 37°C for 1 h. The results are presented in Figure 8 and Table 3. From the presented data, it can be seen that both types of nanoparticles are effective thermosensitizers, causing complete bacterial death after 10 min of irradiation at a very low intensity of about $0.3 \text{ W}\cdot\text{cm}^{-2}$, which is an order of magnitude lower than the intensities typically used for photothermal therapy ($2 \text{ W}\cdot\text{cm}^{-2}$). A small 10% reduction in bacterial viability without laser irradiation could be explained by the possible contribution of residual CTAB/CTAC toxicity.

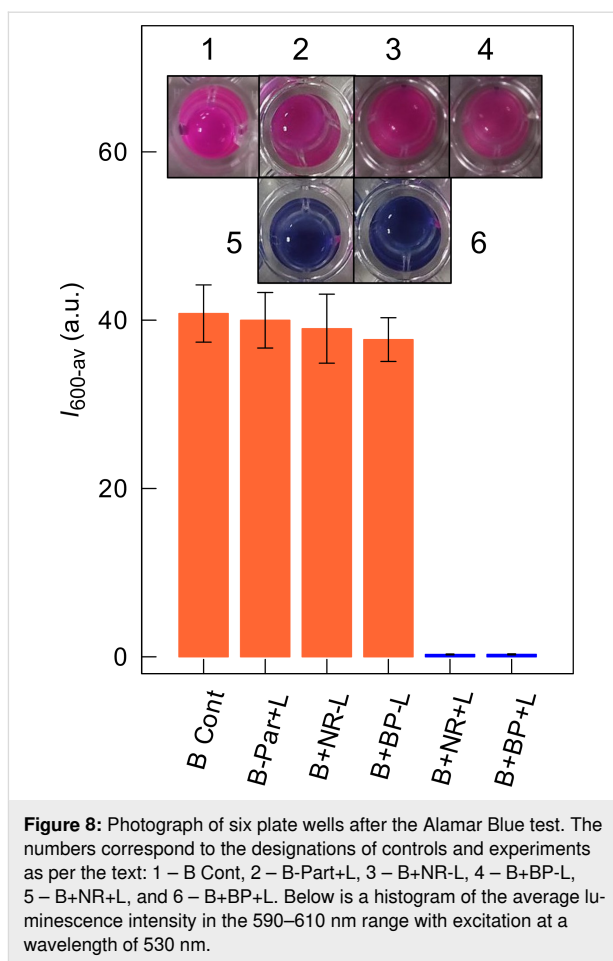


Figure 8: Photograph of six plate wells after the Alamar Blue test. The numbers correspond to the designations of controls and experiments as per the text: 1 – B Cont, 2 – B-Part+L, 3 – B+NR-L, 4 – B+BP-L, 5 – B+NR+L, and 6 – B+BP+L. Below is a histogram of the average luminescence intensity in the 590–610 nm range with excitation at a wavelength of 530 nm.

Table 3: Fluorescence intensities of the samples at 600 nm and assessment of cell viability.

Sample	Intensity I_{600} (a.u.)	Cell viability, %
1 – B Control	44.6	100
2 – B-Part+L	43.5	97.5
3 – B+NR-L	40.5	90.7
4 – B+BP-L	40.1	90
5 – B+NR+L	0.30	0.7
6 – B+BP+L	0.26	0.6

In conclusion to this section, it should be noted that, in addition to the thermal effect, nanoparticles can be potentially toxic [70] at high concentrations. Toxicity of gold nanoparticles was observed only in *in vivo* experiments at small sizes (2–5 nm), high concentrations, and in cases where cellular uptake was observed [71]. Toxicity of gold nanorods is also mainly manifested in *in vivo* experiments [72], and for nanorods, their toxicity is determined primarily not by size or shape, but by surface chemistry. First and foremost, the potential source of toxicity is the toxic surfactant stabilizer CTAB. In experiments with bacterial cells, cellular uptake of particles is not observed due to the

dense cell wall and the presence of an extracellular polymeric component. This is also consistent with our experiments, because in the control with the addition of nanoparticles but without laser heating, no reduction in cell viability was observed. The main effect is cellular adhesion of gold nanoparticles on the bacterial surface [73] rather than bacterial uptake. Therefore, the primary damaging mechanism in experiments with bacteria is precisely local heating of cells adsorbed on their surface.

Conclusion

In this work, we have presented experimental and theoretical data comparing several important plasmonic parameters of the most popular anisotropic particles, gold nanorods, and the less familiar, yet in our view, quite promising plasmonic gold bipyramid nanoparticles. Our goal was to draw researchers' attention to this new class of anisotropic plasmonic nanoparticles with a tunable plasmon resonance spanning 600–1200 nm. Due to their strong shape anisotropy, bipyramids generate a strong local field near their sharp tips when excited at their plasmon resonance wavelength. Such localization has important implications for potential biomedical applications. First, Raman reporter molecules located near these local-field hot spots emit a stronger SERS signal than conventional nanorods under the same conditions. Second, bipyramids exhibit greater analytical sensitivity to global or local dielectric environmental changes, leading to a larger plasmon resonance shift. Third, it can be expected that bipyramids will be promising nanoparticles as a plasmonic platform for bioimaging. Finally, thanks to their high absorption per unit mass of metal, bipyramids show promise as photothermal sensitizers that efficiently generate heat upon laser irradiation.

Supporting Information

Section S1. Additional TEM images and particle statistics; Section S2. Comparison of extinction spectra calculated by COMSOL and the T-matrix method; Section S3. Calculations of the average near field parameters.

Supporting Information File 1

Additional experimental and calculation data.

[<https://www.beilstein-journals.org/bjnano/content/supplementary/2190-4286-17-80-S1.pdf>]

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Compliance with Ethical Standards

This work does not involve studies with human participants or animals.

Conflict of Interest

The authors of this work declare that they have no conflicts of interest.

Declaration of Generative AI and AI-Assisted Technologies

During the preparation of this work, the authors used DeepSeek v3 and Grammarly to check the English. After using these tools, the authors reviewed and edited the content as needed and take full responsibility for the content of the published article.

Author Contributions

Andrey M. Burov: investigation; methodology; visualization; writing – review & editing. Sergey V. Zarkov: investigation; software; visualization. Arina V. Drozd: investigation. Igor V. Borisov: investigation; methodology. Elena G. Zavyalova: funding acquisition; project administration. Nikolai G. Khlebtsov: conceptualization; data curation; formal analysis; methodology; project administration; software; supervision; visualization; writing – original draft; writing – review & editing.

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Data Availability Statement

All modeling and experimental data are available from the authors upon request.

Preprint

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References

1. Liz-Marzán, L. M.; Grzelczak, M. *Science* **2017**, *356*, 1120–1121. doi:10.1126/science.aam8774
2. Reguera, J.; Langer, J.; Jiménez de Aberasturi, D.; Liz-Marzán, L. M. *Chem. Soc. Rev.* **2017**, *46*, 3866–3885. doi:10.1039/c7cs00158d

3. Zhuo, X.; Kumar, V.; Chow, T. H.; Wang, J.; Liz-Marzán, L. M. Synthesis and Functionalization of Anisotropic Silver Nanoparticles. In *Synthesis and Functionalization of Anisotropic Silver Nanoparticles*; Wang, J., Ed.; World Scientific Series in Nanoscience and Nanotechnology, Vol. 22; World Scientific, 2022; pp 349–396. doi:10.1142/9789811235221_0008
4. Ortiz-Castillo, J. E.; Gallo-Villanueva, R. C.; Madou, M. J.; Perez-Gonzalez, V. H. *Coord. Chem. Rev.* **2020**, *425*, 213489. doi:10.1016/j.ccr.2020.213489
5. Nguyen, Q. N.; Wang, C.; Shang, Y.; Janssen, A.; Xia, Y. *Chem. Rev.* **2023**, *123*, 3693–3760. doi:10.1021/acs.chemrev.2c00468
6. Khlebtsov, N. G.; Dykman, L. A.; Khlebtsov, B. N. *Russ. Chem. Rev.* **2022**, *91*, RCR5058. doi:10.57634/rcr5058
7. Jain, P. K.; Lee, K. S.; El-Sayed, I. H.; El-Sayed, M. A. *J. Phys. Chem. B* **2006**, *110*, 7238–7248. doi:10.1021/jp057170o
8. Lu, Z.; Ji, J.; Ye, H.; Zhang, H.; Zhang, S.; Xu, H. *Nat. Commun.* **2024**, *15*, 8803. doi:10.1038/s41467-024-53210-8
9. Liz-Marzán, L., Ed. *Colloidal Synthesis of Plasmonic Nanometals*; Jenny Stanford Publishing: New York City, NY, U.S.A., 2020. doi:10.1201/9780429295188
10. Xia, Y.; Li, W.; Cobley, C. M.; Chen, J.; Xia, X.; Zhang, Q.; Yang, M.; Cho, E. C.; Brown, P. K. *Acc. Chem. Res.* **2011**, *44*, 914–924. doi:10.1021/ar200061q
11. Panfilova, E. V.; Khlebtsov, B. N.; Khlebtsov, N. G. *Colloid J.* **2013**, *75*, 333–338. doi:10.1134/s1061933x13030149
12. Zhang, F.; Xu, L.; Wang, Y.; Wang, P. *Mater. Today Bio* **2025**, *33*, 102022. doi:10.1016/j.mtbio.2025.102022
13. Wang, J.; Fang, W.; Liu, H. *ChemPlusChem* **2023**, *88*, e202200464. doi:10.1002/cplu.202200464
14. Scarabelli, L.; Sun, M.; Zhuo, X.; Yoo, S.; Millstone, J. E.; Jones, M. R.; Liz-Marzán, L. M. *Chem. Rev.* **2023**, *123*, 3493–3542. doi:10.1021/acs.chemrev.3c00033
15. Jin, X.; Khlebtsov, B. N.; Khanadeev, V. A.; Khlebtsov, N. G.; Ye, J. *ACS Appl. Mater. Interfaces* **2017**, *9*, 30387–30397. doi:10.1021/acsami.7b08733
16. Khlebtsov, N. G.; Lin, L.; Khlebtsov, B. N.; Ye, J. *Theranostics* **2020**, *10*, 2067–2094. doi:10.7150/thno.39968
17. Zheng, J.; Cheng, X.; Zhang, H.; Bai, X.; Ai, R.; Shao, L.; Wang, J. *Chem. Rev.* **2021**, *121*, 13342–13453. doi:10.1021/acs.chemrev.1c00422
18. Vigderman, L.; Khanal, B. P.; Zubarev, E. R. *Adv. Mater. (Weinheim, Ger.)* **2012**, *24*, 4811–4841. doi:10.1002/adma.201201690
19. Lohse, S. E.; Murphy, C. J. *Chem. Mater.* **2013**, *25*, 1250–1261. doi:10.1021/cm303708p
20. Requejo, K. I.; Liopo, A. V.; Zubarev, E. R. *Langmuir* **2020**, *36*, 3758–3769. doi:10.1021/acs.langmuir.0c00302
21. Dement'eva, O. V.; Kartseva, M. E. *Colloid J.* **2023**, *85*, 500–519. doi:10.1134/s1061933x23700187
22. Chow, T. H.; Li, N.; Bai, X.; Zhuo, X.; Shao, L.; Wang, J. *Acc. Chem. Res.* **2019**, *52*, 2136–2146. doi:10.1021/acs.accounts.9b00230
23. Montañó-Priede, J. L.; Rao, A.; Sánchez-Iglesias, A.; Grzelczak, M. *Sci. Adv.* **2025**, *11*, eadx2299. doi:10.1126/sciadv.adx2299
24. Naik, A. M.; Sánchez-Iglesias, A.; Montañó-Priede, J. L.; D'souza, N. M.; Sancho-Parramon, J.; Mezzasalma, S. A.; Rao, A.; Grzelczak, M. *Adv. Opt. Mater.* **2025**, *13*, e01006. doi:10.1002/adom.202501006
25. Campu, A.; Brezestean, I. A.; Tripon, S.-C.; Astilean, S.; Focsan, M. *J. Mater. Chem. C* **2025**, *13*, 16378–16386. doi:10.1039/d5tc01502b
26. Alba-Molina, D.; Cano, M.; Blanco-Blanco, M.; Ortega-Llamas, L.; Jiménez-Gómez, Y.; Gonzalez-Lopez, A.; Perez-Perdomo, M.; Camacho, L.; Giner-Casares, J. J.; Gonzalez-Andrades, M. *J. Mater. Chem. B* **2025**, *13*, 3000–3010. doi:10.1039/d4tb02688h
27. Martín-Sánchez, C.; Sánchez-Iglesias, A.; Barreda-Argüeso, J. A.; Itié, J.-P.; Chauvigne, P.; Liz-Marzán, L. M.; Rodríguez, F. *Nano Lett.* **2025**, *25*, 3588–3596. doi:10.1021/acs.nanolett.4c06473
28. Chen, H.; Kou, X.; Yang, Z.; Ni, W.; Wang, J. *Langmuir* **2008**, *24*, 5233–5237. doi:10.1021/la800305j
29. Lu, M.; Wang, C.; Fan, R.; Lin, M.; Guang, J.; Peng, W. *Photonic Sens.* **2024**, *14*, 240202. doi:10.1007/s13320-024-0709-1
30. Fang, C.; Zhao, G.; Xiao, Y.; Zhao, J.; Zhang, Z.; Geng, B. *Sci. Rep.* **2016**, *6*, 36706. doi:10.1038/srep36706
31. Lee, J.-H.; Gibson, K. J.; Chen, G.; Weizmann, Y. *Nat. Commun.* **2015**, *6*, 7571. doi:10.1038/ncomms8571
32. Chateau, D.; Liotta, A.; Vadcad, F.; Navarro, J. R. G.; Chaput, F.; Lermé, J.; Lerouge, F.; Parola, S. *Nanoscale* **2015**, *7*, 1934–1943. doi:10.1039/c4nr06323f
33. Chateau, D.; Desert, A.; Lerouge, F.; Landaburu, G.; Santucci, S.; Parola, S. *ACS Appl. Mater. Interfaces* **2019**, *11*, 39068–39076. doi:10.1021/acsami.9b12973
34. Nguyen, A. L.; Griffin, Q. J.; Wang, A.; Zou, S.; Jing, H. *J. Phys. Chem. C* **2025**, *129*, 4303–4312. doi:10.1021/acs.jpcc.4c08818
35. Sánchez-Iglesias, A.; Grzelczak, M. *Small* **2025**, *21*, 2407735. doi:10.1002/smll.202407735
36. Salsabiila, N.; Morsin, M.; Nafisah, S.; Razali, N. L.; Mahmud, F.; Tukiran, Z.; Omar, M. A. *J. Adv. Res. Appl. Mech.* **2025**, *127*, 100–119. doi:10.37934/aram.127.1.100119
37. Marcheselli, J.; Chateau, D.; Lerouge, F.; Baldeck, P.; Andraud, C.; Parola, S.; Baroni, S.; Corni, S.; Garavelli, M.; Rivalta, I. *J. Chem. Theory Comput.* **2020**, *16*, 3807–3815. doi:10.1021/acs.jctc.0c00269
38. Montañó-Priede, J. L.; Sánchez-Iglesias, A.; Mezzasalma, S. A.; Sancho-Parramon, J.; Grzelczak, M. *J. Phys. Chem. Lett.* **2024**, *15*, 3914–3922. doi:10.1021/acs.jpclett.4c00582
39. Khlebtsov, N. G.; Zarkov, S. V. *J. Phys. Chem. C* **2025**, *129*, 10958–10974. doi:10.1021/acs.jpcc.5c01652
40. Li, Q.; Zhuo, X.; Li, S.; Ruan, Q.; Xu, Q.-H.; Wang, J. *Adv. Opt. Mater.* **2015**, *3*, 801–812. doi:10.1002/adom.201400505
41. Qi, Y.; Xing, T.-y.; Zhao, J.; Weng, G.-j.; Li, J.-j.; Zhu, J.; Zhao, J.-w. *J. Alloys Compd.* **2019**, *776*, 934–947. doi:10.1016/j.jallcom.2018.10.321
42. Khanadeev, V. A.; Khlebtsov, N. G.; Burov, A. M.; Khlebtsov, B. N. *Colloid J.* **2015**, *77*, 652–660. doi:10.1134/s1061933x15050117
43. Ciraci, C.; Urzhumov, Y.; Smith, D. R. *Opt. Express* **2013**, *21*, 9397–9406. doi:10.1364/oe.21.009397
44. Khlebtsov, N. G.; Zarkov, S. V. *J. Phys. Chem. C* **2024**, *128*, 15029–15040. doi:10.1021/acs.jpcc.4c03126
45. Gladyshev, S.; Pashina, O.; Proskurin, A.; Nikolaeva, A.; Sadrieva, Z.; Petrov, M.; Bogdanov, A.; Frizyuk, K. *ACS Photonics* **2024**, *11*, 404–418. doi:10.1021/acsphotonics.3c01166
46. Khlebtsov, N. G.; Zarkov, S. V.; Khanadeev, V. A.; Avetisyan, Y. A. *Nanoscale* **2020**, *12*, 19963–19981. doi:10.1039/d0nr02531c
47. Olmon, R. L.; Slovick, B.; Johnson, T. W.; Shelton, D.; Oh, S.-H.; Boreman, G. D.; Raschke, M. B. *Phys. Rev. B* **2012**, *86*, 235147. doi:10.1103/physrevb.86.235147
48. Novo, C.; Gomez, D.; Perez-Juste, J.; Zhang, Z.; Petrova, H.; Reismann, M.; Mulvaney, P.; Hartland, G. V. *Phys. Chem. Chem. Phys.* **2006**, *8*, 3540–3546. doi:10.1039/b604856k

49. Khlebtsov, B.; Khanadeev, V.; Pylaev, T.; Khlebtsov, N. *J. Phys. Chem. C* **2011**, *115*, 6317–6323. doi:10.1021/jp2000078
50. Mosquera, J.; Wang, D.; Bals, S.; Liz-Marzán, L. M. *Acc. Chem. Res.* **2023**, *56*, 1204–1212. doi:10.1021/acs.accounts.3c00101
51. Yu, R.; Liz-Marzán, L. M.; García de Abajo, F. J. *Chem. Soc. Rev.* **2017**, *46*, 6710–6724. doi:10.1039/c6cs00919k
52. Khlebtsov, N. G. *J. Quant. Spectrosc. Radiat. Transfer* **2013**, *123*, 184–217. doi:10.1016/j.jqsrt.2012.12.027
53. Vigderman, L.; Zubarev, E. R. *Chem. Mater.* **2013**, *25*, 1450–1457. doi:10.1021/cm303661d
54. Lohse, S. E.; Burrows, N. D.; Scarabelli, L.; Liz-Marzán, L. M.; Murphy, C. J. *Chem. Mater.* **2014**, *26*, 34–43. doi:10.1021/cm402384j
55. Tong, W.; Walsh, M. J.; Mulvaney, P.; Etheridge, J.; Funston, A. M. *J. Phys. Chem. C* **2017**, *121*, 3549–3559. doi:10.1021/acs.jpcc.6b10343
56. Khlebtsov, B. N.; Burov, A. M.; Zarkov, S. V.; Khlebtsov, N. G. *Phys. Chem. Chem. Phys.* **2023**, *25*, 30903–30913. doi:10.1039/d3cp04541b
57. Eremeev, A. V.; Pikina, A. S.; Vladimirova, T. V.; Bogomazova, A. N. *Genes Cells* **2023**, *18*, 5–21. doi:10.23868/gc312198
58. Khlebtsov, N. G.; Khlebtsov, B. N.; Kryuchkova, E. V.; Zarkov, S. V.; Burov, A. M. *J. Phys. Chem. C* **2022**, *126*, 19268–19276. doi:10.1021/acs.jpcc.2c05843
59. Engel, A.; Draut, F.; Demoustier-Champagne, S. M.; Hermans, S. *Langmuir* **2025**, *41*, 15805–15818. doi:10.1021/acs.langmuir.5c00326
60. Etchegoin, P. G.; Le Ru, E. C. *Phys. Chem. Chem. Phys.* **2008**, *10*, 6079–6089. doi:10.1039/b809196j
61. Mayer, K. M.; Hafner, J. H. *Chem. Rev.* **2011**, *111*, 3828–3857. doi:10.1021/cr100313v
62. Miller, M. M.; Lazarides, A. A. *J. Phys. Chem. B* **2005**, *109*, 21556–21565. doi:10.1021/jp054227y
63. Luo, X.; Qiao, L.; Xia, Z.; Yu, J.; Wang, X.; Huang, J.; Shu, C.; Wu, C.; He, Y. *Langmuir* **2022**, *38*, 6454–6463. doi:10.1021/acs.langmuir.2c00663
64. Khlebtsov, N. G. *Quantum Electron.* **2008**, *38*, 504. doi:10.1070/qe2008v038n06abeh013829
65. Khlebtsov, B. N.; Khanadeev, V. A.; Ye, J.; Mackowski, D. W.; Borghs, G.; Khlebtsov, N. G. *Phys. Rev. B* **2008**, *77*, 035440. doi:10.1103/physrevb.77.035440
66. Khlebtsov, N. G.; Dykman, L. A.; Bogatyrev, V. A.; Khlebtsov, B. N. *Colloid J.* **2003**, *65*, 508–518. doi:10.1023/a:1025137322720
67. Khlebtsov, B. N.; Khanadeev, V. A.; Khlebtsov, N. G. *Langmuir* **2008**, *24*, 8964–8970. doi:10.1021/la8010053
68. Khlebtsov, B. N.; Khanadeev, V. A.; Burov, A. M.; Le Ru, E. C.; Khlebtsov, N. G. *J. Phys. Chem. C* **2020**, *124*, 10647–10658. doi:10.1021/acs.jpcc.0c00991
69. Le Ru, E. C.; Blackie, E.; Meyer, M.; Etchegoin, P. G. *J. Phys. Chem. C* **2007**, *111*, 13794–13803. doi:10.1021/jp0687908
70. Khlebtsov, N.; Dykman, L. *Chem. Soc. Rev.* **2011**, *40*, 1647–1671. doi:10.1039/c0cs00018c
71. Dykman, L. A.; Khlebtsov, N. G. *Chem. Rev.* **2014**, *114*, 1258–1288. doi:10.1021/cr300441a
72. AlGhosain, H. M.; Nie, J.; Liu, T.; Lee, J. *Toxicol. Res. (Cham, Switz.)* **2025**, *41*, 489–502. doi:10.1007/s43188-025-00295-y
73. Dykman, L.; Khlebtsov, N. *Chem. Soc. Rev.* **2012**, *41*, 2256–2282. doi:10.1039/c1cs15166e

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