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Al-assisted models to predict chemotherapy drugs modified with C_{60} fullerene derivatives

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Full Research Paper

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Abstract

Employing quantitative structure–activity relationship (QSAR)/ quantitative structure–property relationship (QSPR) models, this study explores the application of fullerene derivatives as nanocarriers for breast cancer chemotherapy drugs. Isolated drugs and two drug–fullerene complexes (i.e., drug–pristine C_{60} fullerene and drug–carboxyfullerene C_{60} –COOH) were investigated with the protein CXCR7 as the molecular docking target. The research involved over 30 drugs and employed Pearson's hard–soft acid–base theory and common QSAR/QSPR descriptors to build predictive models for the docking scores. Energetic descriptors were computed using quantum chemistry at the density functional-based tight binding DFTB3 level. The results indicate that drug–fullerene complexes interact more with CXCR7 than isolated drugs. Specific binding sites were identified, with varying locations for each drug complex. Predictive models, developed using multiple linear regression and IBM Watson artificial intelligence (AI), achieved mean absolute percentage errors below 12%, driven by AI-identified key variables. The predictive models included mainly quantitative descriptors collected from datasets as well as computed ones. In addition, a water-soluble fullerene was used to compare results obtained by DFTB3 with a conventional density functional theory approach. These findings promise to enhance breast cancer chemotherapy by leveraging fullerene-based drug nanocarriers.

Introduction

Breast cancer is the most diagnosed cancer in women and the second leading cause of cancer-related mortality in women [1,2]. Heritage is the most critical risk factor, and 15 to 20% of breast cancer is familiar [3]. One of the characteristics of breast cancer is that it can be wholly cured given an early diagnosis [4]. The mortality rate from breast cancer has been reduced by 1.9% annually from 2002 to 2011 and 1.3% from 2011 to 2020 [5]. Diagnostics and treatments have continuously improved through the years. However, the situation is different in each country considering the costs and technological advances in each country. In the United States, 300,590 cases of breast cancer had been estimated for the year 2023, with a total of 43,700 deaths [6]. Latin America has over 210,000 new cases and around 60,000 deaths yearly [7]. For the year 2020, it was estimated that about 2.3 million breast cancer cases were diagnosed in women globally, and about 685,000 died from this disease [8]. A recurrent problem with standard treatments are the side effects. Regarding the use of chemotherapeutic drugs, such issues are nephrotoxicity of cisplatin, cardiotoxicity of doxorubicin, and pulmonary fibrosis from the use of bleomycin [9-11]. Besides, in the case of radiotherapy, fibrosis, atrophy, and neuronal damage caused by irradiation can occur [12,13]. Consequently, novel treatments try to reduce the secondary effects while retaining the benefits of standard approaches.

Chemotherapy is one of the most extensively applied treatments for breast cancer, with different drug targets depending on the type of cancer. Progesterone- or estrogen-receptor-positive tumors are related to cancers with low mortality [14]. Another common target in chemotherapy is human epidermal growth factor receptor 2 (HER2). Only 15 to 20% of all tumors are HER2-positive, overexpressing Erb-B2 receptor tyrosine kinase 2 (ERBB2) in the cell membrane. HER2 tumors are usually more aggressive than other ones, but the advantage is that their treatment is very effective [15]. Another chemotherapy target is the chemokine C-X-C motif receptor 7 (CXCR7) [16,17]. This G-protein is targeted because studies show a possible positive effect on inhibiting the metastasis of cervical cancer cells [18]. However, more clinical and preclinical studies on CXCR7 and its co-player CXCR4 are required since alterations have been detected in diseases such as cancer, central nervous system and cardiac disorders, and autoimmune diseases [16].

In recent years, nanomaterials have attracted the attention of different scientific communities by providing them with new solutions for drug delivery [19,20]. These nanotechnological applications have made it possible to obtain treatments that release substances at specific sites of interest, reducing the required drug amount and side effects. Nanostructures to form

these drug delivery systems can be divided into organic and inorganic [19,20], with the latter one being the less extensively studied. One option currently considered in pharmacy and medicine is carbon-based nanomaterials because of their physicochemical, mechanical, electrical, thermal, and optical properties [19,20], as well as their capacity to modify existing drugs. Fullerene derivatives have been proposed recently, particularly those obtained from fullerene C₆₀ [21]. The unmodified fullerene C₆₀ is known as a "free radical sponge" because its double bonds tend to accept free radicals [22]. Because of its size, surface area, and capacity to extinguish or generate reactive oxygen species, C₆₀ is very promising in medicine and clinical therapy [23,24]. It is also possible to modify pristine fullerenes by adding polar functional groups (e.g., -COOH, -OH, or -NH₂), to improve water solubility, antioxidant properties, and even biological activity [25]. For instance, polyhydroxy fullerenes (PHFs) exhibit properties suitable for biomedical applications, such as water solubility, biodegradability, biocompatibility, and hypoallergic response. It has been shown that PHFs can inhibit cancer tumor growth and positively regulate the immune system [26]. The same is valid for carboxylated fullerenes [27]; for instance, C₆₀[C(COOH)₂]₃ is well known for its high biological activity in plants [28] and within mitochondrial dynamics [29].

Since the evaluation of novel drugs is a task that requires significant human and material resources, innovative strategies have been formulated as alternatives. Quantitative structure-activity and quantitative structure-property relationships (QSAR/QSPR) are a paradigm that can be useful in choosing promising molecules, considering the information on inactive and active compounds, through in silico approaches. According to the QSAR/ QSPR paradigm, a given activity/property, f, can be modeled using a set of quantitative descriptors, $x_1, x_2, x_3, ..., x_n$, theoretically determined or measured by experiments [30]. A relationship $f(x_1, x_2, x_3,..., x_n)$ can be defined to predict the activity or property of molecules after the evaluation of their quantitative descriptors. However, the QSAR/QSPR paradigm does not explain how to select the descriptors or how to build the mathematical function. Consequently, the following paragraphs discuss basic concepts about selecting descriptors and regression techniques implemented in this manuscript.

Lipinski's rule of five is a compendium of guidelines commonly used to determine if a molecule can be proposed as an orally delivered drug according to its physicochemical properties. According to this rule, a drug compound should have a molecular weight below 500 g/mol, a octanol—water partition coefficient (LogP) below 5, less than five hydrogen bond donor sites, and less than ten hydrogen bond acceptors sites. It is possible to add

two other conditions, namely polar surface area (PSA) $\leq 140 \text{ Å}^2$ and less than ten rotatable bonds [31]. Taking advantage of the readiness of these quantities in public datasets, the current study proposes some of these quantities as potentially suitable descriptors for predictive models. Besides, Pearson's hard-soft acid-base (HSAB) theory suggests other descriptors to describe and predict the interactions between chemical species, such as those between a drug molecule as a ligand and a protein [32]. These quantitative values are based on the vertical ionization energy (I) and electron affinity (A). According to Koopmans' theorem, both can be approximated by $I = -E_{HOMO}$ and A = $-E_{\text{LUMO}}$, where E_{HOMO} is the energy of the highest occupied molecular orbital (HOMO), and E_{LUMO} is the energy of the lowest unoccupied molecular orbital (LUMO). It is advantageous to combine these properties to find out if an interaction between two species will occur and to obtain new quantitative relationships. Another helpful descriptor is the global electrophilicity, calculated as $\omega = \chi^2/2\eta$ [33]. Electrophilicity is related to the energetic stabilization that a species gains by obtaining an additional electron.

Methods

First, 42 drugs related to chemotherapy treatments for breast cancer were proposed. Although the most notable fullerene derivatives for biological applications are those with several hydrophilic groups, the carboxylic acid derivative C₆₀-COOH has been studied as well. Baglayan and coworkers carried out a conformation analysis within DFT to obtain the ground state structure for C₆₀-COOH [34]. In addition, they discussed its usage as a potential drug carrier for the antimetabolic and anticancer drug 5-fluoruracil [34]. Similarly, Parlak and Alver reported a theoretical study on the interactions and stability of paracetamol complexes with C₆₀-COOH [35]. Consequently, this work proposes the interaction of C₆₀-COOH fullerene with anticancer drugs. As a complement, a water-soluble fullerene predicted as stable at the normal human body temperature was proposed to study the interactions with doxorubicin and gemcitabine [36]. The water-soluble fullerene is introduced to avoid known mutagenic reactions related to breast cancer [36]. It was also studied as a potential carrier for bedaquiline, an agent against tuberculosis [37]. The current study only considered molecules and complexes formed with up to 100 atoms to be affordable with our computational resources.

A set of descriptors was chosen to build the dataset, including molecular weight and pK_a [38]. Also, LogP was included, as a descriptor associated with the concentration of a given substance in the aqueous phase of a two-phase octanol-water mixture [39]. Similarly, LogS, related to the water solubility of a substance, was considered. Besides, PSA, as molecular surface associated with charge accumulation due to heteroatoms

and polar groups, as well as polarizability (α) associated with the tendency of a given molecule to acquire an electric dipole moment in the presence of an external electric field were taken into account. The mentioned QSAR/QSPR descriptors were obtained from the Drugbank dataset (https://go.drugbank.com). Initial drug structures and connectivity were also obtained from the simplified molecular input line entry specification (SMILES) retrieved from Drugbank.

Molecular mechanics and density functional-based tight binding (DFTB) with dispersion and solvation corrections were used to obtain the optimized structures of the molecules under study and to compute E_{HOMO} , E_{LUMO} , and ω as quantitative descriptors. As an alternative to the most robust but computationally more expensive density functional theory (DFT) method, DFTB was used. A reference electron density ρ_0 represents the sum of the neutral atomic densities [40]. Within the third-order approach DFTB3, the ground state density $\rho(r)$ is obtained as the reference density ρ_0 perturbed by density fluctuations $\delta\rho$, that is,

$$E_{\text{DFTB3}} \left[\rho_0 + \delta \rho \right] = E_0 \left[\rho_0 \right] + E_1 \left[\rho_0 + \delta \rho \right]$$
$$+ E_2 \left[\rho_0 + \left(\delta \rho \right)^2 \right] + E_3 \left[\rho_0 + \left(\delta \rho \right)^3 \right].$$
(1)

For all calculations within DFTB3, the 3OB parameter set was used [41]. To carry out the global optimization procedure, Balloon 1.8.2 [42] and DFTB+ 17.1 [40] were used for the initial conformational study by genetic algorithms and final optimization at the DFTB3 level, respectively. London dispersion forces were considered in the DFTB3 and global optimization procedures by Lennard-Jones potentials, as implemented in UFF and MMFF94 force fields, respectively. The solvent effect was included by the Born solvation model within DFTB3. The study considered the chemotherapy drugs isolated and interacting with pristine C₆₀ fullerene as well as its carboxylic acid derivative C₆₀-COOH. Eight initial drug-fullerene structures were proposed to obtain their global optimization by means of DFTB3. The drugs were initially set at 1.5 Å of minimal distance from the fullerene. Once the global optimization was done, the same steps as for the isolated drugs were carried out for the molecular docking. The datasets were modified to take into account the effect of the fullerenes. Also, the validation set was reduced because of the large size of the complexes.

The atypical chemokine receptor 3, also known as CXCR7 or G-protein-coupled receptor 159 (GPR159) [16,18,43], was selected as the target protein for molecular docking. The iterative assembly refinement server (I-Tasser) was used to produce an initial structure for the CXCR7 protein by the homology approach. The sequence was extracted from the UniProtKB/

Swiss-Prot dataset. From all homology structures produced by the I-Tasser server, the one with the highest confidence coefficients was selected to produce a reliable initial structure [44]. The lowest-energy structure, as in the study of Muthiah and coworkers [45], was validated using PROCHECK [46] to check the quality of the protein structure. The PDB produced with the previous step was subsequently optimized by an energy minimization through Amber force fields using the USCF Chimera 1.14 toolkit [47]. The secondary structural features were stabilized by TMpred [48] and HMMTOP [49] during energy minimization. Last, the protein was prepared by setting atomic charges and hydrogen atoms and merging the nonpolar groups. Once the structures were optimized, molecular docking was performed with the CXCR7 protein, using Autodock Vina 1.1, to obtain the docking score, established hydrogen bonds, and the binding site (pocket). The above was done for all drugs in the dataset and an external validation set.

IBM Watson AI was used to build the models and to predict the docking score through the Extra Trees regressor algorithm [50,51]. It was also used to obtain the most significant quantum descriptors used in each model. Extra Trees, an abbreviation of "extremely randomized trees", is a mathematical method used to estimate a relationship between data and the covariates [52]. The Extra Trees algorithm creates many decision trees [52], but the sampling of each one is random. Thus, a dataset for each tree contains unique samples. The optimization of the hyperparameters associated with the decision trees obtained was performed by the derivative-free global search algorithm known as RBfOpt, which fits a radial basis function mode to accelerate the discovery of the hyperparameters [53]. All the above was used through the AutoAI tool within IBM Watson, an automatized routine to select the model with the best performance among those available in the platform. Since this method does not produce exportable mathematical models, another approach was used as detailed below [50].

Multiple linear regression (MLR) could be a tool to solve the problem in a complementary way to Extra Trees regression. MLR is a mathematical model that can be seen as an extension of linear regression. In terms of n input variables, $x_1, x_2, ..., x_n$, the outcome y can be expanded by the following linear expansion [54]:

$$y = \beta_0 + \beta_1 \times x_1 + \beta_2 \times x_2 + \beta_3 \times x_3 + \dots + \beta_n \times x_n.$$
 (2)

In Equation 2, β_k are the partial regression coefficients, and β_0 is the value of y when all variables are set to zero.

To obtain the AI and MLR models, a fivefold approach was implemented, by using 80% of the data available to obtain the

predictive model as training set and the remaining 20% as testing set. Supporting Information File 1 gives the results of the cross-validation for all the models reported in the current manuscript. Once the models were built, an additional external validation set was used to obtain evaluation metrics and to determine the most accurate models between methodologies. The metrics proposed to evaluate the performance of the predictive models were mean squared error (MSE), mean absolute percentage error (MAPE), mean absolute error (MAE), and root mean squared error (RMSE). These metrics were computed as follows:

MAPE =
$$(1/n)\sum_{i=1}^{n} |(y_i - \hat{y}_i)/y_i| \times 100\%$$
,
MAE = $(1/n)\sum_{i=1}^{n} |(y_i - \hat{y}_i)|$,
MSE = $(1/n)\sum_{i=1}^{n} (y_i - \hat{y}_i)^2$,

and

RMSE =
$$[(1/n)\sum_{i=1}^{n}(y_i - \hat{y}_i)^2]^{1/2}$$
.

Here, y_i is the docking score for compound I, \hat{y}_i is the estimated value of the docking score for compound I provided by the model. The workflow diagram in Figure 1 summarizes the procedure followed to obtain the models.

Results and Discussion

Table 1 presents the quantum descriptors proposed for the current study and the symbols used for them. The physical unit of each descriptor, as well as references to their usage in similar QSAR/QSPR models, were included as well.

Isolated drugs

A dataset containing all the descriptors of Table 1 for 33 drugs was created to obtain the predictive models. Also, another nine compounds were considered to build an external validation set, allowing for the comparison between methodologies (Supporting Information File 1, Table S1). In the case of the training set, the molecular weight was obtained with values between 130.08 and 915.4 g/mol. Water solubility values varied between 0.0004 mg/mL and 22.3 mg/mL. The LogP values varied between -2 and 6.54, whereas LogS ranged from -6 to -1.1. Besides, pK_a values varied between -8 and 14.55. The hydrogen acceptor count varied widely between 2 and 13, whereas the hydrogen donor count varied between 0 and 6. In addition, the polar surface area had variations between 12.47 and 221.29 Å². The cases of thiotepa and aldoxorubicin were not considered because they are part of the validation set. Rotatable bonds were obtained ranging from 0 to 15. The polarizability varied from 9.46 to 87.46 Å³; everolimus was excluded as part of the

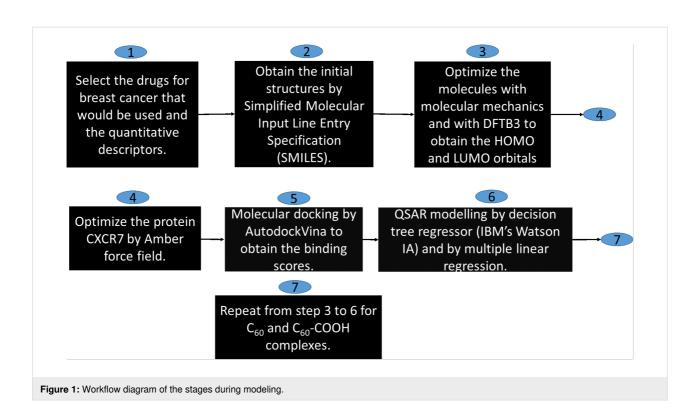


Table 1: Quantitative descriptors proposed to model the docking score of the isolated drugs, as well as of those modified with fullerenes C_{60} and C_{60} –COOH, interacting with the protein CXCR7.

/ariable	Descriptor	Symbol	Unit	Reference
1	molecular weight	MW	g/mol	[55,56]
2	water solubility	WS	mg/mL	[57]
′ 3	octanol-water partition coefficient	LogP	_	[58,59]
4	solubility coefficient	LogS	_	_
5	acid dissociation constant	р K_{a}	_	[60]
6	hydrogen acceptor count	Ac	_	[61]
7	hydrogen donor count	Dn	_	[61]
' 8	polar surface area	PSA	Å ²	[62]
. 9	rotatable bond count	RBC	_	[63]
10	polarizability	α	Å ³	[64]
11	number of rings	NOR	_	[65]
12	energy of HOMO	E_{HOMO}	eV	[66,67]
13	energy of LUMO	E_{LUMO}	eV	[66,67]
14	electrophilicity	ω	eV	[66,67]

external validation set. Also, values of number of rings were obtained from 0 to 9. The energy of the HOMO was computed ranging from -7.400 to -4.392 eV, and the LUMO energy from -5.341 to -0.889 eV. Finally, the electrophilicity varied from 2.12 to 180.39 eV. Figure S1 (Supporting Information File 1) shows the correlation matrix between the ten most relevant quantum descriptors used to obtain the mathematical models. There are significant correlations between the molecular weight

and the polarizability of about 0.93 and between polarizability and the number of rings of about 0.88. Also, molecular weight and number of rings, as well as WS and LogS exhibited considerable correlations, with values of 0.87 and 0.73, respectively. However, all variables showed a correlation below 0.95. Once the drugs were optimized, blind molecular docking was performed with the CXCR7 protein to obtain the docking score, number of established hydrogen bonds, and the protein residues

interacting with the ligands in a coordination sphere of 3 Å. The results obtained with Autodock Vina [47,68] for training-testing and validation sets are shown in Table 2.

The docking scores ranged from -10.1 to -4.6 kcal/mol for the training set. The molecule with the most significant bond strength, according to its docking score, was olaparib, whereas the one with the lowest bond strength was fluorouracil. The number of hydrogen bonds was computed ranging from 0 to 5. It is important to note that the number of hydrogen bonds is not directly related to the docking score since there are weak and

strong hydrogen bonds. This assumption was proved by the analysis performed to obtain the predictive models, as discussed below.

According to the results of the interacting residues annotated in Table 2, two things can be highlighted. First, the analyzed isolated drugs bind inside the protein CXCR7 (Figure 2); second, the pocket is similar for several analyzed drugs. For example, the leucine residue Leu297 is shared by eleven drugs, indicating that their binding zone is close to each other. Comparing the interacting residues with those recently obtained by Muthiah et

Table 2: Docking score, number of established H-bonds, and protein–ligand interacting residue in three-letter symbol, up to 3 Å distance. Drugs marked with an asterisk were used as external validation set.

Ligand	Docking score (kcal/mol)	H-bonds	Interacting residues
doxorubicin	-8.4	5	Asp275, Phe294, Leu297, Tyr200, His298, Arg197, Ser198, Cys196, Asp179, Trp100
neratinib maleate	-8.5	2	lle276, Leu297, Phe294, His298, Tyr 268, Gln30, Ser216, Ser198, Phe129, Asp179, Leu183, His121, Phe124
epirubicin	-8.1	2	Asp275, Leu297, Tyr200, Ser198, Arg197, Cys196, His121, Trp100, Leu104, His298, Phe294, Gln301,
lapatinib ditosylate	-8.9	1	Glu213, Ile276, Val272, Leu297, Tyr200, His298, Arg197, Ser198, Leu183, His121, Cys196,
fulvestrant	-7.9	0	Leu297, Ile276 Val272, Tyr268, Gln301, Trp100, Phe124, Cys196, Asp179, Ser198, Ser216, Arg197
dinaciclib	-8.4	1	Leu297, Gln301, Tyr268, Trp100, Cys196, Arg197, Phe124, Leu128, Ser198, Asp179, Phe129, Met212, Ser216
abemaciclib	-9.8	1	Asn108, Trp100, Trp110, Leu104, Arg197, Lys40, Asn36, Pro38, Leu43, His298
gemcitabine	-6.2	0	Asn36, Phe294, Met37, Pro38, Asn39, Leu43, Lys40, Leu104, Asn108
voruciclib	-8.8	0	Asn36, Pro38, Asn39, His298, Leu43, Lys40, Leu104, Asn108, Trp100 Ser103, Trp110, Arg197
fluorouracil	-4.6	1	Asn319, Asn321, Lys73, Ile83, His80
letrozole	-8.1	0	His298, Gln301, Arg197, Leu104, Trp100, Ser198, His121, Trp110, Leu183
olaparib	-10.1	2	Tyr268, Val272, Ile276, Glu213, Leu209, Ile205, Met212, Tyr200, Phe199, Ser198, Leu128, Phe124
paclitaxel	-9.6	0	His298, Phe294, Leu297, Tyr268, Gln301, Val272, Leu104, Trp100, Phe124, His121, Trp110, Arg197, Ser198, Leu209, Tyr200, Leu183
seliciclib	-7.6	0	Leu183, Asp179, Ser125, Phe124, Phe129, Leu128, Ser216, Tyr268, Leu297
ixabepilone	-8.2	0	Leu297, Val272, Tyr268, Trp265, Phe124, Arg197, His298, Leu128, Ser198, Leu209, Tyr200, Met212, Ser216
anastrozole	-7.7	0	Met212, Leu209, Ser216, Tyr200, Phe129, Asp179, Ser125, Ser198, Phe124, Leu183, His121
pentostatin	-6.3	3	Phe294, Lys40, Leu104, Trp100, Asn108, Trp110
alvocidib	-8.6	1	His298, Leu43, Lys40, Leu104, Trp100, Asn108, Ser103, Trp110, Arg197
methotrexate	-8.5	3	Leu104, Gln301, Tyr268, Val272, Trp100, Trp110, Cys196, Phe124, His121, Ser198, Leu183, Asp179, Phe129, Glu213, Ser216
milciclib	-9.2	0	Lys40, Phe294, Leu297, Leu104, Asn108, Tyr268, Arg197, Val272, His121
ribociclib	-9.5	1	Lys40, Leu104, Leu43, Tyr195, Cys196, Arg197, Phe124, His121, Ser198, Leu183, Asp179

Table 2: Docking score, number of established H-bonds, and protein–ligand interacting residue in three-letter symbol, up to 3 Å distance. Drugs marked with an asterisk were used as external validation set. (continued)

exemestane	-8.0	0	Phe294, Leu43, Lys40, Leu104, Trp110
tamoxifen	-7.9	1	Asn39, Pro38, Leu43, Lys40, Leu104, Trp110, Tyr195, Arg197
idarubicin	-8.8	1	Phe294, His298, Leu43, Leu104, Asn108, Ser103, Trp110, Cys196, His121
palbociclib	-8.3	1	Phe294, Asn36, Pro38, Leu43, Lys40, Leu104, Asn108, Arg197, Cys196
toremifene	-7.4	0	Leu43, Asn39, Leu104, Pro38, Lys40, Asn36, Phe294, Trp110, Tyr195 Arg197
vinblastine	-9.5	2	Leu43, Asn39, Leu104, His298, Phe294, Lys40, Asn108, Trp110, Arg197, Tyr195, Ile205, Leu209
pirarubicin	-9.4	1	Val272, Leu297, Tyr268, Gln301, Met212, Ile205, Leu209, Glu213, Arg197, Phe199, Trp110, Cys196, Phe199,
roniciclib	-8.4	1	Asn39, Met37, His298, Phe294, Pro38, Asn36, Trp100, Leu104, Lys40 Ser103, Asn108, Trp110,
capecitabine	-7.7	2	Gln301, Phe129, Phe124, Ser198, Arg197, His121, Cys196, Trp110, Leu183
sulfanilamide	-5.4	2	Tyr268, Val272, Glu213, Ser216, Leu128, Phe129, Met212, Ser125
zoledronic acid hydrate	-5.7	1	Tyr268, Glu213, Ser216, Phe129, Phe124, Ser125, Ser198, Met212, Asp179
pamidronic acid	-4.8	4	Val272, Tyr268, Trp265, Val217, Glu213, Ser216, Val215, Phe129, Met212, Val175, Phe124, Ser125, His121, Ser76, Tyr200, Asp179
docetaxel*	-9.5	2	Leu209, Glu213, Met212, Ser198, Ser216, Arg197, Leu183, Phe124, His121, Trp110, Ile276, Val272, Leu297, Tyr268, His298, Gln301, Leu104
topotecan*	-8.2	2	Tyr268, His298, Gln301, Trp100, Leu104, Trp110, Cys196, Phe124, Leu183, Ser198
tucatinib*	-10.1	1	Val272, Tyr268, Leu104, Arg197, Ser198, Trp110, His121, Phe124, Asp179, Leu183
aldoxorubicin*	-8.4	1	lle205, Leu209, Tyr200, Ser198, Asp179, Arg197, Leu193, Asp179, His121, Phe124, Ser125, Trp110, Asn108, Leu104, His298, Gln301, Leu297, Val272, Tyr268
vinorelbine*	-9.1	0	Leu297, Tyr268, Phe294, His298, Gln301, Leu43, Leu47, Leu104, Arg197, Trp110, Ser198, Tyr200, Leu209
etoposide*	-9.3	2	Lys 40, Leu43, Asn108, Leu104, Trp100, Trp110, His121, Phe124, Leu183, Gln301
pemetrexed*	-9.1	2	Leu104, Ser103, Trp100, Trp110, Cys196, His121, Phe124, Leu183, Ser198, Tyr200, Val272, Glu213, Ser216
everolimus*	-9.2	3	Leu209, Met33, Asn36, Phe294, His298, Asn39, Leu43, Lys40, Leu104, Asn108, Tyr195, Arg197
thiotepa*	-3.9	1	Tyr268, Val272, Ser216, Phe124

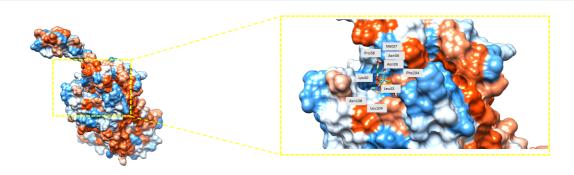


Figure 2: Gemcitabine after blind docking with the protein CXCR7. Blue color indicates the most hydrophilic sites and orange-red the most hydrophobic ones. Interacting residues are labeled as well.

al. [45], it is possible to conclude that the pockets are similar. For instance, doxorubicin was obtained in both cases with Asp179, Cys196, and Trp100 as interacting residues. Also, similar pockets could be obtained because the selected drugs are mostly designed to serve as chemotherapy agents for breast cancer. Thus, it is possible to assume that several drugs share a common mechanism of action and, subsequently, a common protein target, such as CXCR7. For instance, gemcitabine, shown in Figure 2, shares the pocket within CXCR7 with several drugs.

The quantitative descriptors included in the produced models are discussed next. Table 3 contains the used quantitative descriptors and the importance that each one has in the mathematical models. The docking score was the predicted variable in all cases. In addition, the correlation matrix (Supporting Information File 1, Figure S1) was used to build the models. Among the IBM Watson AI models, the first one was obtained using all the initially proposed quantitative descriptors (Table 1). In the second and fourth models, polarizability was not used because, as shown in the correlation matrix, it was found to be closely related to molecular weight. The third and fifth models did not consider molecular weight because of the same relationship with polarizability. Also, the descriptors pK_a , Ac, and PSA were not considered because their importance in the previous models was below 10% (Table 3). Although conceptually different, $E_{\rm HOMO}$ was discarded because it is related to $E_{\rm LUMO}$ and electrophilicity. The last model did not include α because of its relationship with MW. It is important to notice that the most important variables in the six models were NOR, polarizability, LogS, MW, and WS. The least important variables in the six models were $E_{\rm LUMO}$, p $K_{\rm a}$, PSA, $E_{\rm HOMO}$, and Ac; they all had less than 10% importance in all models. Hence, the computed $E_{\rm HOMO}$ and $E_{\rm LUMO}$ values were not particularly useful in predicting the ligand–protein docking score and, subsequently, the docking score.

To compare the performance offered by the Extra Trees algorithm of Watson AI, a comparison with a family of MLR models was made. Supporting Information File 1 contains the cross-validation for all reported MLR models. Table S2 (Supporting Information File 1) shows the docking scores obtained for the validation set by using both methodologies. With these values, it is possible to appreciate the difference between the docking score obtained directly by molecular docking through Autodock Vina and the prediction of the mathematical models for the external validation set.

To clearly state the performance comparison between AI and MLR, Table 4 reports the values computed for the evaluation metrics proposed for each model. The MSE ranged from 0.30 to 1.73 kcal²/mol², whereas the MAPE varied from 6.1 to 16.37%. Also, MAE values from 0.46 to 1.13 kcal/mol and RMSE values from 0.55 to 1.32 kcal/mol were obtained. The above shows that both AI and MLR approaches accurately model the protein–ligand docking score, yielding higher confidence in the case of Extra Tree regressor models. The best performance, according to the computed minimum errors, was obtained in the

Table 3: Input variables (IV) and output importance (OI) of six Extra Tree regressor models obtained from IBM Watson. Variables are annotated according to Table 1. The best model, according to the MAPE values, is highlighted in bold.

Мо	del 1	Мо	del 2	Мо	del 3	Мо	del 4	Мо	odel 5	Мо	odel 6
IV	OI (%)	IV	OI (%)	IV	OI (%)	IV	OI (%)	IV	OI (%)	IV	OI (%)
NOR	100	WS	100	ws	100	LogS	100	NOR	100	LogS	100
α	57	NOR	97	LogS	99	MW	53	α	74	NOR	15
LogS	43	LogP	94	α	56	NOR	26	LogS	49	E_{LUMO}	7
WS	38	LogS	87	NOR	39	WS	17	WS	31	MW	6
MW	33	MW	20	ω	18	ω	12	LogP	19	ω	5
LogP	32	Dn	14	E_{LUMO}	6	E_{LUMO}	5	Dn	13	RBC	0
Ac	8	RBC	12	E_{HOMO}	5	E_{HOMO}	3	E_{LUMO}	5	Dn	0
Dn	6	E_{LUMO}	7	PSA	4	PSA	3	RBC	0	Ws	0
E_{HOMO}	2	PSA	4	р K_a	1	р K_{a}	1	ω	0	LogP	0
RBC	1	E_{HOMO}	3	LogP	1	RBC	0	_	_	_	_
PSA	1	р K_{a}	2	RBC	0	Ac	0	_	_	_	_
ω	1	Ac	1	Dn	0	Dn	0	_	_	_	_
pK _a	0	Ω	0	Ac	0	LogP	0	_	_	_	_
E_{LUMO}	0	_	_	_	_	_	_	_	_	_	_

Error	Мо	del 1	Mo	odel 2	Mo	odel 3	Mo	odel 4	Мо	del 5	Мо	del 6
	Al	MLR	AI	MLR	AI	MLR	Al	MLR	Al	MLR	Al	MLR
MSE (kcal²/mol²)	0.93	1.69	0.64	1.73	0.30	1.02	0.43	1.51	0.82	1.37	0.73	1.10
MAPE (%)	11.51	16.11	9.65	16.37	6.17	11.98	6.70	15.69	10.92	14.10	10.34	12.49
MAE (kcal/mol)	0.77	1.11	0.64	1.13	0.46	0.82	0.51	1.08	0.71	0.98	0.66	0.83
RMSE (kcal/mol)	0.97	1.30	0.80	1.32	0.55	1.01	0.66	1.23	0.91	1.17	0.85	1.05

Table 4: Comparison metrics obtained by the use of Al and MLR in the case of isolated drugs. The best model, according to the MAPE values, is highlighted in bold. The values were computed relative to the validation set.

case of Watson AI model 3. In contrast, the maximum error was obtained in MLR model 2. Thus, the variables denoted in this work can be used for other authors to propose novel chemotherapy drugs assuming CXCR7 as a target.

As mentioned above, the best model was AI model 3 with a MAPE of about 6.17%. Since the AI models are not exportable, our best model is represented by the following functional form: AI3_{DRUG} = f(WS, LogS, α , NOR, ω , E_{LUMO} , E_{HOMO} , PSA, pK_a , LogP). The model with the lowest MAPE among those obtained by MLR, computed as 11.98%, is model 3, represented as follows:

$$\begin{split} \text{MLR3}_{\text{DRUG}} &= -5.288 + 0.049 \times \text{WS} + 0.318 \times \text{LogS} \\ &+ 0.048 \times \alpha - 0.637 \times \text{NOR} + 0.009 \times \omega \\ &- 0.054 \times E_{\text{HOMO}} + 0.113 \times E_{\text{LUMO}} \\ &- 0.008 \times \text{PSA} - 0.234 \times \text{LogP} - 0.014 \times \text{pK}_{\text{a}}. \end{split} \tag{3}$$

Thus, the AI was useful in selecting the most relevant variables for the formulation of the linear, accurate, and exportable model. Thus, AI is a valuable guide to obtain mathematical models with other methodologies such as MLR. Another significant model is MLR model 6 with the second lowest MAPE of 12.49%. It is worth noting that the model is compact and includes the computed descriptors E_{HOMO} and ω , as well as experimentally determined ones.

$$\begin{split} \text{MLR6}_{\text{DRUG}} &= -4.2179 + 0.4131 \times \text{LogS} - 0.5274 \times \text{NOR} \\ &+ 0.2869 \times E_{\text{HOMO}} + 0.0009 \times \text{MW} \\ &+ 0.0088 \times \omega. \end{split} \tag{4}$$

Although the best AI model was model 3, model 4 is relevant as well. It has the following functional form: $AI4_{DRUG} = f(LogS, MW, NOR, WS, \omega, E_{LUMO}, E_{HOMO}, PSA, pK_a)$. For this model, the MAPE is about 6.70%. The variables used can be mostly evaluated by computational methods, except for pK_a and LogS.

Drugs modified with C₆₀

Since this study aims to elucidate the potential use of AI suites, such as Watson, to predict the docking score of pristine and modified chemotherapy drugs, the following paragraphs detail the extension of our datasets and models to drugs modified with potential nanocarriers. First, a dataset with 28 drugs, extracted from public datasets or modified from the data annotated in the previous case, was built with the corresponding quantitative descriptors to study complexes of the drugs with fullerene C₆₀ or a simple C₆₀-COOH derivative [29]. The resultant dataset is shown in Supporting Information File 1, Table S3. Complexes of the drugs with C₆₀ or its derivative with more than one hundred atoms were excluded to save computational resources. Because of that, some maximums and minimums were modified in the dataset. Also, quantities such as the molecular weight or number of rings were shifted to the correspondent values for the drug-fullerene complexes since these modifications are only additive constants. Molecules interacting with C₆₀ were studied in this subsection and those modified with the fullerene derivative are the described in the subsequent subsection.

In case of the complexes with C_{60} , the molecular weight is obtained with values above 907.1639 g/mol. The above imposes some restrictions on the usage of fullerene derivatives as drug nanocarriers, since it is accepted that common pharmacological agents applied in topical therapies are under 500 Da [69]. For simplicity, WS, LogP, LogS, p K_a , and α values were taken from isolated drugs. In the case of Dn, Ac, RBC, and PSA, values of the isolated drugs were also used. This is because fullerene is not expected to modify these descriptors since its chemical constitution lacks polar groups or donor/acceptor atoms. The number of rings was only modified with the addition of the fullerene rings from 32 to 39.

As stated in the Methods section, HSAB descriptors were computed for the drug– C_{60} complexes in their ground states while drugs with C_{60} were structurally optimized at the DFTB3 level. Since introducing a species known to act as an electron acceptor, such as fullerene C_{60} , could modify the electron struc-

ture of the modified species [70,71], the energies of the frontier orbitals were recomputed for the modified drugs in their ground states. The energy of the HOMO varied between -3.458 and -5.718 eV. The energy of LUMO ranged from -3.179 to -5.388 eV. The electrophilicity, computed through Koopman's theorem, varied from -134.88 to 280.8 eV. Once the drugs with fullerene C_{60} were globally optimized, molecular docking was conducted with the CXCR7 protein to acquire the docking

score, number of established hydrogen bonds, and protein residues interacting with the complex at a distance of 3 Å. The results obtained with Autodock Vina for the 24 complexes and the validation set are presented in Table 5.

The protein-ligand docking score between drug- C_{60} and CXCR7 varied from -13.6 to -9.2 kcal/mol. These values are consistently higher than those computed for the isolated drugs.

Table 5: Docking score, number of established H-bonds after docking, and protein–ligand interacting residues up to 3 Å distance obtained for the drugs modified with C_{60} fullerene. Drugs with the asterisk were used as an external validation set.

Ligands	Score (kcal/mol)	H-bonds	Interacting residues
doxorubicin	-9.2	2	Arg323, Ser316, Val313, Leu326, Phe330, Leu340, Leu343, Val64, Ile57, Ile60
epirubicin	-9.2	0	Arg323, Ser316, Leu326, Tyr322, Leu340, Leu343, Lys342, Val64, Pro312, Ala61, Ile60
lapatinib ditosylate	-9.4	0	Arg323, Ser316, Phe330, Leu326, Tyr322, Leu340, Lys342, Leu343, Val64, Ala61, lle60
fulvestrant	-11.7	0	Tyr322, Pro312, Leu326, Ala61, Val64, Ile60, Leu340, Leu343, Lys342
dinaciclib	-10.2	1	Val313, Ile60, Ala61, Val64, Leu326, Leu340, Thr340
abemaciclib	-10	0	Arg323, Leu326, Ser316, Tyr322, Leu340, Leu343, Lys342, Val64, Ala61, Ile60
gemcitabine	-10.9	1	Val313, Ile57, Ala61, Ile60, Val64, Leu326, Leu340, Leu343, Phe330
voruciclib	-10.2	0	Val64, Ala61, Ile57, Ile60, Val313, Leu326, Leu340, Thr341, Leu343
fluorouracil	-11.6	0	Pro312, Tyr322, Leu326, Leu340, Leu343, Lys342, Ala61, Val64, Ile60
olaparib	-12.4	2	Val313, Ala61, Ile60, Ile57, Arg323, Leu326, Leu340, Lys342
ixabepilone	-11.7	2	Arg323, Ser316, Val313, Ala61, Val64, Ile60, Leu326, Leu340, Lys342, Leu343
alvocidib	-12.1	0	Val313, Ser316, Tyr322, Leu326, Leu340, Leu343, Lys342, Val64, Ala61, Ile60, Ile57
methotrexate	-9.7	2	Ser316, Pro312, Tyr322, Leu326, Val65, Ala61, Val64, Ile60, Phe330, Leu340, Leu343, Thr341, Lys342
ribociclib	-12.4	0	Ser198, Arg197, Cys196, Tyr195, Trp110, Asn108, Leu104, Lys40, Ser90, Phe94, Asn36, Asp30, Val28
exemestane	-13.6	0	Val313, Pro312, Ser316, Tyr322, Val65, Ala61, Ile57, Val64, Ile60, Leu326, Leu340, Leu343, Lys34
tamoxifen	-9.8	0	lle66, Leu84, lle70, lle88, Leu91, Trp92
idarubicin	-10.1	1	Ser316, Tyr322, Leu326, Leu340, Leu343, Lys342, Ile57, Ile60, Ala61, Val64
palbociclib	-11.2	0	Val313, Ala61, Ile57, Ile60, Val64, Leu343, Leu326
toremifene	-9.9	0	Val313, Ile57, Ile60, Val64, Leu326, Ser350, Leu340, Leu343
roniciclib	-11	2	Met33, Pro35, Asn36, Asp30, Val28, Asn39, Pro38, Ile27, Leu43, Lys40 His107
capecitabine	-9.5	0	Ser316, Tyr322, Leu326, Ala61, Val64, Ile60, Ile57, Leu340, Leu343, Lys342
sulfanilamide	-10.6	1	Cys165, Arg162, Ile166, Leu84, Ile70, Trp92
zoledronic acid hydrate	-9.8	2	Val313, Ile60, Ala61, Ile57, Leu326, Leu340, Lys342, Thr341
pamidronic acid	-9.9	1	Val313, Ser316, Tyr322, Pro312, Leu326, Val64, Ala61, Ile60, Leu340, Lys34, Leu343
topotecan*	-10.1	0	Lys342, Leu343, Leu340, Phe330, Leu326, Ser316, Val313, Ala61, Ile57 Val64, Ile60
tucatinib*	-11	0	Lys342, Leu343, Leu340, Tyr322, Leu326, Val64, Ala61, Ile60, Ile67
pemetrexed*	-11.5	0	Lys342, Thr341, Leu343, Leu340, Phe330, TTyr322, Val64, Ser316, Ala61, Ile60
thiotepa*	-9.9	0	Lys342, Leu343, Leu340, Tyr322, Leu326, Ser316, Val313, Val64, Ile60

In this case, examestane had the highest docking score, whereas epirubicine had the lowest docking score. The number of hydrogen bonds was between 0 and 2. Considering the results of the interacting residues, two things can be highlighted. First, drugs modified with fullerene C₆₀ bind outside the protein; second, these binding sites are similar for the analyzed compounds (Figure 3). For example, the serine residue labeled as Ser316 is shared by 14 drugs, indicating that their binding zone is close to each other. Figure 3 shows the binding site between the protein and gemcitabine and the interacting residues.

The predictive QSAR/QSPR models were obtained as in the previous section. Table 6 shows the used quantum descriptors and the importance that each one has in the mathematical models for the drug– C_{60} complexes. The docking score was the predicted variable in all cases. The models for C_{60} were taken directly from the models of the isolated drugs. For example, model 1 for drug– C_{60} complexes considered variables with

importance greater than zero from model 1 in the case of isolated drugs. The previous procedure was performed for all models. The most critical variables for the six models were LogP, pK_a , and PSA. The least important variables in the six models were WS and Ac because they have less than 10% importance in the models.

To give a quantitative reference about the performance of the predictive models, Supporting Information File 1, Table S4, shows the scores obtained for the validation set using both methods, the Extra Tree algorithm of IBM Watson and multiple linear regression. With these values, it is possible to appreciate the difference between the docking score obtained by molecular docking and the predictive models for the drug–C₆₀ complexes. Table 7 shows the values obtained for the different evaluation metrics for each predictive model. The MSE ranges from 0.44 to 12.77 kcal²/mol², and the MAPE varies from 4.97 to 31.5%. The MAE ranged from 0.5 to 3.26 kcal/mol, whereas the RMSE varied from 0.66 to 3.57 kcal/mol. The minimum

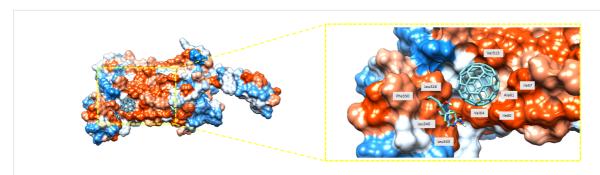


Figure 3: Binding site of gemcitabine—C₆₀ and protein CXCR7. Blue color indicates the most hydrophilic sites and orange-red the most hydrophobic ones. Interacting residues are labeled as well.

Table 6: Input variables (IV) and output importance (OI) for six Extra Tree regressor models obtained from IBM Watson. Variables are annotated according to the notation introduced in Table 1. The best models, according to their MAPE values, are highlighted in bold.

Мо	del 1	Мс	del 2	Мо	odel 3	Мо	del 4	Мо	odel 5	Мо	odel 6
IV	OI (%)	IV	OI (%)	IV	OI (%)	IV	OI (%)	IV	OI (%)	IV	OI (%)
LogP	100	pK _a	100	p <i>K</i> a	100	pK _a	100	LogP	100	MW	100
PSA	97	LogP	55	PSA	69	PSA	66	Dn	58	E_{LUMO}	96
RBC	24	PSA	49	LogP	56	LogS	32	α	18	ω	71
E_{HOMO}	23	E_{LUMO}	29	E_{LUMO}	24	E_{LUMO}	25	E_{LUMO}	10	NOR	34
Dn	17	Dn	20	WS	8	NOR	20	NOR	6	LogS	0
ω	11	LogS	7	E_{HOMO}	5	WS	8	LogS	1	_	_
α	8	E_{HOMO}	1	ω	3	E_{HOMO}	3	WS	0	_	_
MW	4	Ac	0	α	3	ω	1	_	_	_	_
LogS	3	RBC	0	LogS	0	MW	0	_	_	_	_
NOR	1	MW	0	NOR	0	_	_	_	_	_	_
WS	1	NOR	0	_	_	_	_	_	_	_	_
Ac	0	WS	0	_	_	_	_	_	_	_	_

Error	Мо	del 1	Мо	del 2	Mo	odel 3	Мо	del 4	Mo	odel 5	Mo	odel 6
	Al	MLR	Al	MLR	Al	MLR	Al	MLR	Al	MLR	Al	MLR
MSE (kcal²/mol²)	1.62	12.77	3.9	0.86	3.61	3.75	2.38	0.44	1.96	2.22	0.93	1.77
MAPE (%)	10.99	31.5	18.62	7.58	17.3	17.36	12.96	4.97	12.6	12.8	7.53	11.94
MAE (kcal/mol)	1.17	3.26	1.97	0.77	1.8	1.8	1.38	0.5	1.36	1.34	0.82	1.24
RMSE (kcal/mol)	1.27	3.57	1.97	0.92	1.9	1.94	1.54	0.66	1.4	1.49	0.96	1.33

Table 7: Metrics obtained by using IA and MLR for drug–C₆₀ complexes. The best models, according to their MAPE values, are highlighted in bold. The values were computed relative to the validation set

error for all four metrics was obtained in MLR model 4, while the maximum error was obtained in MLR model 1. Considering the MAPE, the best model, with a value of 4.97%, to predict the docking score is MLR model 4. The explicit form of this model is:

$$\begin{split} \text{MLR4}_{\text{DRUG+C60}} &= -10.253 + 0.179 \times \text{p}K_{\text{a}} - 0.003 \times \text{PSA} \\ &+ 0.292 \times \text{LogS} + 0.719 \times E_{\text{LUMO}} \\ &- 0.071 \times \text{NOR} - 0.057 \times \text{WS} \\ &- 1.020 \times E_{\text{HOMO}} + 0.006 \times \omega. \end{split} \tag{5}$$

This linear model exhibited a higher performance than the non-exportable approaches provided by the AI. However, one needs to remember that the variables included in model 4 were selected by the initial AI screening. Thus, the selection of variables using AI offers a significant improvement for modeling using other mathematical methods. In addition, all metrics obtained in the case of MLR model 4 are better than those calculated in the case of the best MLR model of the isolated drugs and are comparable to those of the best AI model. Another significant MLR model is the model 2 with a MAPE of 7.58% and the following linear function:

$$\begin{split} \text{MLR2}_{\text{DRUG+C60}} &= -12.884 + 0.141 \times \text{p} K_{\text{a}} + 0.139 \times \text{LogP} \\ &- 0.10 \times \text{PSA} + 0.649 \times E_{\text{LUMO}} \\ &- 0.261 \times \text{Dn} + 0.268 \times \text{LogS} \\ &- 0.828 \times E_{\text{HOMO}}. \end{split} \tag{6}$$

The best AI model is model 6 with a MAPE of about 7.53% and the functional form AI6_{DRUG+C60} = $E(MW, E_{LUMO}, \omega, NOR)$; all variables can be evaluated by theoretical approaches without the necessity of experimental results. The other significant model from AI, model 1, yielded a higher MAPE value of about 10.99%. In this case, the functional form is AI1_{DRUG+C60} = $E(LogP, PSA, RBC, E_{HOMO}, Dn, \omega, \alpha, MW, LogS, NOR, WS)$. Despite the large number of variables, including theoretical and experimental ones, the error is larger than those of the previously discussed models.

Drugs modified with C₆₀-COOH

To elucidate the effect of a fullerene derivative, the carboxyfullerene C₆₀-COOH was chosen. A dataset with 19 drugs for the predictive model and four drugs as the validation set was built. The resultant dataset is shown in Supporting Information File 1, Table S5. As in the previous systems, the dataset was reduced to systems with less than 100 atoms. Because of this, ranges of the descriptors and their contributions to the predictive models were modified in the dataset. The molecular weights were increased to values ranging from 915.8022 to 1292.425 g/mol. As in the previous case, WS, LogS, pK_a , and LogP are the same as those obtained for the isolated drugs. The hydrogen acceptor count varied between 3 and 13, whereas the hydrogen donor count varied from 1 to 7. Since the carboxylic group is polar, polar surface area values, ranging from 49.77 to 243.22 Å², were modified. Also, after the introduction of the polar group, the RBC varied from 1 to 10.

The fullerene derivative C₆₀–COOH was expected to modify the electronic structure of the composed systems. In consequence, the energy of the HOMO of the complexes was recomputed for the globally optimized systems at the DBTB3 level with solvation effects; the results ranged from -3.504 to -5.164 eV. Similarly, the energy of the LUMO varied from -3.48 to -4.437 eV. The electrophilicity computed by Koopman's theorem had variations between 21.675 and 508.086 eV. Molecular docking was performed with the CXCR7 protein to obtain the docking score, number of established hydrogen bonds, and protein residues interacting with the complex at a distance of 3 Å. The results obtained with Autodock Vina for the 19 complexes and the validation set are shown in Table 8.

The resultant docking score between drug– C_{60} –COOH and CXCR7 ranged from -11.7 to -8.8 kcal/mol. As in the previous case, the water-soluble fullerene increased the docking score compared with the isolated drugs (Table 2). Modified ixabepilone had the highest docking score, and the capecitabine had the lowest. The numbers of hydrogen bonds were obtained in a narrow range from 0 to 3.

Table 8: Docking score, number of H-bonds established after docking, and interacting residues of CXCR7 with drug–fullerene C₆₀–COOH complexes at 3 Å distance.

Ligand	Score (kcal/mol)	H-bonds	Interacting residues
dinaciclib	-9.7	0	Lys342, Leu340, Phe330, Leu343, Leu326, Tyr322, Val64, Ile60, Ala61
gemcitabine	-10.6	0	Phe294, Asn36, Asp30, Leu104, Arg197. Trp100, Tyr195, Ser190, Asn108, Lys40, Leu43
voruciclib	-10.3	0	Lys342, Leu343, Leu340, Val64, Leu326, Ser316, Val313, Ala61, Ile60, Ile57, Val56
fluorouracil	-11.4	1	Lys342, Leu343, Leu340, Thr341, Phe330, Leu326, Tyr322, Ser316, Val64, lle60
olaparib	-11.6	0	Lys342, Leu343, Leu340, Phe330, Val64, Leu326, Tyr322, Arg323, Ser316, Ile60, Ala61, Val313
ixabepilone	-11.7	0	Ser190, Glu193, His107, Lys40, Arg197, Ile205, Pro38, Val28, Ile27
alvocidib	-10.9	1	Lys342, Leu343, Leu340, Val64, Leu326, Tyr322, Ala61, lle60, lle57
methotrexate	-10.9	2	Asn108, Lys40, Tyr195, Ser190, Arg197, Ser198, Tyr200, Leu43, Gln301, Tyr268, His298, Phe294, Asn39, Pro38, Asn36, Met37, His291, Asp30, Val28
ribociclib	-11.2	0	Leu104, Asn36, Phe294, Met33, Val32, Arg197, Ser190, Trp110, Tyr195, Asp30, Ile205, Lys206
exemestane	-11.6	1	Lys342, Leu343, Leu340, Leu326, Phe330, Ala61, Ile60, Ile57, Val313
tamoxifen	-9.1	0	Leu340, Leu326, Phe330, Ser316, Ile60, Val313, Ala61, Ile60, Ile57, Val56
idarubicin	-10.6	0	Leu343, Leu340, Lys342, Val64, Tyr322, Pro312, Ser316, Val313, Phe330, Ile60, Ile57
palbociclib	-11.4	0	lle166, Arg162, Cys165, Cys81, Asn85, lle88, Trp92, lle70, Val66
toremifene	-9	0	Leu343, Lys342, Ser350, Leu340, Val64, Leu326, Ala61, Ser316, Ile60, Ile57
roniciclib	-9.2	0	Lys342, Leu343, Leu340, Leu326, Arg323, Val64, Ala61, Ile60, Ile57
capecitabine	-8.8	0	Thr341, Leu343, Leu340, Val64, Phe330, Leu326, Ala61, Ile60, Ile57, Val313
sulfanilamide	-10.7	1	Lys342, Leu343, Thr341, Leu340, Leu326, Tyr322, Ser316, Pro312, Val64, Ala61, Ile60
zoledronic acid hydrate	-10	0	Lys342, Leu343, Val64, Val65, Leu326, Tyr322, Ser316, Val313, Pro312, Ala61, Ile57
pamidronic acid	-10.6	3	Leu122, Ile126, Ile123, Trp169, Trp92, Ile166, Asn85, Cys165
topotecan*	-12.6	2	Thr341, Phe330, Leu340, Leu343, Leu326, Val64, Tyr322, Ser316, Val65, Pro312, Val313, Ala61, Ile60, Ile57
tucatinib*	-10.9	1	Leu343, Lys342, Leu340, Thr341, Val64, Tyr322, Leu326, Ser316, Val313, Ile60, Ala61, Val56, Ile57
pemetrexed*	-9.9	1	lle166, Arg162, Trp92, lle88, Leu84, Asn85, Thr75, Cys81
thiotepa*	-9.4	0	Lys342, Leu343, Leu340, Thr341, Val64, Leu326, Phe330, Ile60

Considering the residues shown in Table 8 for the drug-fullerene complex with the protein, there are three possible binding sites. The first one is located inside the protein, near the pocket determined for isolated drugs. This binding site is near phenylalanine Phe294 and arginine Arg197 (Figure 4a) as prominent residues. The second possible binding site is outside the protein, near where drug-C₆₀ binds; common residues are serine Ser316 and valine Val313 (Figure 4b). The third binding site is also at the outside of the protein and characterized by isoleucine Ile166 and arginine Arg162 (Figure 4c).

Table 9 shows the quantitative descriptors used and the importance that each one has in the predictive models for drug– C_{60} –COOH. The docking score was the predicted variable in all the cases. In addition, the variables used to obtain the models in the case of drug– C_{60} were initially considered. The variables to model the interaction with C_{60} –COOH were taken directly from those of drug– C_{60} . For example, AI model 1 with C_{60} –COOH used the variables that had an importance greater than zero from AI model 1 of drug– C_{60} . The most important variables in the six models were LogP, RBC, and Dn. The least important variable in the six models was WS because had less than 10% importance in all models.

Supporting Information File 1, Table S6 shows the docking scores obtained for the validation set using both methods, that

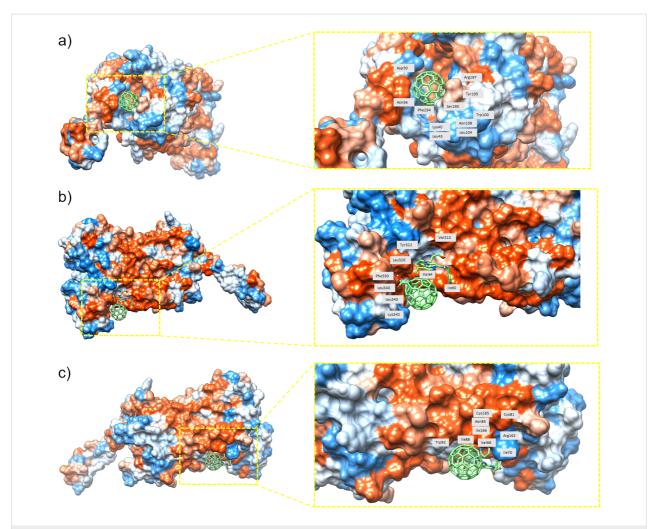


Figure 4: Binding sites for (a) gemcitabine, (b) dinaciclib, and (c) palbociclib with C_{60} –COOH and protein CXCR7. Blue color indicates the most hydrophilic sites and orange-red the most hydrophobic ones. Interacting residues are labeled as well.

 Table 9: Input variables (IV) and output importance (OI) obtained for six Extra Tree regressor models obtained from IBM Watson. Variables are annotated according to the notation introduced in Table 1. The best models, according to their MAPE values, are highlighted in bold.

Mo	odel 1	Mo	del 2	Mo	del 3	Mo	del 4	Mo	odel 5	Mo	odel 6
IV	OI (%)	IV	OI (%)	IV	OI (%)	IV	OI (%)	IV	OI (%)	IV	OI (%)
RBC	100	pK _a	100	PSA	100	Ω	100	LogP	100	MW	100
LogP	34	LogP	96	LogP	77	р K_a	93	LogS	90	NOR	96
Dn	20	Dn	69	pK_a	72	PSA	70	α	83	E_{LUMO}	38
WS	10	LogS	64	Α	42	E_{LUMO}	31	NOR	61	ω	0
LogS	6	E_{HOMO}	30	E_{HOMO}	32	NOR	5	Dn	52	_	_
NOR	5	PSA	12	E_{LUMO}	26	E_{HOMO}	2	E_{LUMO}	0	_	_
ω	2	E_{LUMO}	0	WS	3	LogS	1	_	_	_	_
MW	1	_	_	ω	0	WS	0	_	_	_	_
Еномо	0	_	_	_	_	_	_	_	_	_	_
PSA	0	_	_	_	_	_	_	_	_	_	_
α	0	_	_	_	_	_	_	_	_	_	_

is, the Extra Tree regressors implemented in IBM Watson and the MLR. With these values, it is possible to compare the difference in the docking score obtained by molecular docking and the prediction of the mathematical models for drug-C₆₀-COOH and the protein CXCR7. Table 10 shows the values obtained for the different types of errors in each of the models. The MSE ranged from 0.77 to 4.73 kcal²/mol², whereas the MAPE varied from 6.70 to 16.22%. Also, the MAE was obtained ranging from 0.69 to 6.52 kcal/mol. Finally, the RMSE varied from 0.88 to 2.18 kcal/mol. Considering the MAPE, the best model, with a value of 6.7%, to predict the docking score is MLR model 1. Once again, the synergistic effect of using AI with a mathematical tool such as MLR is observed. The benefits are the predictive model's clarity, whereas AI was useful in determining the most important descriptors to be included in the QSAR/QSPR model. The explicit form of this model is:

$$\begin{split} \text{MLR1}_{\text{DRUG+C60-COOH}} &= -13.597 + 0.047 \times \text{pK}_{\text{a}} + 0.33 \times \text{LogP} \\ &+ 0.18 \times \text{Dn} + 0.284 \times \text{LogS} \\ &- 0.696 \times E_{\text{HOMO}} - 0.007 \times \text{PSA}. \end{split} \tag{7}$$

The other significant model is MLR model 5 with a MAPE of 10.17%. In this model, experimental and theoretical descriptors were mixed. The following is the explicit form of MLR model 5:

$$\begin{aligned} \text{MLR5}_{\text{DRUG+C60-COOH}} &= 2.480 + 0.552 \times \text{LogP} \\ &+ 0.21 \times \text{LogS} + 0.002 \times \alpha \\ &- 0.402 \times \text{NOR} + 0.186 \times \text{Dn}. \end{aligned} \tag{8}$$

Considering the MAPE, the best AI model is model 4 with a value of 8.18% and the functional form AI4 $_{DRUG+C60-COOH} = f$ (E_{HOMO} , ω , p K_a , PSA, E_{LUMO} , LogS, NOR). The other significant model obtained from AI is model 5, with a MAPE value of 8.69%; the functional form of this model is AI5 $_{DRUG+C60-COOH} = f$ (LogP, α , LogS, NOR, Dn). Thus, although the Extra Trees algorithm was competitive in the case of drugs modified with a

carboxyfullerene, this approach was surpassed by the MLR with the AI choosing the most important variables.

Doxorubicin and gemcitabine with a watersoluble fullerene

Finally, doxorubicin and gemcitabine were selected to compare the DFTB3 approach with the regular DFT method. In addition, their interactions with a water-soluble fullerene derivative were studied as well. Both anticancer agents are presented in Figure 5 interacting with a water-soluble fullerene [36,37]. Doxorubicin, an antibiotic that belongs to the family of tetracycline pharmaceutical agents, has gained popularity among chemotherapy agents and was recently modified with fullerene C_{60} [72-75]. Its anticarcinogenic activity comes from its ability to intercalate into DNA, inducing damage of the DNA strands and inhibiting its replication. Also, doxorubicin contributes to stopping the action of the enzyme topoisomerase II, leading to apoptosis of living tissues [71]; therefore, it is important to study its intrinsic chemical reactivity.

At the B3PW91/6-31G level of DFT, it is possible to appreciate that the periphery of the doxorubicin molecule is saturated by organic substituents. Its oxy, carbonyl, and carboxy terminal groups are active sites to interact with DNA or amino acids. The molecular orbital scheme of this molecule is shown in Supporting Information File 1, Figure S2, together with its molecular electrostatic potential (ESP). In the case of doxorubicin, from frontier molecular orbitals theory, HOMO and LUMO were found on the tetracycline moiety. However, the HOMO is confined to the quinoid ring, whereas the LUMO is completely delocalized (Supporting Information File 1, Figure S2). Oxygen atoms are the regions with the most negative ESP (red color in Supporting Information File 1, Figure S2). In contrast, a high electrostatic potential (blue color in Supporting Information File 1, Figure S2) is found on the lateral substituted cycle. Both central rings, the aromatic one and the quinoid one, are the main regions for the reactivity, including all the substituent oxygen atoms. The frontier molecular orbitals are similar, and it is ex-

Table 10: Metrics obtained by the use of AI and MLR in the case of drug-C₆₀-COOH. The best models, according to their MAPE values, are highlighted in bold. Values were computed relative to the validation set.

Error	Model	1	Model	2	Model	3	Model	4	Model	5	Model 6	6
	Al	MLR	Al	MLR	Al	MLR	AI	MLR	ΑI	MLR	Al	MLR
MSE (kcal²/mol²)	0.97	0.77	1.67	1.77	1.80	4.73	1.05	1.53	1.41	2.08	1.46	1.80
MAPE (%)	8.75	6.70	10.51	12.03	10.71	16.22	8.18	10.26	8.69	10.17	10.86	11.39
MAE (kcal/mol)	0.93	0.69	1.13	1.29	1.17	6.52	0.86	1.10	0.93	0.99	1.16	1.18
RMSE (kcal/mol)	0.98	0.88	1.29	1.33	1.34	2.18	1.02	1.24	1.19	1.44	1.21	1.34

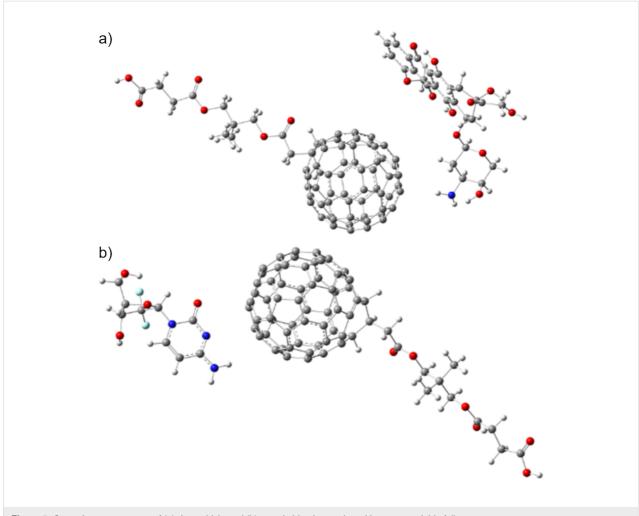


Figure 5: Ground state structures of (a) doxorubicin and (b) gemcitabine interacting with a water-soluble fullerene.

pected that electronic transit can occur in this region accepting and donating negative charges. The ESP map reinforces this suggestion showing negative density sites as well as a positive center, which can receive electrons. The energy of the HOMO was computed as -5.978 eV and that of the LUMO as -4.221 eV at the DFTB3 level. In comparison, the B3PW91 method yielded -6.116 and -3.242 eV, respectively. Thus, to consider the models obtained here, it is recommended to use DFTB3 to compute the electronic and energetic properties instead of DFT calculations.

Gemcitabine includes an active pyrimidinone fragment as a very reactive zone. The frontier molecular orbitals and the ESP map are shown Supporting Information File 1, Figure S2. Again, there is a strong polarization, which can induce a route for reaction. The ring nitrogen atom in alpha position concerning the carbonyl group is the more nucleophilic center, whereas there are two positive-density regions near the carbonyl group and in the C–C bond next to the amine-substituted carbon atom.

Both pharmaceutical agents are susceptible to interaction with fullerenes to form a force dispersion complex as it has been previously suggested. However, these complexes should be watersoluble to be delivered to their host. Considering all these factors, a water-soluble species was used to form such complexes, the structure of which [76] is shown in Figure 5. In both cases, a strong hydrogen bond is present; the distances are 1.97 Å for the doxorubicin complex and 2.25 Å for the gemcitabine complex. Furthermore, the energy of these interactions was calculated taking advantage of the Grimme module; they are 23.7 kcal/mol for doxorubicin and 18.9 kcal/mol for gemcitabine.

Conclusion

A QSAR/QSPR study of drugs commonly used for breast cancer chemotherapy modified with fullerene derivatives as drug nanocarriers was carried out. The CXCR7 protein was selected as a target for molecular docking calculations; the drugs were studied in the isolated form and modified with C_{60}

fullerene and with the water-soluble C₆₀-COOH fullerene derivative. An initial dataset was built by analyzing more than 30 drugs. The models to predict the docking score were obtained concerning Pearson's HSAB concept and common QSAR/ QSPR descriptors. The energetic descriptors were computed quantum chemically by using density functional-based tight binding at the DFTB3 level. The highest docking score in the case of isolated drugs was -10.1 kcal/mol for olaparib. In contrast, in the case of the drugs modified with pristine C₆₀ fullerene, it was -13.6 kcal/mol for exemestane. In the case of the drugs modified with the water-soluble fullerene derivative C₆₀-COOH, the maximum docking score was -11.7 kcal/mol for ixabepilone. Hence, the complexes are supposed to dock with stronger interactions with the CXCR7 protein than the isolated drugs. Also, characteristic binding sites were determined. The pocket of the isolated drugs was found within the protein, sharing residues including Trp100, Leu297, and Ser198. In the case of the drugs with fullerene C₆₀, the binding site was outside the protein with the complex pointing away from the pocket. The interacting residues included Arg323, Ser316, and Lys342. In the case of the drugs with C_{60} -COOH fullerene, there were three possible binding sites. The first two are the same as those in the previous cases The third binding site was found outside the protein, near the residues Ile166, Arg192, and Cys165. The docking score for the drug-fullerene complex is higher than that of the isolated drugs. QSAR/QSPR predictive models for the docking score were obtained from MLR and from IBM Watson artificial intelligence, yielding models with a MAPE of lower than 12% in all three cases. Although MLR exhibits the best evaluation metrics in the case of drug-C₆₀ and drug-C₆₀-COOH complexes, an improvement is obtained based on the variables detected by the AI as the most important ones.

Supporting Information

Supporting Information File 1

Additional tables and figures.

[https://www.beilstein-journals.org/bjnano/content/supplementary/2190-4286-15-95-S1.pdf]

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Data Availability Statement

The data that supports the findings of this study is available from the corresponding author upon reasonable request.

References

- Giaquinto, A. N.; Miller, K. D.; Tossas, K. Y.; Winn, R. A.; Jemal, A.; Siegel, R. L. *Ca-Cancer J. Clin.* **2022**, *72*, 202–229. doi:10.3322/caac.21718
- Miller, K. D.; Ortiz, A. P.; Pinheiro, P. S.; Bandi, P.; Minihan, A.; Fuchs, H. E.; Martinez Tyson, D.; Tortolero-Luna, G.; Fedewa, S. A.; Jemal, A. M.; Siegel, R. L. Ca-Cancer J. Clin. 2021, 71, 466–487. doi:10.3322/caac.21695
- Wendt, C.; Margolin, S. Acta Oncol. 2019, 58, 135–146. doi:10.1080/0284186x.2018.1529428
- Faruk, T.; Islam, M. K.; Arefin, S.; Haq, M. Z. Clin. Breast Cancer 2015, 15, 313–324. doi:10.1016/j.clbc.2015.01.002
- Giaquinto, A. N.; Sung, H.; Miller, K. D.; Kramer, J. L.; Newman, L. A.; Minihan, A.; Jemal, A.; Siegel, R. L. Ca-Cancer J. Clin. 2022, 72, 524–541. doi:10.3322/caac.21754
- Siegel, R. L.; Miller, K. D.; Wagle, N. S.; Jemal, A. Ca-Cancer J. Clin. 2023, 73, 17–48. doi:10.3322/caac.21763
- Sung, H.; Ferlay, J.; Siegel, R. L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Ca-Cancer J. Clin. 2021, 71, 209–249. doi:10.3322/caac.21660

- Arnold, M.; Morgan, E.; Rumgay, H.; Mafra, A.; Singh, D.; Laversanne, M.; Vignat, J.; Gralow, J. R.; Cardoso, F.; Siesling, S.; Soerjomataram, I. *Breast* 2022, 66, 15–23. doi:10.1016/j.breast.2022.08.010
- Anderson, D. Mutat. Res., Fundam. Mol. Mech. Mutagen. 1995, 329, 37–47. doi:10.1016/0027-5107(95)00017-d
- Hay, J.; Shahzeidi, S.; Laurent, G. Arch. Toxicol. 1991, 65, 81–94. doi:10.1007/bf02034932
- Meyer, K. B.; Madias, N. E. *Miner Electrolyte Metab.* 1994, 20, 201–213.
- Bentzen, S. M. Nat. Rev. Cancer 2006, 6, 702–713. doi:10.1038/nrc1950
- Delanian, S.; Porcher, R.; Rudant, J.; Lefaix, J.-L. J. Clin. Oncol. 2005, 23, 8570–8579. doi:10.1200/jco.2005.02.4729
- Dunnwald, L. K.; Rossing, M. A.; Li, C. I. Breast Cancer Res. 2007, 9, R6. doi:10.1186/bcr1639
- Asif, H. M.; Sultana, S.; Ahmed, S.; Akhtar, N.; Tariq, M. Asian Pac. J. Cancer Prev. 2016, 17, 1609–1615. doi:10.7314/apjcp.2016.17.4.1609
- Shi, Y.; Riese, D. J., II; Shen, J. Front. Pharmacol. 2020, 11, 574667. doi:10.3389/fphar.2020.574667
- Sherif, M. F.; Ismail, I. M.; Ata, S. M. S. Ann. Diagn. Pathol. 2020, 49, 151621. doi:10.1016/j.anndiagpath.2020.151621
- 18. Xu, L.; Li, C.; Hua, F.; Liu, X. *Med. Oncol.* **2021,** *38*, 58. doi:10.1007/s12032-021-01481-2
- Ding, C.; Li, Z. Mater. Sci. Eng., C 2017, 76, 1440–1453. doi:10.1016/j.msec.2017.03.130
- 20. Hossen, S.; Hossain, M. K.; Basher, M. K.; Mia, M. N. H.; Rahman, M. T.; Uddin, M. J. J. Adv. Res. 2019, 15, 1–18. doi:10.1016/j.jare.2018.06.005
- 21. Krätschmer, W.; Lamb, L. D.; Fostiropoulos, K.; Huffman, D. R. *Nature* **1990**, *347*, 354–358. doi:10.1038/347354a0
- Krusic, P. J.; Wasserman, E.; Keizer, P. N.; Morton, J. R.; Preston, K. F. Science 1991, 254, 1183–1185. doi:10.1126/science.254.5035.1183
- Markovic, Z.; Trajkovic, V. Biomaterials 2008, 29, 3561–3573. doi:10.1016/j.biomaterials.2008.05.005
- Castro, E.; Garcia, A. H.; Zavala, G.; Echegoyen, L. J. Mater. Chem. B 2017, 5, 6523–6535. doi:10.1039/c7tb00855d
- 25. Lichota, A.; Krokosz, A. *Med. Pr.* **2016**, *67*, 817–831. doi:10.13075/mp.5893.00466
- Krishna, V.; Singh, A.; Sharma, P.; Iwakuma, N.; Wang, Q.; Zhang, Q.; Knapik, J.; Jiang, H.; Grobmyer, S. R.; Koopman, B.; Moudgil, B. Small 2010, 6, 2236–2241. doi:10.1002/smll.201000847
- 27. Kumar, M.; Raza, K. *Pharm. Nanotechnol.* **2018**, 5. doi:10.2174/2211738505666170301142232
- 28. Panova, G. G.; Zhuravleva, A. S.; Khomyakov, Y. V.; Vertebnyi, V. E.; Ageev, S. V.; Petrov, A. V.; Podolsky, N. E.; Morozova, E. I.; Sharoyko, V. V.; Semenov, K. N. J. Mol. Struct. 2021, 1235, 130163. doi:10.1016/j.molstruc.2021.130163
- Ye, S.; Zhou, T.; Cheng, K.; Chen, M.; Wang, Y.; Jiang, Y.; Yang, P. Nanoscale Res. Lett. 2015, 10, 246. doi:10.1186/s11671-015-0953-9
- Danishuddin; Khan, A. U. *Drug Discovery Today* 2016, 21, 1291–1302. doi:10.1016/j.drudis.2016.06.013
- 31. Chen, X.; Li, H.; Tian, L.; Li, Q.; Luo, J.; Zhang, Y. *J. Comput. Biol.* **2020**, *27*, 1397–1406. doi:10.1089/cmb.2019.0323
- 32. Pearson, R. G. J. Am. Chem. Soc. 1963, 85, 3533–3539. doi:10.1021/ja00905a001

- Chattaraj, P. K.; Giri, S.
 Annu. Rep. Prog. Chem., Sect. C: Phys. Chem. 2009, 105, 13. doi:10.1039/b802832i
- Bağlayan, Ö.; Parlak, C.; Dikmen, G.; Alver, Ö. Comput. Theor. Chem.
 1221, 114036. doi:10.1016/j.comptc.2023.114036
- Parlak, C.; Alver, Ö. J. Indian Chem. Soc. 2022, 99, 100769. doi:10.1016/j.jics.2022.100769
- 36. Pérez-Manríquez, L.; Ramos, E.; Rangel, E.; Salcedo, R. Mol. Simul. **2013.** *39.* 612–620. doi:10.1080/08927022.2012.758845
- Bosquez, M.; Fomina, L.; Salcedo, R. Mol. Simul. 2020, 46, 398–407. doi:10.1080/08927022.2020.1714042
- 38. Muratov, E. N.; Bajorath, J.; Sheridan, R. P.; Tetko, I. V.; Filimonov, D.; Poroikov, V.; Oprea, T. I.; Baskin, I. I.; Varnek, A.; Roitberg, A.; Isayev, O.; Curtalolo, S.; Fourches, D.; Cohen, Y.; Aspuru-Guzik, A.; Winkler, D. A.; Agrafiotis, D.; Cherkasov, A.; Tropsha, A. Chem. Soc. Rev. 2020, 49, 3525–3564. doi:10.1039/d0cs00098a
- Selassie, C.; Verma, R. P. History of Quantitative Structure–Activity Relationships. In *Burger's Medicinal Chemistry and Drug Discovery*; Abraham, D. J., Ed.; Wiley, 2010; pp 1–96. doi:10.1002/0471266949.bmc001.pub2
- 40. Hourahine, B.; Aradi, B.; Blum, V.; Bonafé, F.; Buccheri, A.; Camacho, C.; Cevallos, C.; Deshaye, M. Y.; Dumitrică, T.; Dominguez, A.; Ehlert, S.; Elstner, M.; Van Der Heide, T.; Hermann, J.; Irle, S.; Kranz, J. J.; Köhler, C.; Kowalczyk, T.; Kubař, T.; Lee, I. S.; Lutsker, V.; Maurer, R. J.; Min, S. K.; Mitchell, I.; Negre, C.; Niehaus, T. A.; Niklasson, A. M. N.; Page, A. J.; Pecchia, A.; Penazzi, G.; Persson, M. P.; Řezáč, J.; Sánchez, C. G.; Sternberg, M.; Stöhr, M.; Stuckenberg, F.; Tkatchenko, A.; Yu, V. W.-z.; Frauenheim, T. J. Chem. Phys. 2020, 152, 124101. doi:10.1063/1.5143190
- Gaus, M.; Goez, A.; Elstner, M. J. Chem. Theory Comput. 2013, 9, 338–354. doi:10.1021/ct300849w
- 42. Vainio, M. J.; Johnson, M. S. *J. Chem. Inf. Model.* **2007**, *47*, 2462–2474. doi:10.1021/ci6005646
- Wang, C.; Chen, W.; Shen, J. Front. Pharmacol. 2018, 9, 10.3389/fphar.2018.00641. doi:10.3389/fphar.2018.00641
- Yang, J.; Zhang, Y. Nucleic Acids Res. 2015, 43, W174–W181. doi:10.1093/nar/gkv342
- Muthiah, I.; Rajendran, K.; Dhanaraj, P.; Vallinayagam, S. J. Biomol. Struct. Dyn. 2021, 39, 4807–4815. doi:10.1080/07391102.2020.1783365
- Laskowski, R. A.; MacArthur, M. W.; Moss, D. S.; Thornton, J. M. J. Appl. Crystallogr. 1993, 26, 283–291. doi:10.1107/s0021889892009944
- Eberhardt, J.; Santos-Martins, D.; Tillack, A. F.; Forli, S. J. Chem. Inf. Model. 2021, 61, 3891–3898. doi:10.1021/acs.jcim.1c00203
- Ganapathiraju, M.; Balakrishnan, N.; Reddy, R.; Klein-Seetharaman, J. BMC Bioinf. 2008, 9. S4. doi:10.1186/1471-2105-9-s1-s4
- Tusnády, G. E.; Simon, I. Bioinformatics 2001, 17, 849–850. doi:10.1093/bioinformatics/17.9.849
- Chen, Y.; Elenee Argentinis, J.; Weber, G. Clin. Ther. 2016, 38, 688–701. doi:10.1016/j.clinthera.2015.12.001
- Geurts, P.; Ernst, D.; Wehenkel, L. Mach. Learn. 2006, 63, 3–42. doi:10.1007/s10994-006-6226-1
- Suzuki, J. Statistical Learning with Math and Python: 100 Exercises for Building Logic; Springer Singapore: Singapore, 2021. doi:10.1007/978-981-15-7877-9
- Costa, A.; Nannicini, G. Math. Program. Comput. 2018, 10, 597–629. doi:10.1007/s12532-018-0144-7

- 54. Methods in molecular biology. In *Topics in Biostatistics;* Ambrosius, W. T., Ed.; Humana Press: Totowa, N.J., 2007. doi:10.1007/978-1-59745-530-5
- Yang, L.; Sang, C.; Wang, Y.; Liu, W.; Hao, W.; Chang, J.; Li, J. *Chemosphere* 2021, 285, 131456. doi:10.1016/j.chemosphere.2021.131456
- Keshavarz, M. H.; Shirazi, Z.; Kiani Sheikhabadi, P. *Process Saf. Environ. Prot.* 2021, 150, 137–147. doi:10.1016/j.psep.2021.04.011
- Lomba, L.; Ribate, M. P.; Zuriaga, E.; García, C. B.; Giner, B. *Ecotoxicol. Environ. Saf.* 2019, 172, 232–239. doi:10.1016/j.ecoenv.2019.01.081
- Sadeghi, F.; Afkhami, A.; Madrakian, T.; Ghavami, R. J. Iran. Chem. Soc. 2021, 18, 2785–2800. doi:10.1007/s13738-021-02233-9
- 59. Zuriaga, E.; Giner, B.; Valero, M. S.; Gómez, M.; García, C. B.; Lomba, L. Chemosphere 2019, 227, 480–488. doi:10.1016/j.chemosphere.2019.04.054
- Fadilah, F.; Arsianti, A.; Yanuar, A.; Andrajati, R.; Indah Paramita, R.; Hernawati Purwaningsih, E. *Orient. J. Chem.* 2018, 34, 2656–2660. doi:10.13005/ojc/340558
- El Rhabori, S.; El Aissouq, A.; Chtita, S.; Khalil, F. J. Indian Chem. Soc.
 2022, 99, 100675. doi:10.1016/j.jics.2022.100675
- Er-Rajy, M.; ElFadili, M.; Mrabti, N. N.; Zarougui, S.; Elhallaoui, M. Chin. J. Anal. Chem. 2022, 50, 100163. doi:10.1016/j.cjac.2022.100163
- Silva, A. M.; Martins-Gomes, C.; Silva, T. L.; Coutinho, T. E.; Souto, E. B.; Andreani, T. *Toxics* 2022, 10, 378. doi:10.3390/toxics10070378
- Hansch, C.; Steinmetz, W. E.; Leo, A. J.; Mekapati, S. B.; Kurup, A.;
 Hoekman, D. J. Chem. Inf. Comput. Sci. 2003, 43, 120–125.
 doi:10.1021/ci020378b
- Sudhakaran, S.; Calvin, J.; Amy, G. L. Chemosphere 2012, 87, 144–150. doi:10.1016/j.chemosphere.2011.12.006
- Ogunyemi, B. T.; Latona, D. F.; Adejoro, I. A. Sci. Afr. 2020, 8, e00336. doi:10.1016/j.sciaf.2020.e00336
- Bodun, D. S.; Omoboyowa, D. A.; Omotuyi, O. I.; Olugbogi, E. A.;
 Balogun, T. A.; Ezeh, C. J.; Omirin, E. S. Comput. Biol. Chem. 2023, 104, 107865. doi:10.1016/j.compbiolchem.2023.107865
- Trott, O.; Olson, A. J. J. Comput. Chem. 2010, 31, 455–461. doi:10.1002/jcc.21334
- Bos, J. D.; Meinardi, M. M. H. M. Exp. Dermatol. 2000, 9, 165–169. doi:10.1034/j.1600-0625.2000.009003165.x
- Haddon, R. C. Philos. Trans. R. Soc., A 1993, 343, 53–62. doi:10.1098/rsta.1993.0040
- Sergio, M.; Behzadi, H.; Otto, A.; van der Spoel, D. *Environ. Chem. Lett.* 2013, 11, 105–118. doi:10.1007/s10311-012-0387-x
- Panchuk, R. R.; Prylutska, S. V.; Chumak, V. V.; Skorokhyd, N. R.; Lehka, L. V.; Evstigneev, M. P.; Prylutskyy, Y. I.; Berger, W.; Heffeter, P.; Scharff, P.; Ritter, U.; Stoika, R. S. J. Biomed. Nanotechnol. 2015, 11, 1139–1152. doi:10.1166/jbn.2015.2058
- Butowska, K.; Kozak, W.; Zdrowowicz, M.; Makurat, S.; Rychłowski, M.; Hać, A.; Herman-Antosiewicz, A.; Piosik, J.; Rak, J. Struct. Chem.
 2019, 30, 2327–2338. doi:10.1007/s11224-019-01428-4
- 74. Grebinyk, A.; Prylutska, S.; Grebinyk, S.; Prylutskyy, Y.; Ritter, U.; Matyshevska, O.; Dandekar, T.; Frohme, M. Nanoscale Res. Lett. 2019, 14, 61. doi:10.1186/s11671-019-2894-1

- Liu, J.-H.; Cao, L.; Luo, P. G.; Yang, S.-T.; Lu, F.; Wang, H.;
 Meziani, M. J.; Haque, S. A.; Liu, Y.; Lacher, S.; Sun, Y.-P.
 ACS Appl. Mater. Interfaces 2010, 2, 1384–1389.
 doi:10.1021/am100037v
- 76. Martín, N.; Altable, M.; Filippone, S.; Martín-Domenech, A. *Synlett* **2007**, 3077–3095. doi:10.1055/s-2007-990939

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Heterogeneous reactions in a HFCVD reactor: simulation using a 2D model

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Abstract

In this study, a simulation of the elementary chemical reactions during SiO_x film growth in a hot filament chemical vapor deposition (HFCVD) reactor was carried out using a 2D model. For the 2D simulation, the continuity, momentum, heat, and diffusion equations were solved numerically by the software COMSOL Multiphysics based on the finite element method. The model allowed for the simulation of the key parameters of the HFCVD reactor. Also, a thermochemical study of the heterogeneous reaction between the precursors quartz and hydrogen was carried out. The obtained equilibrium constants (K_{eq}) were related to the temperature profile in the deposition zone and used in the proposed simulation. The validation of the model was carried out by measuring the temperature experimentally, where the temperature range on the substrate is 450 to 500 °C for different deposition parameters. In the simulation, the laminar flow of species contributing to the film growth was confirmed, and the simulated concentration profiles of H° and SiO near the filaments and the sources were as expected. H° and SiO are essential species for the subsequent growth of the SiO_x films. These SiO_x films have interesting properties and embedded nanostructures, which make them excellent dielectric, optoelectronic, and electroacoustic materials for the fabrication of devices compatible with silicon-based technology.

Introduction

The growth of materials such as non-stoichiometric silicon oxide (SiO_x) is an important step in semiconductor devices development. Control of deposition parameters determines the success of the process and the properties of the films, with the most important parameters being substrate temperature, gas pressure, species concentration, and flow velocity [1]. The structural, optical, and electrical properties of the SiO_x , more generally known as silicon-rich oxide (SRO), films are determined by the ratio x = [O]/[Si], which is determined by controlling key parameters in the deposition process [2]. This ratio determines optical and electrical properties such as bandgap energy, absorption coefficient, photoluminescence, refractive index, and electrical conductivity [3]. SiO_x cannot only be obtained by different CVD techniques, but also by sputtering and ion implantation, among others [4,5]. The key parameters are particular for each of these techniques. Hot filament chemical vapor deposition (HFCVD) is an excellent alternative for obtaining SRO films. It is also very versatile and economical because the input gases and materials are accessible; also, it is scalable to larger areas [6].

The SiO_x films obtained by HFCVD possess excellent optical and electrical properties, which makes such films suitable for applications in the manufacture of metal–insulator–semiconductor and metal–insulator–metal devices exhibiting blue and white electroluminescence [7,8]. It was found that these films exhibit photoconductive and photoelectric effects suitable for electroluminescence and photovoltaics applications [9,10], as well as for other applications such as solar cells and anodes for Li batteries [11]. The basic steps of the general CVD process are classified and described in [12].

The optimization of this technique improves the properties of the films; however, the complexity of the CVD processes makes it difficult to understand clearly the different mechanisms involved in such optimization. Different tools such as ab initio density functional theory [13,14], kinetic Monte Carlo simulations [15,16], and reactive molecular dynamics simulations [17,18] have been used to understand the chemical reactions underlying the growth of the films [19].

Modeling the reaction mechanism in both two-dimensional (2D) and three-dimensional (3D) systems is a tool that allows us to understand the key steps regarding the reproducibility and uniformity of the films [19,20]. From a computational point of view, prior works focused on the growth mechanism of SiO_x in a plasma-enhanced chemical vapor deposition reactor in zero dimensions (0D); the model used was solved using CHEMKIN III and AURORA software. In the model, a set of reactions was established that approximately describe the mechanisms of the

material growth, and the model results were compared with those obtained by experimental measurements [21]. Also, modeling of CVD microreactors at atmospheric pressure using tetraethyl orthosilicate as a source to obtain SiO₂ has been achieved through computational fluid dynamics (CFD) simulations [22]. The gas-phase and surface reactions were analyzed using direct Monte Carlo simulations of a hot wire chemical vapor deposition reactor for the growth of polycrystalline SiO₂ [23]. Most of these models describe CVD reactors at low pressure and low temperature, but there are not enough models regarding CVD systems at high temperature (>800 K) and high pressure (atmospheric pressure).

In this investigation, we focus on the simulation and analysis of key steps in a HFCVD deposition process to obtain SiO_x films by means of continuity, momentum, heat, and diffusion equations, which were solved numerically by the software COMSOL Multiphysics based on the finite element method; we also carry out a thermochemical analysis using FactSage. Some of the theoretical results are compared to experimental results. This work consists of five sections apart from this one. In Section "Experimental", there is a complete description of the HFCVD reactor and the deposition parameters. Section "Theoretical and Numerical Simulations" explains the equations for the 0D and 2D models. Also, the hypothesis used to develop this study and the methodology for the use of COMSOL and FactSage are given. In section "Results and Discussion", some theoretical results are compared with experimental ones. Further, the results obtained from the simulation are discussed regarding the profiles of temperature, gas velocity, and concentration of the species. Finally, the main conclusions of this research are expressed in section "Conclusion".

The study focuses on the convective transfer of the reactive gases to the solid source and the surface diffusion to the substrate. The main objective is to optimize the process for an HFCVD reactor and, thus, improve the quality and reproducibility of the films.

Experimental

The analyzed HFCVD system is a vertical reactor that can be divided into three zones. The first zone is the gas inlet, the second one is the reaction zone, and the third one is the gas outlet. In the first zone, molecular hydrogen (H₂) gas is pumped in through a stainless steel piping system that reaches a diffuser inside the reaction chamber. The gas gets in contact with eleven tungsten filaments from incandescent lamps, which are activated by an externally applied voltage generating a current; these filaments have a temperature of approximately 2300 K. The second zone includes the region where the chemical reac-

tions takes place. Here, molecular hydrogen, exposed to the high temperature of the filaments, dissociates to form atomic hydrogen, which reacts with the eleven solid quartz sources. A cloud or plasma is formed and finally reaches the substrate for the formation of the thin films. Finally, zone three is the exit of the gases that were not deposited in the film. The entire process is carried out under atmospheric pressure. Table 1 summarizes the values of the parameters and dimensions complementary to the experimental conditions in the reactor for the deposition of SRO films previously described and depicted below in Figure 2 [24], as well as the corresponding boundary conditions.

Theoretical and Numerical Simulations Hypothesis

Obtaining non-stoichiometric silicon oxide films in a HFCVD reactor is mainly based on two main heterogeneous reactions, which are the dissociation of atomic hydrogen and the reaction of atomic hydrogen with a solid source. The thermochemical study will allow us to obtain thermodynamic parameters of the heterogeneous reaction between H° in the gas phase and quartz to describe its behavior as a function of temperature at constant pressure. The thermodynamic data obtained and those taken from the bibliography will be used in the Arrhenius equation to calculate the thermodynamic and transport properties of the system in the gas phase in a dimensionless and temporal model. The thermodynamic and transport properties of the mixture will permit a study in the steady state considering the dimensions of the HFCVD reactor and the deposition parameters through numerically solving the equations of continuity, momentum, and heat transfer by the finite element method.

Mathematical method and equations

The complex growth of non-stoichiometric silicon oxide films in a HFCVD reactor involves different physics. For the description of the behavior of all systems, it is necessary to incorporate mathematical models that can explain the different phenomena involved. In this section, the mathematical models and the different pieces of software used in this study will be de-

scribed. Figure 1 shows a flowchart that describes the modeling and simulation process. The different mathematical models used, and their mathematical equations will be described in detail in the following sections.

The well-mixed reactor equations (0D model)

The modeling of reactions is based on the mass action law given by Equation 1:

$$aA + bB + \cdots \xrightarrow{k_j^f} xX + yY + \cdots, \tag{1}$$

where A, B,... are the reactants and X, Y,... are the products. a, b,... are the stoichiometric coefficients of the reactants, and x, y,... are the coefficients of the products. In the case of a set of reactions, the reaction rates r_j (mol·m⁻³·s⁻¹) can be described by the law of mass action given by Equation 2:

$$r_{j} = k_{j}^{f} \prod_{i \in \text{react}} c_{i}^{-\nu_{ij}} - k_{j}^{r} \prod_{i \in \text{prod}} c_{i}^{\nu_{ij}}, \qquad (2)$$

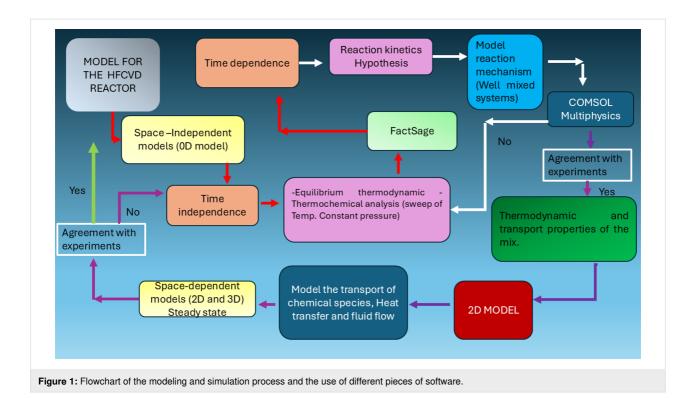
where k_j^f and k_j^r refer to the forward and reverse rate constants, respectively. The concentration of species i is indicated as c_i (mol·m⁻³). The stoichiometric coefficients are expressed by v_{ij} and are negative for reactants and positive for products [25].

The dependence of concentration and temperature on reaction rates can be included using the Arrhenius expression (Equation 3):

$$k = AT^n e^{\left(-E_{\mathcal{A}}/RT\right)},\tag{3}$$

where *A* is the frequency factor, T(K) is the absolute temperature, n is the temperature exponent, $E_A(J \cdot mol^{-1})$ is the activation energy, and R is the gas constant $(8.314 \text{ J} \cdot mol^{-1} \cdot \text{K}^{-1})$.

Table 1: Elements, dimensions, an	d parameters of the depos	ition in the HFCVD reactor.	
Parameters and dimensions o	f the HFCVD reactor	Deposition parameters and boundary cond	itions
parameter	value	parameter	value
filament diameter	0.5 [mm]	temperature of the filament (no-slip wall)	2000 [°C]
filament length	10 [mm]	pressure (outlet)	1 [atm]
distance between filaments	3.8 [mm]	mass flow (inlet)	25, 50, and 100 [sccm]
deposition time	3 [min]	substrate temperature (no-slip wall)	26 [°C]
reactor volume	0.009 [m ³]	distance source-substrate (DSS)	3, 4, and 5 [mm]
surface area of the sources	$2.19 \times 10^{-5} [\text{m}^2]$	distance filament-source	6 mm
		substrate diameter	50.8 mm



For chemical reactions in equilibrium, the equilibrium constants are defined in terms of the equilibrium expression K_{eq} by Equation 4:

$$K_{\text{eq}} = \prod_{i} c_i^{\nu_i} = \frac{\prod_{i \in \text{prod}} c_i^{\nu_i}}{\prod_{i \in \text{prod}} c_i^{-\nu_i}}.$$
 (4)

The constants K_{eq} were calculated through data obtained below from Table 2 and using Equation 5:

$$\Delta G^{\circ} = -RT \ln K_{\rm eq}, \tag{5}$$

where ΔG is the change in the Gibbs energy and R is the gas constant.

The equations in the spatial model

The used model assumes conservative species transport by diffusion and convection through a mass balance as described by Equation 6:

$$\nabla \cdot \mathbf{J}_i + \mathbf{u} \cdot \nabla c_i = R_i. \tag{6}$$

In this equation, J_i (mol·m⁻²·s⁻¹) is the diffusive flow vector, R_i (mol·m⁻³·s⁻¹) is a rate expression for the species, and \mathbf{u} (m·s⁻¹) is the mass-averaged velocity vector. The suffix i denotes the species.

The laminar behavior of the gas-phase fluid is based on the Navier–Stokes equations; for an incompressible flow, ρ = constant. The continuity equation in the general form is expressed by Equation 7, the momentum equation in the general form is given by Equation 8, and the transfer of heat in fluids is described by Equation 9:

$$\nabla \cdot (\rho \mathbf{u}) = 0, \tag{7}$$

$$\rho(\mathbf{u} \cdot \nabla)\mathbf{u} = \nabla \cdot \left[-p\mathbf{I} + \mu \left(\nabla \mathbf{u} + (\nabla \mathbf{u})^T \right) \right] + \mathbf{F}, \quad (8)$$

$$\rho C_p \left(\mathbf{u} \cdot \nabla T \right) + \nabla \cdot \left(\mathbf{q} + \mathbf{q}_r \right) = -\frac{1}{\rho} \frac{\partial \rho}{\partial T} T \left(\mathbf{u} \cdot \nabla p \right) + \tau : \nabla \mathbf{u} + Q, (9)$$

$$\boldsymbol{\tau} : \nabla \mathbf{u} = \sum_{i} \sum_{j} \left[\frac{\boldsymbol{\tau}_{ij}}{2} + \frac{1}{3} \nabla \cdot \mathbf{u} \delta_{ij} \right]^{2}, \tag{10}$$

$$\mathbf{\tau}_{ij} = -\mu \left(\frac{\partial u_i}{\partial x_j} + \frac{\partial u_j}{\partial x_i} \right),\tag{11}$$

where **u** is the velocity vector (m·s⁻¹), ρ is the density (kg·m⁻³), p is the pressure (Pa), **F** is the volume force vector (N·m⁻³), C_p is the specific heat capacity at constant pressure (J·kg⁻¹·K⁻¹), T is the absolute temperature (K), **q** is the heat flux vector

 $(\mathbf{W}\cdot\mathbf{m}^{-2})$, \mathbf{q}_r is the heat flux vector by radiation $(\mathbf{W}\cdot\mathbf{m}^{-2})$, \mathbf{I} is the identity matrix (unitless), $(\nabla \mathbf{u})^T$ is the transposed velocity gradient tensor, $\boldsymbol{\tau}$ is the viscous stress tensor (Pa), $\boldsymbol{\mu}$ is the dynamic viscosity (Pa·s), Q includes heat sources other than viscous dissipation $(\mathbf{W}\cdot\mathbf{m}^{-3})$, and δ_{ij} is the Kronecker delta symbol. All equations in this section were taken from [26], except Equation 5, which was taken from [27].

The modeling of chemical reactions

A prior thermodynamic equilibrium study of the heterogeneous reaction of SiO_2 (s) + H° (g) at 1 atm pressure in the temperature range from 100 to 2000 °C using FactSage software found that the gas species with higher concentrations were H°, SiO, OH, H₂O, and O₂; those with lower concentrations were SiH₄, SiH, Si, O, Si₂, and Si₃ [24]. This study was the basis for establishing the four main chemical reactions in zone 2 in the HFCVD reactor, which are listed below in Table 3. Here, an additional thermochemical study of the reaction SiO_2 (s) + H° (g) was developed to obtain the extensive properties of this reaction using FactSage in the temperature range from 500 to 1500 °C. Enthalpy (H), Gibbs energy (G), entropy (G), heat capacity (G_p), and Helmholtz energy (G) were obtained to calculate G_{eq} through Equation 5 [27]. The thermodynamic properties are listed in Table 2.

It should be noted that the surface film growth reactions are not considered in this study. The approximation of the thermodynamic properties of the superficial species is a study that we are still developing.

Numerical simulation

A 2D model of the HFCVD reactor was created using COMSOL Multiphysics 5.3 software [28]. Figure 2 shows the 2D geometry model with the three zones of the HFCVD reactor. For the numerical solution, the finite element method

discretizes the domain into a mesh of smaller parts called elements. The computational time for the numerical solution is related to the mesh in the 2D model with longer computational time for a greater number of elements.

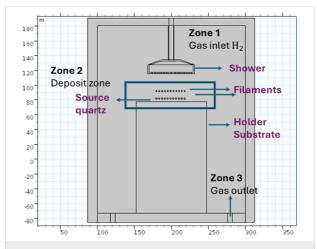


Figure 2: The HFCVD reactor zones and elements of the 2D model built in COMSOL Multiphysics 5.3.

Based on the thermodynamic equilibrium study, we propose four main reactions in zone 2, which are listed in Table 3. For the generation of atomic hydrogen, reactions 1 and 2 are considered, where M is a tertiary species or H₂. Atomic hydrogen diffuses and reacts superficially with the quartz sources; this corresponds to the process described by the heterogeneous reaction 3. Finally, reaction 4 is considered for the generation of OH, considering that the source of oxygen is the interaction of atomic H° with the sources. The Arrhenius parameters used are described in Table 3.

Initially, a well-mixed reactor approach will be used. This is a 0D model in which the reactor volume is assumed to be con-

Temperature T (°C)	Enthalpy <i>H</i> (J)	Gibbs energy <i>G</i> (J)	Entropy S (J/K)	Volume <i>V</i> (L)	Heat capacity C_p (J·K ⁻¹)	Helmholtz energy A (J)	Equilibrium constant $K_{ m eq}$
500	-677803.9	-774752.6	193.897	4.11 × 10 ¹	80.284	-778912.2	8.62784 × 10 ⁸⁰
600	-669523.0	-794911.9	208.981	4.93×10^{1}	85.279	-799902.9	1.59062 × 10 ⁶⁹
700	-660742.4	-816497.1	222.507	5.75×10^{1}	90.434	-822319.6	8.43911 × 10 ⁶⁰
800	-651404.4	-839377.8	234.967	6.57×10^{1}	96.539	-846031.8	6.37426×10^{54}
900	-645606.2	-863342.5	241.929	7.39×10^{1}	20.828	-870827.9	1.27612 × 10 ⁵⁰
1000	-643523.4	-887647.1	244.124	8.21×10^{1}	20.828	-895964.0	2.31640 × 10 ⁴⁶
1100	-641440.6	-912160.3	246.109	9.03×10^{1}	20.828	-921308.6	2.05986 × 10 ⁴³
1200	-639357.8	-936863.1	247.921	9.85×10^{1}	20.828	-946842.9	6.02018×10^{40}
1300	-637275.1	-961739.7	249.588	1.07×10^{1}	20.828	-972551.0	4.38780×10^{38}
1400	-635192.3	-986776.6	251.132	1.15 × 10 ¹	20.828	-998419.4	6.54940×10^{36}
1500	-633109.5	-1011962.0	252.569	1.2311 × 10 ¹	20.828	-1024436.7	1.73301 × 10 ³⁵

Table 3: Reactions for the 0D and 2D mode	els.			
Reaction	Frequency factor A	Temperature exponent n	Activation energy E_{A}	Ref.
1. H + H + M = H ₂ + M	1.00 × 10 ¹⁸	-1.00	0.0	[16]
2. $H + H + H_2 = H_2 + H_2$	9.2×10^{16}	-0.6	0.0	[16]
3. SiO_2 (s) + H° (g) = SiO + OH	4.0×10^{12}	0.0	5700	[17,18]
$4. H_2 + O_2 = OH + OH$	1.7×10^{13}	0	48100	[16]

stant and spatially uniform; the calculated concentrations of the species, and the temperature are instantaneous values within the reactor. The 0D model scales to a spatial 2D model. For the numerical solution, the following considerations are made: Molecular hydrogen behaves like an ideal gas in a two-dimensional model with position coordinates (x,y). The fluid is considered laminar and incompressible. The mathematical equations that describe the motion of gas are the conservation of mass (continuity equation), momentum (Navier-Stokes equations), and energy. The boundary conditions used in this model are listed in Table 1. Through the finite element method, all equations were solved via COMSOL Multiphysics. The Dirichlet boundary conditions were employed to solve this model numerically. The types of boundaries are listed in Table 1. For the boundary wall on the filaments, we use a constraints boundary condition; for the outlet, a pressure boundary condition was used. For the inlet, we use a mass flow boundary condition.

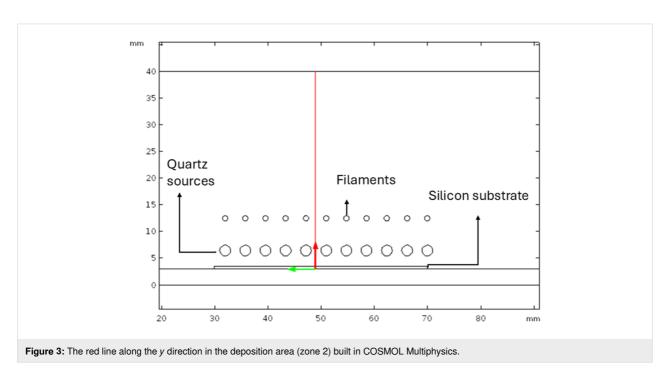
The simulations give us a broad overview on the probable reaction mechanisms during the deposition of thin films in the HFCVD reactor. This allows us to get an estimate of the types of species that will be deposited for the growth of thin films, which depend strongly on the parameters and precursors employed.

Results and Discussion Experimental results

The SiO_x films obtained using the HFCVD reactor have already been characterized using different optical, electrical, and structural characterization techniques [29-31]. Also, the process of the growth of the films was optimized through a computational fluid analysis study, which made it possible to increase the deposition area, in addition to enhancing the optoelectronic properties of the films [24].

Theoretical results

The 2D model was numerically solved in the steady state with a computing time of 286 s. For the simulation, the 2D model was analyzed with a mesh of 45,769 elements. First, the distribution of temperature and velocity of the fluid and, second, the approx-



imation of the concentration and distribution of the species are presented. All results reported in this research were obtained along a line on the y axis of the 2D model in zone 2. The line crosses all horizontal zones of the HFCVD reactor. Figure 3 shows the line in red. The data obtained on the mentioned line is presented and analyzed separately according to the main variables in the HFCVD system during deposition.

Temperature

The theoretical model was experimentally validated through temperature profiles for 20 and 30 sccm flows, which are shown in Figure 4a and Figure 4b, respectively [32]. The temperature measurements were made using a K-type thermocouple located on the surface of the deposition area. The deposition time for SiO_x was 3 min at a fixed position of the thermocouple on the substrate. Thereafter, the position of the sources was moved by 3, 4, and 5 mm. The experimental temperature measurements range from 450 to 550 °C after a deposition time of 3 min considering the mentioned DSSs.

The 2D temperature map of the HFCVD reactor is shown in Figure 5. The temperature map of deposition zone 2 is shown in Figure 6a. Figure 6b describes the temperature profile along the y coordinate from the substrate to the diffuser for the different DSSs in the steady state. The maximum temperature is near the filaments, and the minimum temperatures are found on substrate and diffuser. Heat transfer in the reactor is due to different mechanisms, such as radiation, conduction, convection, and diffusion. The theoretical and experimental temperature results were compared and analyzed near the position of the thermocouple, where the experimental temperature is only a function of the deposition time but not of the position. From the theoreti-

cal results, the temperature increases from 300 K at the substrate to 1000 K near the quartz sources, that is 3–5 mm away from the sources in the steady state.

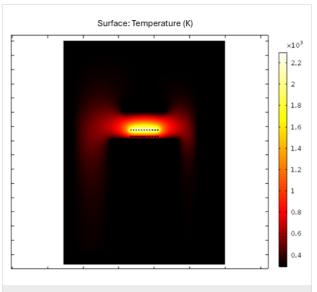
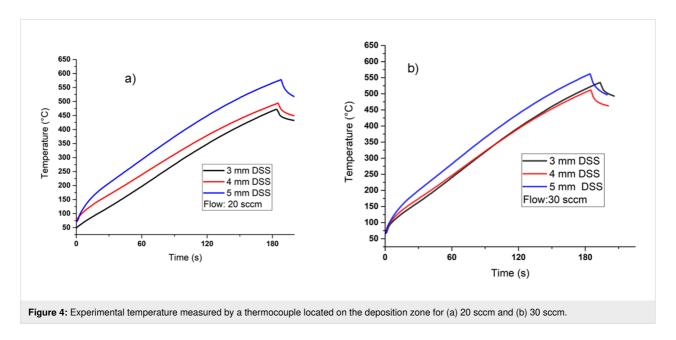


Figure 5: Temperature map of the HFCVD reactor.

The results obtained show us that the HFCVD is a high-temperature system regarding the filaments (2300 K), compared with other CVD reactors like low-pressure CVD (800 K). Hence, diffusion is expected to dominate the reactions for the formation of the precursor species. The growth of SiO_{x} films is controlled by nucleation effects. According to results for high temperatures over 700 K, the supersaturation is high and the nucleation is homogeneous in the process, leading to the precipitation of solid particles on the substrate and powder formation



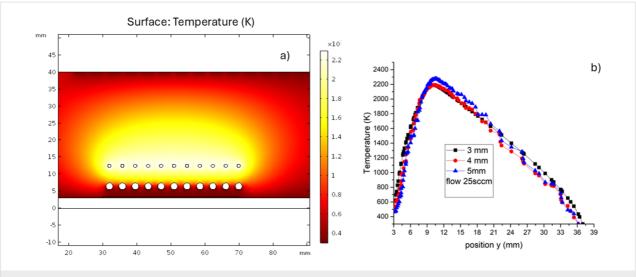
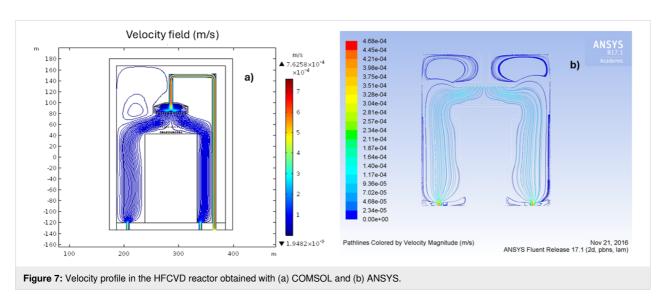


Figure 6: (a) Temperature map of the surroundings of the deposition zone. (b) Temperature profile along the y axis from the substrate to the diffuser for the different DSSs in the steady state.

[23]. SiO_x powders are obtained in the HFCVD reactor when the distance between the filaments and the source is less than 6 mm. According to what was discussed above, the distance decreases the temperature, increases the size of the clusters, and decreases the diffusion of the species, resulting in powder formation. Heterogeneous nucleation on the substrate promotes the growth of SiO_x films. The distance between the filaments and the substrate is greater than 9 mm; this reduces the temperature below 700 K according to experimental and theoretical results, promoting the growth of films. Temperature control in these systems can be modulated through the longitudinal distance between filaments, sources, and substrates. The transverse distance between sources and filaments also plays an important role in the deposition process; however, the proposed 2D model limits the study of this effect.

Velocity

Flow dynamics play a critical role when the deposition takes place at atmospheric pressure. Fluid velocity and distribution analysis were previously performed and analyzed by CFD using ANSYS Fluent [24], as depicted in Figure 7b. From this study, it was determined that it is necessary to make a change in the configuration of the reaction chamber outlets to homogenize the flow distribution and, thus, be able to optimize the deposition area to two inches. The previously reported ANSYS velocity profile is in very good agreement with that obtained in COMSOL, which is shown in Figure 7a. Both profiles in Figure 7 describe the laminar flow of the gas in zone 2 and the formation of a turbulent flow at the diffuser due to the interaction with the walls. In a study of the deposition of silicon dioxide using an atmospheric-pressure plasma-enhanced CVD



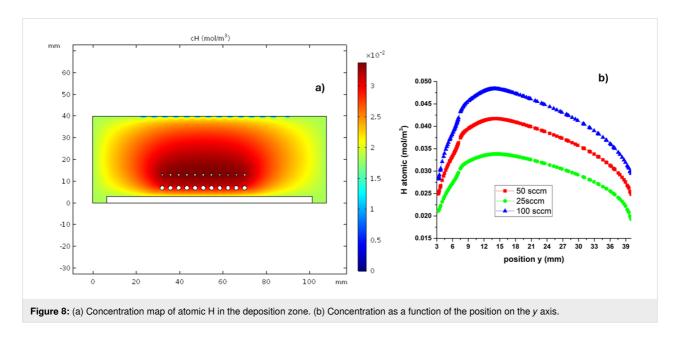
reactor, the reactor performance was shown to be strongly affected by the flow dynamics [33,34].

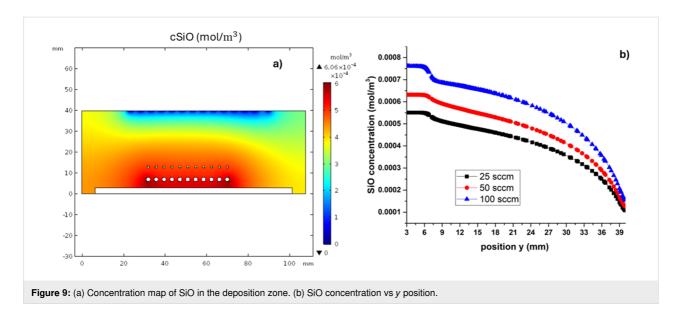
Distribution and concentration of species

As previously mentioned, temperature and fluid dynamics play a very important role in the formation and diffusion of precursor species. The solution of the model allowed us to obtain preliminary results for the concentration of the species in the reactions proposed in Table 3. The concentration profiles for H°, SiO, O2, and OH were analyzed at three different flow levels of 25, 50, and 100 sccm. The H° concentration map is shown in Figure 8a, and the concentration as a function of the y position is shown in Figure 8b. Figure 8b shows different concentration profiles for different H2 fluxes. The profiles vary slightly, and the concentration increases with the flux of H2. This effect was already studied experimentally, and the most significant results were the analysis of how the hydrogen flow influences the composition SiO_x films by X-ray photoelectron spectroscopy; as the hydrogen flow increases, the concentration of Si also increases in comparison with that of oxygen, modifying stoichiometry and bandgap [35]. The distribution of H° in zone 2 is a result of the temperature distribution. The concentration is greater near the filaments, decreasing with distance along the y axis away from the quartz sources, where the temperature varies in the range of 800-1200 K. When the distances from the filament to the source are greater, no film deposition occurs. This effect is mainly due to the recombination of H° to H2 as the temperature decreases.

The formation of these gaseous species is an important step since they contain the radical species that give rise to the surface reactions for the growth of the films. In the reaction mechanisms reported for the growth of SiO₂ films, the role of intermediate 'H and 'OH and 'O2 radicals has been pointed out through in situ Fourier-transform infrared spectroscopy. These species react with silane-derived and surface hydroxy groups, which leads to deposition. They are also responsible for the incorporation of Si-OH bonds into the silicon oxide matrix. The 'H and 'OH species attack unsaturated surface species with dangling bonds. Oxidation of surface hydrogens is the predominant reaction for O to convert H to OH. Finally, 'H and 'OH radicals can attack saturated surface species, removing H to form H₂ and H₂O, respectively, in the gas phase [21,36]. The presence of dangling bonds in the SiO_x films was detected previously. In this study, SiO_x films with thermal treatment and without thermal treatment were analyzed, and a decrease in thickness, refractive index, and excess silicon was observed. This behavior was attributed to the structural rearrangement at the atomic level by the thermal treatment due to the desorption of hydrogen from dangling bonds. The films after thermal treatment exhibited greater photoluminescence compared to those that did not undergo thermal treatment [29].

The SiO species plays an important role in the proposed mechanism; the reaction of SiO with *OH and *H radicals leads to the formation of HSiO and silanes [15]. Silane and silanol species were also found in the equilibrium thermodynamic study; they can be the precursors of nanocrystalline silicon (nc-Si) in the SiO_x films [24]. The nc-Si obtained by this technique has been studied and characterized [35]. The calculated concentration of SiO and OH is shown in Figure 9a. These species exhibit a higher concentration in the region among the quartz sources and in the vicinity of the substrate. These concentrations are gener-



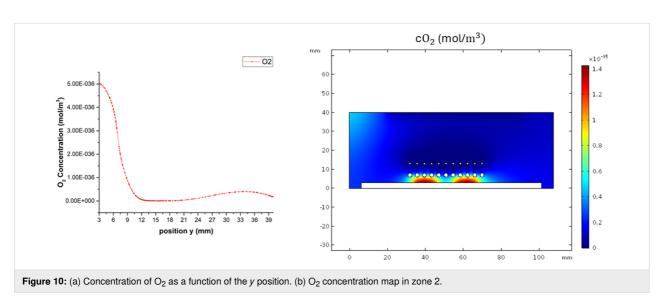


ated in chemical equilibrium by reaction 3 in Table 3. Figure 9a shows the SiO concentration map, and Figure 9b shows the concentration as a function of the *y* position for different hydrogen fluxes.

The oxygen in the deposition process is only from the solid quartz sources. In our model, there is not an additional source for O_2 . According to the proposed mechanism in reaction 4 in Table 3, the formed O_2 reacts with H_2 on the surface of the quartz sources to form OH. The O_2 content is lower than that of the other species because it reacts with H_2 and H° . Therefore, this species is predominantly found below the quartz sources. The O_2 concentration map is shown in Figure 10b. The concentration of O_2 above the sources is almost zero due to the reaction with atomic H leading to the formation of OH. Figure 10a shows the concentration of O_2 as a function of the position.

Conclusion

A model of the reaction mechanism of quartz sources with atomic hydrogen was established in thermodynamic equilibrium, and a thermochemical study of the heterogeneous chemical reaction was carried out using FactSage. We obtained the thermodynamic and transport properties to set up a 2D model. Through 2D simulation with COMSOL Multiphysics software, it was possible to obtain the concentration profiles of the main species H° , O_2 , OH, and SiO, which contribute to the growth of SRO films. In addition, the temperature and flow velocity profiles were obtained. Furthermore, it was possible to make a comparison between the theoretical results and those obtained from experiments. It was observed that the effect of the temperature on the distribution of the species is the most important. Regarding the temperature variation on the substrate, it was found that it is mainly caused by the increase in the dis-



tance between sources and substrate. The temperature on the substrate is influenced by radiation, conduction, und convection mechanisms. The transport of species is due to the convection and diffusion effects.

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Data Availability Statement

Data generated and analyzed during this study is available from the corresponding author upon reasonable request.

References

- Wang, M.; Jia, L.; Xu, H.; Li, A.; Peng, Y.; Tang, Z. Ceram. Int. 2020, 46, 4843–4849. doi:10.1016/j.ceramint.2019.10.218
- Iskhakzay, R. M. K.; Kruchinin, V. N.; Aliev, V. S.; Gritsenko, V. A.; Dementieva, E. V.; Zamoryanskaya, M. V. Russ. Microelectron. 2022, 51, 24–35. doi:10.1134/s1063739721060081
- Simón, Z. J. H.; López, J. A. L.; de la Luz, Á. D. H.; Salgado, G. G.; Leyva, K. M.; López, J. C.; Conde, G. O. M.; Lara, A. B.; Hernández, H. P. M.; Hernández, E. G.; Méndez, J. F. Surf. Interfaces 2021, 26, 101411. doi:10.1016/j.surfin.2021.101411
- Tong, L. Z. J. Phys.: Conf. Ser. 2014, 518, 012006. doi:10.1088/1742-6596/518/1/012006
- Miyazaki, H.; Goto, T. J. Non-Cryst. Solids 2006, 352, 329–333. doi:10.1016/j.jnoncrysol.2005.12.008
- Jansen, F.; Machonkin, M. A.; Kuhman, D. E. J. Vac. Sci. Technol., A 1990, 8, 3785–3790. doi:10.1116/1.576494
- Martínez, H. P.; Luna, J. A.; Morales, R.; Casco, J. F.; Hernández, J. A. D.; Luna, A.; Hernández, Z. J.; Mendoza, G.; Monfil, K.; Ramírez, R.; Carrillo, J.; Flores, J. Nanomaterials 2021, 11, 943. doi:10.3390/nano11040943
- Gao, W.; Li, T.-k.; Ono, Y.; Hsu, S.-T. J. Rare Earths 2006, 24, 673–678. doi:10.1016/s1002-0721(07)60007-6

- Koryazhkina, M. N.; Filatov, D. O.; Tikhov, S. V.; Belov, A. I.; Serov, D. A.; Kryukov, R. N.; Zubkov, S. Y.; Vorontsov, V. A.; Pavlov, D. A.; Gryaznov, E. G.; Orlova, E. S.; Shchanikov, S. A.; Mikhaylov, A. N.; Kim, S. *Nanomaterials* 2023, *13*, 2082. doi:10.3390/nano13142082
- Jambois, O.; Garrido, B.; Pellegrino, P.; Carreras, J.;
 Pérez-Rodríguez, A.; Montserrat, J.; Bonafos, C.; BenAssayag, G.;
 Schamm, S. Appl. Phys. Lett. 2006, 89, 253124.
 doi:10.1063/1.2423244
- Chen, T.; Wu, J.; Zhang, Q.; Su, X. J. Power Sources 2017, 363, 126–144. doi:10.1016/j.jpowsour.2017.07.073
- Sabzi, M.; Mousavi Anijdan, S.; Shamsodin, M.; Farzam, M.; Hojjati-Najafabadi, A.; Feng, P.; Park, N.; Lee, U. Coatings 2023, 13, 188. doi:10.3390/coatings13010188
- Hohenberg, P.; Kohn, W. Phys. Rev. 1964, 136, B864–B871. doi:10.1103/physrev.136.b864
- Kohn, W.; Sham, L. J. Phys. Rev. 1965, 140, A1133–A1138. doi:10.1103/physrev.140.a1133
- 15. Gillespie, D. T. J. Comput. Phys. **1976**, 22, 403–434. doi:10.1016/0021-9991(76)90041-3
- Voter, A. F. Introduction to the Kinetic Monte Carlo Method. In Radiation Effects in Solids; Sickafus, K. E.; Kotomin, E. A.; Uberuaga, B. P., Eds.; NATO Science Series, Vol. 235; Springer Netherlands: Dordrecht, Netherlands, 2007; pp 1–23. doi:10.1007/978-1-4020-5295-8 1
- van Duin, A. C. T.; Dasgupta, S.; Lorant, F.; Goddard, W. A.
 J. Phys. Chem. A 2001, 105, 9396–9409. doi:10.1021/jp004368u
- 18. Senftle, T. P.; Hong, S.; Islam, M. M.; Kylasa, S. B.; Zheng, Y.; Shin, Y. K.; Junkermeier, C.; Engel-Herbert, R.; Janik, M. J.; Aktulga, H. M.; Verstraelen, T.; Grama, A.; van Duin, A. C. T. npj Comput. Mater. 2016, 2, 15011. doi:10.1038/npjcompumats.2015.11
- De Wilde, J.; Lorant, C.; Descamps, P. J. Phys. D: Appl. Phys. 2017, 50, 135202. doi:10.1088/1361-6463/aa5c1d
- Wu, Y.; Zhang, H.; Yan, G.; Liu, L.; Cristea, D.; Wang, H.; Yang, Y.;
 Shen, J. *Mater. Res. Express* 2021, 8, 116403.
 doi:10.1088/2053-1591/ac3278
- Riera, M.; Rodríguez, J. A.; Barreto, J.; Domínguez, C. Thin Solid Films 2007, 515, 3380–3386. doi:10.1016/j.tsf.2006.09.032
- Konakov, S. A.; Krzhizhanovskaya, V. V. J. Phys.: Conf. Ser. 2015, 574, 012145. doi:10.1088/1742-6596/574/1/012145
- Holt, J. K.; Swiatek, M.; Goodwin, D. G.; Muller, R. P.;
 Goddard, W. A., III; Atwater, H. A. Thin Solid Films 2001, 395, 29–35.
 doi:10.1016/s0040-6090(01)01202-0
- 24. Morán Martínez, X. A.; Luna-López, J. A.; Apreza Sies, A.; Hernández-de La Luz, Á. D.; Martinez Hernández, H. P. Adv. Mater. Res. 2021, 1165, 99–111. doi:10.4028/www.scientific.net/amr.1165.99
- Zanoni, M. A. B.; Gardner, L.; Gnanapragasam, N.; Liang, Z. Int. J. Hydrogen Energy 2024, 57, 39–51.
 doi:10.1016/j.ijhydene.2023.12.192
- 26. COMSOL. Multiphysics 5.5 Reference Manual.

 https://doc.comsol.com/5.5/doc/com.comsol.help.comsol/COMSOL_ReferenceManual.pdf (accessed Nov 29, 2024).
- Bale, C. W.; Bélisle, E.; Chartrand, P.; Decterov, S. A.; Eriksson, G.; Gheribi, A. E.; Hack, K.; Jung, I.-H.; Kang, Y.-B.; Melançon, J.; Pelton, A. D.; Petersen, S.; Robelin, C.; Sangster, J.; Spencer, P.; Van Ende, M.-A.
 CALPHAD: Comput. Coupling Phase Diagrams Thermochem. 2016, 54, 35–53. doi:10.1016/j.calphad.2016.05.002

- COMSOL Multiphysics®, v.5.3; COMSOL AB: Stockholm, Sweden, https://www.comsol.com.
- Mendoza Conde, G. O.; Luna López, J. A.; Hernández Simón, Z. J.; Hernández de la Luz, J. Á. D.; García Salgado, G.; Gastellou Hernández, E.; Martínez Hernández, H. P.; Flores Méndez, J. Sensors 2022, 22, 3904. doi:10.3390/s22103904
- 30. Martínez Hernández, H.; Luna López, J.; Hernández de la Luz, J.; Luna Flores, A.; Monfil Leyva, K.; García Salgado, G.; Carrillo López, J.; Ordoñez Flores, R.; Pérez García, S.; Hernández Simón, Z.; Mendoza Conde, G.; Ramírez Amador, R. Crystals 2020, 10, 127. doi:10.3390/cryst10020127
- 31. López, J. A. L.; Valerdi, D. E. V.; Salgado, G. G.; de la Luz, A. D. H.; Conde, G. M.; Simon, Z. H.; Gracia, F. J. F.; Sanchez, A. M.; Dominguez, M. A. *Procedia Eng.* **2016**, *168*, 1296–1299. doi:10.1016/j.proeng.2016.11.349
- Tépox, N. C. Efectos Fotoconductivos de Heteroestructuras Con Oxidos Nanoestructurados. Ph.D. Thesis, Benemérita Universidad Autónoma de Puebla, México, 2021.
- Descamps, P.; Asad, S. S.; Wilde, J. D. J. Phys. D: Appl. Phys. 2013, 46, 365201. doi:10.1088/0022-3727/46/36/365201
- 34. Meshkova, A. S.; Elam, F. M.; Starostin, S. A.; van de Sanden, M. C. M.; de Vries, H. W. *Surf. Coat. Technol.* **2018**, *339*, 20–26. doi:10.1016/j.surfcoat.2018.02.010
- 35. Luna López, J. A.; Vázquez Valerdi, D. E.; Benítez Lara, A.; García Salgado, G.; Hernández-de la Luz, A. D.; Morales Sánchez, A.; Flores Gracia, F. J.; Dominguez, M. A. J. Electron. Mater. 2017, 46, 2309–2322. doi:10.1007/s11664-016-5271-1
- Meeks, E.; Larson, R. S.; Ho, P.; Apblett, C.; Han, S. M.; Edelberg, E.;
 Aydil, E. S. J. Vac. Sci. Technol., A 1998, 16, 544–563.
 doi:10.1116/1.581096

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TiO₂ immobilized on 2D mordenite: effect of hydrolysis conditions on structural, textural, and optical characteristics of the nanocomposites

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Full Research Paper

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Abstract

A series of novel $TiO_2/2D$ mordenite nanocomposites were synthetized by the introduction of titanium tetraethoxide (TEOT) into the interlamellar space of 2D mordenite, its subsequent hydrolysis in water or a solution of 70% ethanol in water for 6, 12, and 24 h, and calcination. The resulting $TiO_2/2D$ mordenite materials were studied by a set of complementary characterization techniques, including XRD, SEM-EDX, TGA, N_2 sorption, NMR, XPS and UV-vis spectrometry. It was observed that treatment in 70% ethanol solution preserves the ordered layered structure of 2D mordenite because TEOT hydrolysis is slowed down. This, in turn, leads to higher mesoporosity after calcination due to anatase nanoparticles of about 4 nm preventing the collapse of the interlamellar space. Immobilization of TiO_2 on the zeolite surface is evidenced by the formation of Si-O-Ti bonds. The bandgap width of the synthetized nanocomposites was found to be sensitive to the hydrolysis medium.

Introduction

Zeolites are important heterogeneous catalysts in various industrial processes. More and more functional materials based on zeolites are being searched for, including zeolites with hierar-

chical micro- and mesoporosity [1-6]. Zeolites with hierarchical porous structures can be synthesized using different strategies [4], for example, aggregation of nanocrystals, use of

templates, and creation of mesoporosity by forming pillars between 2D lamellae of zeolites separated by cetyltrimethylammonium bromide (CTAB) layers. The latter method involves the synthesis of layered 2D zeolites in the presence of CTAB and organic structure directing agents (OSDAs), followed by calcination to remove them from the resulting products [1].

The choice of the OSDA determines not only the interlamellar distance and, hence, the mesopore size, but also the aluminum distribution and acidic properties of the obtained material, which are key parameters for catalysts [5,7-10]. One widely used component for the synthesis of 2D zeolites is CTAB [5,9,11-13].

To prepare mesoporous materials from hybrid zeolite–CTAB aggregates, inorganic pillars are pre-formed in the CTAB layers, which will keep the zeolite lamellae from collapsing when burning the organic phase. The flexibility in the choice of material to form pillars creates a wide range of potential new materials for targeted applications. Typically, such pillars are amorphous SiO₂ nanoparticles formed during the hydrolysis of tetraethoxysilane (TEOS) introduced into liquid crystalline 2D CTAB layers that fill the interlamellar space between the 2D zeolite nanosheets. Tetraethoxytitanium (TEOT) is a homolog of TEOS, and its hydrolysis similarly leads to the formation of TiO₂.

 TiO_2 is a well-known photocatalyst whose efficiency depends on a number of factors, including the crystalline phase, particle size, and degree of crystallinity. The most active phase of TiO_2 is considered to be anatase. Its nanoparticles usually show higher efficiency than the bulk phase, but the bandgap of anatase particles smaller than 10 nm is very sensitive to their size [14].

One of the disadvantages of such free photocatalyst nanoparticles is the limitation of mass transfer between solid and liquid phases. From this perspective, the problem of immobilization of TiO₂ nanoparticles is of great importance. Many materials are considered as a possible support for immobilization, including porous ceramics [15], glass [16,17], porous carbon materials [18,19], mesoporous silica [20-23], and zeolites [24-27]. Recent studies have confirmed that direct synthesis of TiO₂ in mesoporous silica or zeolites provides strong immobilization of TiO₂ nanoparticles through Ti–O–Si bonding [21-23].

Previously, we reported the results of the trial synthesis of a new $TiO_2/2D$ mordenite nanocomposite [28]. The material was obtained from a composite consisting of lamellar mordenite separated by CTAB layers through the substitution of TEOS for TEOT and subsequent hydrolysis. As a result, the pillars sepa-

rating the mordenite layers were not made of silica as in the routine synthesis method for the preparation of mesoporous materials, but of anatase nanoparticles of about 4 nm in size. It was found that the textural properties of the resulting materials are sensitive to the environment in which TEOT hydrolysis takes place.

Alkoxides are, generally, not stable in protic solvents such as water. However, because of this property they are widely used as starting products in a large number of reactions. The hydrolysis of various metal alkoxides by pure water, or its mixtures with primarily alcohols or other solvents, are the basis of the sol-gel method to obtain oxide materials. The tendency of metal alkoxides to this reaction can be considered as their most important chemical property. For various reasons, the sol-gel method is mainly associated with the hydrolysis of Si(OR)₄; this reagent is readily available, inexpensive, and its hydrolysis proceeds relatively smoothly, as discussed in numerous original papers and reviews [29]. However, the hydrolysis of $M(OR)_4$ (M = metal), unlike the hydrolysis of $Si(OR)_4$, is an extremely fast process. Thus, the basic concepts that were developed specifically for Si(OR)₄ cannot be applied to the hydrolysis of any arbitrary metal alkoxides. The higher coordination number of metals in their alcoholic and hydroxy derivatives compared to Si(OR)4 leads to a high propensity for oligomerization and polymerization of metal alkoxides after the first stages of hydrolysis.

In the case of tetraethoxysilane, the overall reaction can be written as [29]:

$$Si(OEt)_4 + 2H_2O \Rightarrow SiO_2 + 4HOEt.$$
 (1)

The situation in the case of titanium alkoxides is more ambiguous. In a calorimetric hydrolysis study of $Ti(OR)_4$ at different concentrations, the values of reaction enthalpy of the first hydrolysis stage were measured for R = Et, iPr, and n-Bu [29]. It turned out that further hydrolysis proceeds much slower and with very little heat release (for R = Et its value is zero within the accuracy of the experiment).

$$Ti(OR)_4 + H_2O = Ti(OR)_2(OH) + ROH$$
 (2)

Therefore, this step is immediately followed by condensation:

$$n \operatorname{Ti}(\operatorname{OR})_3(\operatorname{OH}) \Rightarrow \operatorname{Ti}_n \operatorname{O}_x(\operatorname{OR})_{4n-2x}.$$
 (3)

The final composition of the hydrolysis products of alkoxides of titanium roughly corresponds to $TiO_{1.5}(OR) \cdot yROH$, where y = 0.15-1.00 depending on the nature of the alcohol. The residual carbon during thermal treatment in air is eliminated in the process of crystallization at 400–550 °C; thus, titanium dioxide nanoparticles are obtained [29].

In this study we investigate in detail the influence of the hydrolysis medium and the duration of the hydrolytic process on composition, local structure, morphology, texture, and optical properties of $\text{TiO}_2/\text{2D}$ mordenite nanocomposites.

Results and Discussion

 $TiO_2/2D$ mordenite compounds were synthetized from the parent layered MOR-L by introduction of TEOT, its further hydrolysis, and calcination. Further details are given in the Experimental section. The calcined (C) samples are labeled as Ti-WNh-C and Ti-ENh-C with N=6, 12, and 24 for materials hydrolyzed in water (W) and 70% ethanol solution (E) for 6, 12, and 24 h, respectively. The non-calcined samples are designated as Ti-WNh and Ti-ENh.

XRD, ²⁷Al NMR, and SEM-EDX studies

Figure 1a and Figure 1b show small-angle and full X-ray diffraction (XRD) patterns, respectively, for the samples containing hydrolyzed forms of Ti after hydrolysis of TEOT. The full XRD patterns of the hydrolyzed samples after calcination are shown in Figure 1c. For comparison, the corresponding patterns for the parent compound MOR-L are also given.

As it was shown in our previous study [5], the as-synthetized lamellar MOR-L exhibits peaks characteristic of a 3D mordenite structure and small-angle peaks at $2\theta = 2.7^{\circ}$ and 5.5° , which correspond to the (001) and (002) peaks of the ordered layered structure with an interplanar distance of 3.2 nm. According to Figure 1b, the introduction of TEOT and its further hydrolysis do not significantly perturb the mordenite peaks. But the small-angle reflection peaks corresponding to the lamellar structure disappear (Figure 1a); traces of a blurred peak are observed only for the sample with minimal hydrolysis time in 70% ethanol solution. An obvious reason for the loss of lamellar structure is the rate of hydrolysis, which is much higher for TEOT than for TEOS [5]. The rapid and apparently uneven growth of oligomers in different regions of the interlamellar layers leads to a disruption of the long-range order of the lamellae.

In addition, the full XRD patterns evidence the formation of an amorphous phase, presumably nanosized TiO₂ (Figure 1b). Zang et al. [30] showed that the XRD pattern of nanometer-sized amorphous TiO₂ consists of broad humps at 2θ values of 35°, 53°, and 75°. In the TiO₂/2D mordenite composites studied in our work, the first hump is masked by the main zeolite peaks, but the remaining two are identical to the ones reported in [30]. Amorphous TiO₂ particles exhibiting such an XRD pattern can be described as small strained anatase-like crystalline cores with strongly distorted shells. These amorphous nanoparticles are responsible for the formation of single-phase nanocrystalline anatase upon heating [30]. This is exactly what we observe.

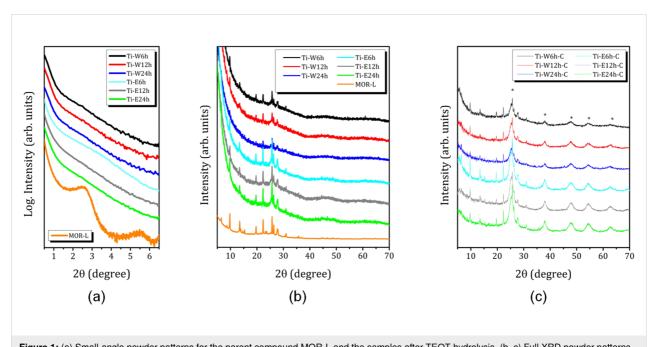


Figure 1: (a) Small-angle powder patterns for the parent compound MOR-L and the samples after TEOT hydrolysis. (b, c) Full XRD powder patterns for the Ti-loaded samples after (b) TEOT hydrolysis and (c) subsequent calcination. Asterisks indicate the anatase structure formed after calcination.

Figure 1c shows that, after calcination, additional large peaks appear in all samples, which can be attributed to anatase TiO_2 nanoparticles. The average size of the TiO_2 nanoparticles, as determined from single diffraction peaks of experimental patterns using the Scherrer formula, is about 4.0 nm for the composites prepared by hydrolysis in water, and slightly larger, about 4.3 nm, after hydrolysis in 70% ethanol solution (Table 1).

In addition, it should be noted that calcination, independent of the medium of TEOT hydrolysis, leads to the complete disappearance of the long-range ordering of the zeolite lamellae (no peak at $2\theta < 5^{\circ}$; the SAXS patterns are not shown here). This is an important difference between TEOS and TEOT, studied in our previous work [5], for which even after hydrolysis for 12 h in water the long-range order of the lamellae was preserved.

The results of the elemental analysis using energy dispersive X-ray (EDX) and X-ray photoelectron spectroscopy (XPS) of the parent lamellar sample and TiO₂-loaded samples are summarized in Table 1. Upon formation of TiO₂ nanoparticles, a partial dealumination of mordenite occurs. However, the Al distribution over the sample depth is not homogenous. XPS shows that Al is accumulated on the sample surface. The hydrolysis medium does not significantly affect the total Si/Al ratio, but hydrolysis reactions longer than 6 h result in a higher aluminum concentration on the surface. It should be also noted that all TiO₂-loaded samples are characterized by low sodium content. We remind the readers that Na⁺ is a charge compensating cation; for an ideal sodium zeolite, Na/Al = 1. The parent MOR-L compound even exhibits a small excess of positive

charge (Na/Al > 1) that is balanced by Br⁻ anions [5]. In the process of post-hydrolysis calcination, the removal of CTAB results in the creation of protonated centers, and the role of compensating cations is eventually passed to protons.

According to the elemental analysis data (Table 1), the TiO₂ loading exceeds 50 wt % in all studied composites. Comparing the data of methods with different measuring depths (i.e., EDX and XPS), we can conclude that Ti is more or less uniformly distributed over the depth in the presented samples. This allows us to state that TiO₂ nanoparticles not only accumulate on the sample surface but are also present in the interlamellar space, which correlates with the nitrogen adsorption data reported next. For both hydrolysis media, the maximum TiO₂ content is reached after 12 h of treatment. For Ti-E24h-C, the lower TiO₂ content compared to TI-E12h-C can be attributed to the formation of 3D mordenite fibers, clearly visible in the SEM images (see below in Figure 3).

From the comparison of the surface (XPS) and volume (EDX) content of ${\rm TiO_2}$, we can not only deduce a fairly uniform distribution of titania over the volume, but also a somewhat higher volume content for samples obtained by hydrolysis in water. This suggests that the formation of anatase particles occurs both on the surface and in the entire accessible mesoporous volume, creating titania pillars for the mesoporous system.

Figure 2 shows the 27 Al magic angle spinning nuclear magnetic resonance (MAS NMR) spectra of the parent compound MOR-L and the TiO_2 -loaded samples. They confirm the regularity of the zeolite frameworks of the as-prepared samples. The

Table 1: Average TiO_2 particle size $\langle d \rangle$ and elemental composition, determined by EDX and XPS, of the as-synthetized and Ti-loaded samples, and their TiO_2 content.

Sample	Method	Si/Al	Na/Al	Ti/Al	TiO ₂ (wt %)	$\langle d \rangle$ (nm) XRD	Ref.
MOR-L	EDX	8.4 ± 0.3	1.14 ± 0.03	_	_	_	[5]
Ti-W6h-C	EDX	10.2 ± 0.4	0.5 ± 0.1	16 ± 1	63 ± 1	4.0 ± 0.1	[28]
	XPS	6.2 ± 0.1	ND ^a	7.6 ± 0.3	52 ± 1		
Ti-W12h-C	EDX	10.2 ± 0.1	0.2 ± 0.1	29 ± 1	73 ± 1	4.1 ± 0.1	this paper
	XPS	4.6 ± 0.1	ND ^a	6.8 ± 0.3	60 ± 1		
Ti-W24h-C	EDX	10.9 ± 0.2	0.4 ± 0.1	30 ± 1	72 ± 1	3.9 ± 0.1	this paper
	XPS	4.8 ± 0.1	ND ^a	8.2 ± 0.3	63 ± 1		
Ti-E6h-C	EDX	9.3 ± 0.6	0.3 ± 0.2	15 ± 2	61 ± 2	4.2 ± 0.1	[28]
	XPS	6.9 ± 0.1	ND ^a	7.0 ± 0.3	51 ± 1		
Ti-E12h-C	EDX	10.6 ± 0.2	0.3 ± 0.1	20 ± 1	66 ± 1	4.3 ± 0.1	this paper
	XPS	4.2 ± 0.1	ND ^a	10.5 ± 0.3	70 ± 1		
Ti-E24h-C	EDX	10.5 ± 0.2	0.4 ± 0.1	10 ± 1	50 ± 1	4.4 ± 0.1	this paper
	XPS	5.3 ± 0.1	ND ^a	6.3 ± 0.3	55 ± 1		

^aThe Na 1s peak overlaps with the Ti LMM Auger peak, which impedes correct evaluation of the Na content.

spectrum consists of only one line at 54 ppm, which corresponds to aluminum in regular tetrahedral sites of the zeolite framework. As it was reported earlier for 2D mordenite pillared by amorphous SiO2, obtained by hydrolysis of TEOT in water, the removal of CTAB leads to a partial removal of Al from the zeolite frameworks [5]. The same is observed for the TiO₂loaded samples. An additional line at about 0 ppm corresponds to extra-framework six-coordinated Al species [31-34], and the relative content of this extra-framework Al does not depend much on duration or medium of hydrolysis (about 20% for all samples except Ti-E6h-C, for which it is 15%) and is mainly determined by the mutual arrangement of the CTA⁺ cations and Al in the zeolite lamellae in the parent MOR-L compound. The CTA+ cations are localized near [AlO₄]- tetrahedra; the removal of organics upon calcination results in a partial collapse of the local zeolite structure.

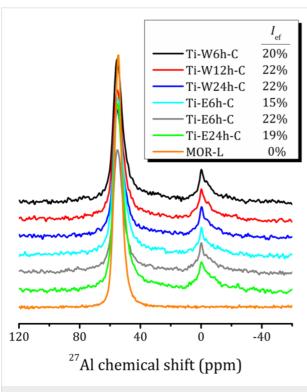


Figure 2: 27 Al MAS NMR spectra of the studied samples. The relative integral intensity of the line at 0 ppm corresponding to extra-framework Al species $I_{\rm eff}$ is shown in the legend.

Figure 3 shows images of all studied composites. The scanning electron microscopy (SEM) image of the initial lamellar mordenite sample MOR-L is also shown for comparison. As can be seen, MOR-L exhibits elongated plates up to 1 μ m long and 0.1 μ m wide, combined into stacks. After introduction of TEOT followed by hydrolysis and calcination, all composites have a similar morphology. Thin plates of about 0.5 μ m in size covered with nanoparticles of about 10 nm in size. The samples

obtained by hydrolysis in 70% ethanol solution exhibit a more foam-like shape. Moreover, hydrolysis in 70% ethanol solution for 24 h results in the formation of thin single-crystalline fibers of mordenite.

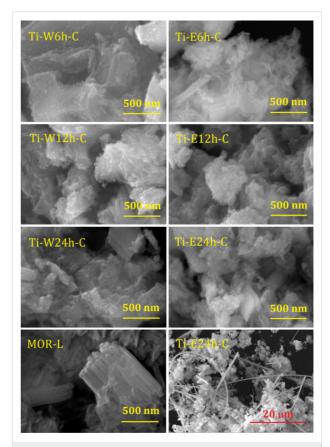


Figure 3: SEM images of the parent lamellar mordenite MOR-L (bottom left) and ${\rm TiO_2}$ -loaded samples. The bottom-right image shows the formation of mordenite crystals in the Ti-E24h-C sample.

Al 2p, Ti 2p, and O 1s XPS studies

The XPS spectra of Al 2p, Ti 2p, and O 1s for the composites are shown in Figure 4. Figure 4a and Figure 4c also show the Al 2p and O 1s spectra of the starting MOR-L compound. No perturbation is observed in the Si 2p region. The only peak at 104.0 eV, corresponding to Si 2p in MOR-L, is slightly shifted towards a lower binding energy (103.3 eV) for all Ti-loaded samples.

The Al 2p XPS spectra of the Ti-loaded composites show differences compared to the initial MOR-L compound (peak at 75.5 eV). The peak is shifted towards lower binding energies (74.6–74.8 eV), and a shoulder appears at 77.5–76.9 and 76.8–76.4 eV for Ti-WNh-C and Ti-ENh-C, respectively. An exemplary decomposition of the Al 2p XPS spectrum for Ti-W24h-C is shown in the inset of Figure 4a. For the Ti-WNh-C series, the position of the additional peak strongly

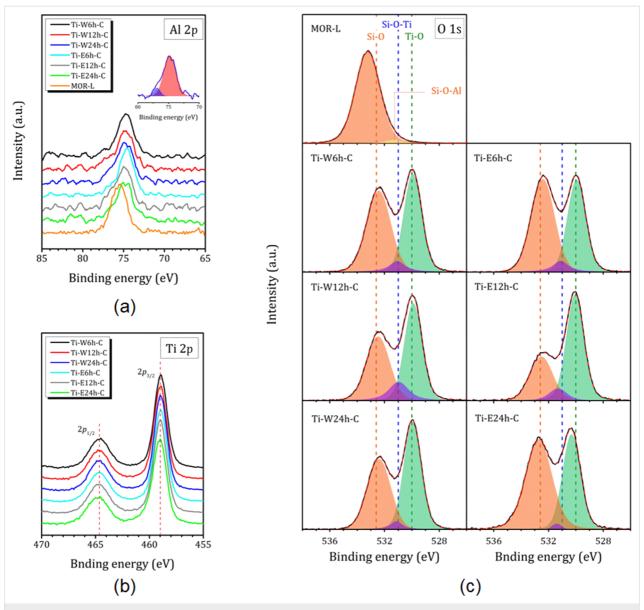


Figure 4: (a) Al 2p, (b) Ti 2p, and (c) decomposed O 1s XPS spectra of the Ti-loaded samples. The Al 2p and O 1s XPS spectra of the MOR-L compound are given for comparison. The inset in (a) shows the decomposition of the Al 2p spectrum for Ti-W24h-C.

depends on the duration of hydrolysis (shift towards lower binding energy with increasing hydrolysis time), whereas the duration of hydrolysis in 70% ethanol solution has no significant effect on the position of the additional peak, but only on its intensity. This peak can be associated with extra-framework Al species, which according to ²⁷Al NMR amount to a total of about 20% (Figure 2). For Ti-E24h-C, a low intensity peak appears at 72.7 eV. This peak can be related to framework Al in the 3D mordenite fibers, which are clearly seen in the SEM image (Figure 3). It should be noted that very often in zeolites the peak at 74.5–74.8 eV is attributed to the presence of Al₂O₃ [35,36]; however, as it will be shown further, this contradicts to the ²⁷Al NMR spectra.

The Ti 2p XPS spectra of the Ti-WNh-C and Ti-ENh-C samples show peaks corresponding to Ti 2p_{1/2} (464.6 eV) and Ti 2p_{3/2} (459.0 eV) transitions, indicating that Ti is in the Ti(IV) state [37] (Figure 3b). To identify the interactions between TiO₂ nanoparticles and zeolite layers in the composites, O 1s XPS spectra were analyzed (Figure 4c). The starting compound MOR-L shows a characteristic peak of Si-O bonds (533.2 eV) with a small shoulder at 531.3 eV, which can be attributed to Si-O-Al. The introduction of TiO₂ shifts the Si-O peak down to 532.4–532.6 eV, and a characteristic peak of Ti-O bonds at 529.9 eV appears (for samples hydrolyzed in 70% ethanol solution for 24 h, it is shifted towards 530.3 eV). The signals from Al-O and Al-OH, expected at 532 and

533 eV, respectively, are evidently masked by a broad signal from the Si–O bonds. The O 1s spectra also reveal the emergence of a new O state at about 530.9–531.2 eV, which can be assigned to the formation of Si–O–Ti bonds [27]. A decrease of the Si–O–Ti signal with the hydrolysis duration suggests that the shortest hydrolysis time provides the most efficient immobilization of TiO₂ nanoparticles on zeolites sheets. However, this

conclusion is difficult to confirm quantitatively as Si-O-Al bonds also may provide a contribution to this line.

Nitrogen sorption and thermogravimetric studies

Figure 5a,b shows the N₂ adsorption/desorption isotherms of the studied nanocomposites. They demonstrate features charac-

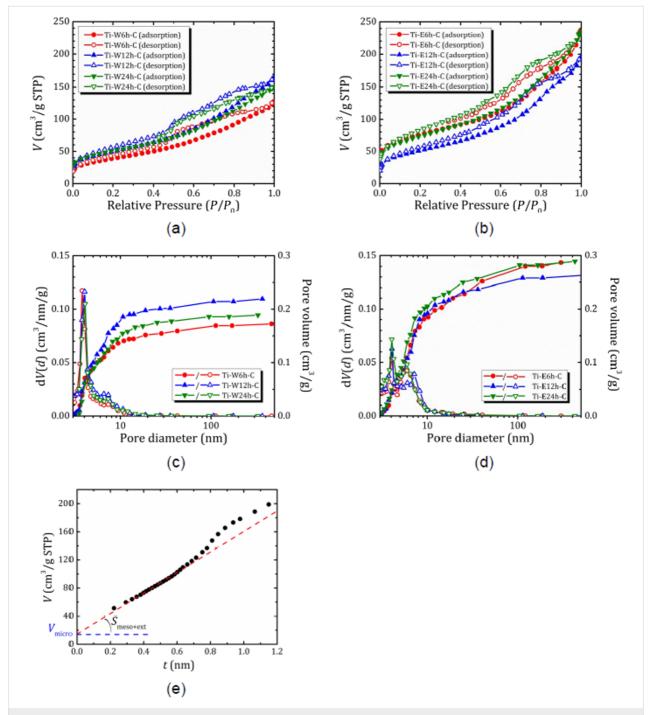


Figure 5: (a, b) Nitrogen adsorption isotherms at 77 K, (c, d) pore size distribution and pore volume in calcined nanocomposites Ti-WNh-C (a, c) and Ti-ENh-C (b, d); (e) t-plot for Ti-E6h-C with a linear fit in the t range from 0.33 to 0.6 nm.

teristic of hierarchical porous structures possessing both microand mesoporosity. At low pressure, there is a sharp gas absorption (isotherm I or II according to IUPAC classification); at increasing pressure, the absorption continues and is accompanied by a hysteresis loop (isotherm IV according to IUPAC classification). For the series of Ti-WNh-C samples hydrolyzed in pure water, the inflection point on the desorption curves appears around $P/P_0 = 0.5$, and it is followed by a broad hysteresis loop. The shape of this loop mainly corresponds to bottle-shaped pores (type H2, more precisely H2b, which corresponds to a pore blocking effect but without percolation, which may also indicate a narrow size distribution of pore cavities), with some presence of slit-shaped pores (type H3). Hydrolysis in 70% ethanol solution (i.e., Ti-ENh-C series) results in a wider distribution of mesopores by size. Volume $V_{\rm BJH}$ and diameter $D_{\rm BJH}$ of the mesopores determined from the desorption branch of the hysteresis loops are given in Table 2. The surface area S_{BET} of the samples were calculated using adsorption data in the range of relative pressures of 0.07-0.22. In addition, to estimate micropore surface area and volume, we applied the t-plot method, which plots the adsorbed volume as a function of the effective thickness (t) of the adsorbed layer. For a correct transformation of relative pressure (P/P_0) to t, we used the formulas proposed in [38,39] for hierarchical microporous/mesoporous zeolites. In this method, a linear fit at low thickness of adsorbate film (low relative pressure) provides the micropore volume $V_{
m micro}$ (the intercept) and the mesopore plus external surface area $S_{\text{meso+ext}}$ (the slope). Then the micropore surface area can be roughly estimated as follows [38]: $S_{\text{micro}} = S_{\text{BET}} - S_{\text{meso+ext}}$. An example of t-plot analysis is shown in Figure 5e. The pore volumes and surface areas evaluated from t-plot are also listed in Table 2.

According to the *t*-plot analysis, the contribution of micropores to the total pore volume does not exceed 10% and is even lower for the sample hydrolyzed in water. As can be seen from Figure 5c, hydrolysis in water leads to the formation of mesopores of about 3.8 nm in size with a mesopore volume of

0.17 cm³/g. Increasing the hydrolysis time up to 12 h results in a slight increase of mesopore size and volume (both evaluated from BJH and *t*-plot methods) that correlates with a more developed surface area. However, prolongation of hydrolysis time does not affect the textural properties of the resulting composites. With an increase hydrolysis duration in the presence of ethanol, first, there is a sharp decrease in porosity. Then, there is an increase to almost the same value as it was after the minimum processing time, which may be due to the formation of microporous zeolite. The decrease in specific surface area for Ti-E12h-C compared to Ti-E6h-C is most likely due to the formation of larger mesopores, which are not observed in Ti-E6h-C and T-E24h-C.

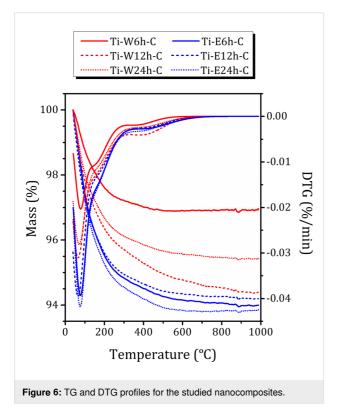
It is interesting to note that the surface area of the studied composites obtained by pillaring of lamellar mordenite with anatase nanoparticles formed in the interlamellar space via hydrolysis is very close to the one reported for $\text{TiO}_2/\text{ZSM-5}$ composites obtained by a related method (by adding the presynthesized zeolite in the synthesis medium of TiO_2 and, vice versa, by adding presynthesized TiO_2 in the synthesis medium of zeolite [27]), lower than in a mechanical mixture of TiO_2 with microporous zeolite, but higher than for composites obtained by a liquid impregnation method [40].

Thermogravimetric (TG) profiles of the studied samples together with the derivative thermogravimetric (DTG) curves are shown in Figure 6. As can be seen, mass loss occurred in two main steps, namely, the rapid desorption of surface water or weakly bound water molecules in the temperature range of 40–100 °C and further mass loss in the temperature range of 100–300 °C, attributed to desorption of the remaining water enclosed in voids and channels [41,42]. However, even at temperatures above 300 °C, all samples exhibit a small peak in the DTG curves at about 400 °C, which can be attributed either to water molecules trapped in hardly accessible voids or, more probably, to a removal of specific hydroxy groups [34]. Moreover, all samples except Ti-W4h-C show a linear mass loss

Table 2: Textural	properties of the Ti-lo	aded samples from N ₂ a	dsorption isotherms	3.		
Sample	$S_{\rm BET}~({\rm m^2/g})$	$S_{\text{meso+ext}}$ (m ² /g)	$S_{\text{micro}} (\text{m}^2/\text{g})$	V _{BJH} (cm ³ /g)	V _{micro} (cm ³ /g)	D _{BJH} (nm)
Ti-W6h-C	134	131	3	0.17	0.004	3.8
Ti-W12h-C	178	178	0	0.22	0.005	4.0
Ti-W24h-C	178	170	8	0.19	0.007	4.1
Ti-E6h-C	268	226	42	0.29	0.022	4.1/6.0 ^a
Ti-E12h-C	183	179	4	0.26	0.005	4.1/7.2 ^a
Ti-E24h-C	259	241	18	0.29	0.015	4.1/6.0 ^a

^aFor the Ti-ENh-C series two pore diameter values correspond to two maxima in the pore size distribution curves.

starting from 300 °C that is not reflected by DTG and can be attributed to progressive dehydroxylation (for hydroxy groups with a significant inhomogeneity) [41]. For the sample Ti-W12h-C, this effect is most pronounced and corresponds to about 7.6% of the total mass loss. The total mass loss Δw as well as the mass loss below and above 300 °C (Δw_1 and Δw_2 , respectively) are listed in Table 3. It should be noted that the samples contain different amounts of TiO₂ (Table 1). Water is mainly located in zeolite voids. Hence, for a more appropriate determination of the amount of water by mass loss, we need to take it into account. The mass losses below and above 300 °C, recalculated assuming that they are due to the zeolite phase alone, Δw_1 and Δw_2 , respectively are also listed in Table 3.



As one can see, there is now obvious correlation between water content determined from Δw_1 and the mesopore volume V_{BJH} estimated from nitrogen adsorption. Despite the larger pore volume and surface area, there is about the same amount of water in the samples obtained by hydrolysis in 70% ethanol solution and in pure water. Nevertheless, there is a strong correlation between the water content $\Delta w_1'$ and the amount of hydroxy groups, $\Delta w_2'$. This points out to a strong interaction of nanoconfined water with hydroxy groups on the inner surface of mesopores, a part of which formed upon calcination [6]. Moreover, the samples obtained in 70% ethanol solution may exhibit different hydrophilic/hydrophobic properties than the samples hydrolyzed in pure water. It is known that the hydrophilicity of TiO₂ depends on the crystalline phase and surface composition, but not on the size [43]. Moreover, it was found that introduction of TiO2 into an amorphous siliceous matrix increases the hydrophilicity of the material. However, as it was shown in [44], the formation of a dense anatase phase has a strong influence on both the value of water adsorption energy and the distribution of water adsorption centers.

UV-vis spectrometry

To determine the bandgap energy $E_{\rm g}$, the Tauc method was applied to the diffuse reflectance spectra. In this method, it is assumed that the energy-dependent absorption coefficient α can be written as

$$(\alpha \cdot h \nu)^{1/n} = B(h \nu - E_g), \tag{4}$$

where h is the Planck constant, hv is the photon energy, and B is a constant. The factor n depends on the nature of the electron transition, that is, n = 1/2 for direct and n = 2 for indirect transition bandgaps. The Kubelka–Munk method allows one to transform the reflectance spectra into the corresponding adsorption spectra using the function

Table 3: Mass loss below $(\Delta w_1, \Delta w_1')$ and above $(\Delta w_2, \Delta w_2')$ 300 °C as directly determined from TG curves $(\Delta w_1, \Delta w_2)$ and recalculated accounting for TiO₂ content $(\Delta w_1', \Delta w_2')$. The total mass losses Δw and $\Delta w'$ are also provided.

Sample	40 °C < T < 300 °C		300 °C < T <	300 °C < T < 1000 °C		300 °C < T < 1000 °C	
	Δw ₁ (%)	Δw ₁ ′ (%)	Δw ₂ (%)	_	Δw ₁ (%)	Δw ₁ ′ (%)	
Ti-W6h-C	2.8 ± 0.1	7.6 ± 0.3	0.3 ± 0.1	Ti-W6h-C	2.8 ± 0.1	7.6 ± 0.3	
Ti-W12h-C	3.9 ± 0.1	14.4 ± 0.3	0.7 ± 0.1	Ti-W12h-C	3.9 ± 0.1	14.4 ± 0.3	
Ti-W24h-C	4.3 ± 0.1	15.3 ± 0.3	1.3 ± 0.1	Ti-W24h-C	4.3 ± 0.1	15.3 ± 0.3	
Ti-E6h-C	5.2 ± 0.1	13.3 ± 0.5	0.9 ± 0.1	Ti-E6h-C	5.2 ± 0.1	13.3 ± 0.5	
Ti-E12h-C	5.0 ± 0.1	14.8 ± 0.3	0.8 ± 0.1	Ti-E12h-C	5.0 ± 0.1	14.8 ± 0.3	
Ti-E24h-C	5.5 ± 0.1	10.9 ± 0.3	0.7 ± 0.1	Ti-E24h-C	5.5 ± 0.1	10.9 ± 0.3	

$$F(R) = \frac{K}{S} = \frac{\left(1 - R\right)^2}{2R},\tag{5}$$

where R is the reflectance of a thick uniform sample, and K and S are the adsorption and scattering coefficients, respectively. When $F(R_{\infty}) \sim \alpha$, by combing Equation 4 and Equation 5, one obtains

$$(F(R) \cdot h\nu)^{1/n} = B(h\nu - E_g). \tag{6}$$

Figure 7a shows the diffuse reflectance spectra of the prepared $TiO_2/2D$ mordenite nanocomposite. All samples exhibit absorption edges near 400 nm due to the anatase TiO_2 bandgap absorption. Figure 7b shows the reflectance spectra of the studied nanocomposites transformed using Equation 6 with n=2 (since TiO_2 is an indirect bandgap semiconductor). Semiconductor materials are characterized by a steep linear increase in light absorption with increasing energy. The bandgap energy can be estimated from the point of intersection of the x-axis of the linear fit of the Tauc plot.

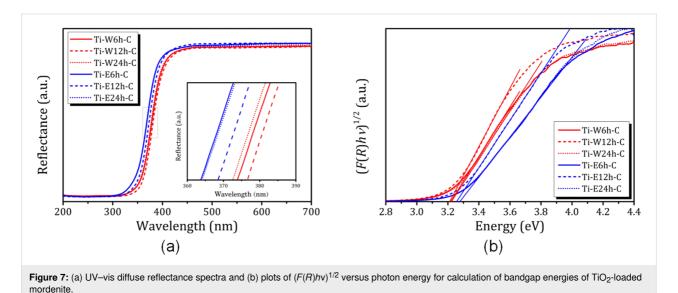
As one can see from Figure 7b, the $E_{\rm g}$ values for the studied TiO₂/2D mordenite nanocomposites are sensitive to the hydrolysis medium. The samples obtained by hydrolysis in water exhibit $E_{\rm g}$ values of about 3.22 eV, which is close to the value of bulk anatase [45,46]. Hydrolysis in 70% ethanol solution results in higher $E_{\rm g}$ values, about 3.27 eV, which are typical for anatase nanoparticles [45].

It should be noticed that according to a number of studies, the bandgap width in TiO₂/zeolite composites depends both on the

topology of the zeolite framework [47] and on the ratio between TiO_2 and zeolite [47,48]. Alvarez et al. [47] reported that for TiO_2 deposited on mordenite during the TiO_2 sol–gel synthesis in a ratio of 75:25, which is close to that in our work (i.e., between 50:50 and 70:30), leads to a noticeable increase in the bandgap (3.47 eV) [47]. It should also be noted that TiO_2 /zeolite composites that were obtained by either incorporating the presynthesized zeolite in the synthesis medium of TiO_2 or incorporating presynthesized TiO_2 in the synthesis medium of ZSM-5 [27] exhibit optical properties very similar to those found in our work.

Conclusion

A series of pillared TiO₂/2D mordenite nanocomposites were successfully synthetized from the initial lamellar CTAB/2D mordenite by introducing TEOT, hydrolyzing the resulting composite in either water or 70% ethanol solution with hydrolysis reaction times of 6, 12, and 24 h, and subsequent calcination. The main results of the comprehensive study of the obtained composites can be summarized as follows. (1) A study of the TEOT hydrolysis process was carried out, and it was found that the properties of the samples depend significantly on the hydrolysis medium and the duration of the process. The hydrolysis of TEOT in the interlamellar CTAB layers in all cases disrupts the long-range ordering of zeolite lamellae. Ethanol allows one to attenuate and slow down this process compared to pure water. In samples prepared in 70% ethanol solution, the residual ordering of the lamellar structure at the first stage of the process is preserved to some extent, and the textural characteristics are better compared to samples prepared in pure water. Further calcination, regardless of the medium in which TEOT hydrolysis occurred, leads to the final disappearance of the long-range ordering of zeolite lamellas. (2) According to XRD data, ana-



tase nanoparticles of about 4 nm in size form pillars separating the mordenite layers, creating the mesoporosity observed in the experiment. The textural properties of the samples strongly depend on the hydrolysis medium. Hydrolysis in 70% ethanol solution leads to a bimodal distribution of pore sizes, with peaks at 4 and 6 nm, as well as to an increase in the volume of mesopores. (3) Hydrolysis in 70% ethanol solution for 24 h leads to recrystallization of 2D mordenite and to the formation of thin single-crystalline fibers of 3D mordenite. (4) Immobilization of TiO₂ nanoparticles on the zeolite support was confirmed by XPS. (5) Samples obtained by hydrolysis in water exhibit a bandgap of 3.22 eV, which is close to that of bulk anatase. Hydrolysis in 70% ethanol solution results to higher bandgap values, around 3.27 eV, which are characteristic of anatase nanoparticles.

Despite the still large bandgap, the immobilization of TiO_2 on the zeolite matrix, combined with the mesopore structure important for high mass transfer properties, suggests that these materials may be promising catalysts under flow conditions. However, it is necessary to further search for parameters to regulate the growth of nanoparticles in the interlayer space (e.g., the temperature of hydrolysis) and their morphology, as well as to investigate the influence of synthesis conditions on the target properties. Nevertheless, the successful application of zeolite pillaring using an oxide material other than SiO_2 opens up a wide range of opportunities for the development of new materials for specific applications.

Experimental

The synthesis of nanocomposites was carried out similarly to the method first proposed in [49] and used previously in [5,50] for the insertion of SiO2 nanopillars. The latter includes the following steps: (1) synthesis of layered zeolite by selfassembly using CTAB as an organic structural guiding agent that creates regular mesopores, (2) introduction of TEOS molecules into the interlayer space, (3) hydrolysis and formation of pillars of amorphous SiO2 separating the layers of two-dimensional zeolite, and (4) calcination for the removal of organic molecules. In this work, to obtain TiO2/2D mordenite composites (layers of two-dimensional mordenite separated by TiO2 pillars), the synthesis method was slightly adjusted and TEOS was replaced by TEOT. This substitution, taking into account the differences in the properties of these compounds, required changes in both the reagents for hydrolysis (processes in pure water and in 70% ethanol solution were compared) and in the choice of process time intervals.

In the synthesis of $TiO_2/2D$ mordenite compounds, the layered MOR-L sample obtained in step (1), was stirred in TEOT at a mass ratio of 1:5 for 6 h at 25 °C. The sample was filtered off

and then dried at 35 °C for 12 h. To hydrolyze TEOT that had diffused into the CTAB layers and to obtain TiO_2 nanoparticles in the interlamellar space, 1.0 g of TEOT-impregnated MOR-L sample was stirred in either 10.0 g of distilled water or a 70% ethanol-in-water solution, at 90 °C, for 6, 12, and 24 h. After completion of the hydrolysis, the samples were filtered, washed with distilled water, dried at 120 °C, and finally calcined at 550 °C for 4 h in air. The calcined (C) samples are labeled as Ti-WNh-C and Ti-ENh-C with N=6, 12, and 24 for materials hydrolyzed in water (W) and 70% ethanol solution (E) for 6, 12, or 24 h, respectively. The non-calcined samples are labeled as Ti-WNh and Ti-ENh.

Powder XRD analysis was carried out on a Bruker D8 DISCOVER diffractometer with monochromatic Cu K α radiation. XRD patterns of the studied compounds were recorded in the 2 θ range of 5–70 $^{\circ}$ with a step width of 0.0302 $^{\circ}$. SAXS patterns were recorded over a 2 θ scanning range of 0.2–7.0 $^{\circ}$ with a step width of 0.01 $^{\circ}$.

Elemental analysis and morphology of synthetized TiO₂/2D mordenite nanocomposites were probed by SEM-EDX using a Zeiss Merlin microscope equipped with an Oxford Instruments INCAx-act EDX console.

N₂ sorption isotherms were recorded at 77 K using a Quadra-Sorb SI instrument. Before analysis, samples were outgassed under vacuum for 6 h at 300 °C. The surface areas were estimated within the mBET method. Thermal gravimetric analysis (TGA) was carried out using a Netzsch STA 449 F1 Jupiter instrument in the temperature range of 40–990 °C at a heating rate of 10 °C/min in an Ar flow of 90 mL/min.

²⁷Al MAS NMR spectra were recorded using a Bruker Avance IIIWB 400 MHz solid-state NMR spectrometer (operating with Topspin version 3.2) using a double-resonance 4 mm MAS probe with a rotor speed of 12.5 kHz.

XPS spectra of the samples were taken using a Thermo Fisher Scientific Escalab 250Xi spectrometer with non-monochromatic Al K α radiation (photon energy 1486.6 eV). Bandgaps energies were determined by diffuse reflectance spectroscopy using a Lambda 1050 spectrophotometer (Perkin Elmer) equipped with an integrating sphere in the spectral range of 200–700 nm. Barium sulfate (BaSO₄) was used as a reference.

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Author Contributions

Marina G. Shelyapina: funding acquisition; project administration; supervision; visualization; writing – original draft; writing – review & editing. Rosario Isidro Yocupicio-Gaxiola: conceptualization; investigation; resources. Gleb A. Valkovsky: data curation; formal analysis; funding acquisition; investigation. Vitalii Petranovskii: conceptualization; methodology; supervision; writing – original draft; writing – review & editing.

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Data Availability Statement

The data that supports the findings of this study is available from the corresponding author upon reasonable request.

References

- Roth, W. J.; Nachtigall, P.; Morris, R. E.; Čejka, J. Chem. Rev. 2014, 114, 4807–4837. doi:10.1021/cr400600f
- Zhang, K.; Ostraat, M. L. Catal. Today 2016, 264, 3–15. doi:10.1016/j.cattod.2015.08.012
- Ge, T.; Hua, Z.; He, X.; Zhu, Y.; Ren, W.; Chen, L.; Zhang, L.;
 Chen, H.; Lin, C.; Yao, H.; Shi, J. Chin. J. Catal. 2015, 36, 866–873.
 doi:10.1016/s1872-2067(14)60263-1
- Jia, X.; Khan, W.; Wu, Z.; Choi, J.; Yip, A. C. K. Adv. Powder Technol. 2019, 30, 467–484. doi:10.1016/j.apt.2018.12.014
- Shelyapina, M. G.; Yocupicio-Gaxiola, R. I.; Zhelezniak, I. V.; Chislov, M. V.; Antúnez-García, J.; Murrieta-Rico, F. N.; Galván, D. H.; Petranovskii, V.; Fuentes-Moyado, S. *Molecules* 2020, 25, 4678. doi:10.3390/molecules25204678
- Shelyapina, M. G.; Nefedov, D. Y.; Antonenko, A. O.; Hmok, H.; Egorov, A. V.; Egorova, M. I.; Ievlev, A. V.; Yocupicio-Gaxiola, R.; Petranovskii, V.; Antúnez-García, J.; Fuentes, S. Appl. Magn. Reson. 2023, 54, 915–928. doi:10.1007/s00723-023-01589-w
- Guo, G.-q.; Sun, Y.-j.; Long, Y.-c. Chem. Commun. 2000, 1893–1894. doi:10.1039/b005453o
- Migliori, M.; Aloise, A.; Giordano, G. Catal. Today 2014, 227, 138–143. doi:10.1016/j.cattod.2013.09.033

- Li, H.-J.; Zhou, X.-D.; Di, Y.-H.; Zhang, J.-M.; Zhang, Y. *Microporous Mesoporous Mater.* 2018, 271, 146–155. doi:10.1016/j.micromeso.2018.05.039
- Catizzone, E.; Migliori, M.; Mineva, T.; van Daele, S.; Valtchev, V.; Giordano, G. *Microporous Mesoporous Mater.* 2020, 296, 109988. doi:10.1016/j.micromeso.2019.109988
- Xu, D.; Feng, J.; Che, S. Dalton Trans. 2014, 43, 3612–3617. doi:10.1039/c3dt53308e
- Wang, X.; Chen, H.; Meng, F.; Gao, F.; Sun, C.; Sun, L.; Wang, S.;
 Wang, L.; Wang, Y. *Microporous Mesoporous Mater.* 2017, 243, 271–280. doi:10.1016/j.micromeso.2017.02.054
- Xu, L.; Ji, X.; Li, S.; Zhou, Z.; Du, X.; Sun, J.; Deng, F.; Che, S.; Wu, P. Chem. Mater. 2016, 28, 4512–4521.
 doi:10.1021/acs.chemmater.6b02155
- 14. Brus, L. E. J. Chem. Phys. 1984, 80, 4403–4409. doi:10.1063/1.447218
- 15. de Araujo Scharnberg, A. R.; de Loreto, A. C.; Wermuth, T. B.; Alves, A. K.; Arcaro, S.; dos Santos, P. A. M.; de Assis Lawisch Rodriguez, A. Bol. Soc. Esp. Ceram. Vidrio 2020, 59, 230–238. doi:10.1016/j.bsecv.2019.12.001
- Cunha, D. L.; Kuznetsov, A.; Achete, C. A.; da Hora Machado, A. E.; Marques, M. PeerJ 2018, 6, e4464. doi:10.7717/peerj.4464
- Zvereva, I. A.; Kalinkina, L. M.; Rodionov, I. A.; Sankovich, A. M.; Kolesnik, I. V.; Gudilin, E. A. *Glass Phys. Chem.* **2012**, *38*, 504–510. doi:10.1134/s1087659612060065
- Zou, X.-X.; Li, G.-D.; Zhao, J.; Su, J.; Wei, X.; Wang, K.-X.;
 Wang, Y.-N.; Chen, J.-S. Int. J. Photoenergy 2012, 2012, 720183.
 doi:10.1155/2012/720183
- Scaria, J.; Karim, A. V.; Divyapriya, G.; Nidheesh, P. V.; Suresh Kumar, M. Carbon-supported semiconductor nanoparticles as effective photocatalysts for water and wastewater treatment; Nano-Materials as Photocatalysts for Degradation of Environmental Pollutants; Elsevier: Amsterdam, Netherlands, 2020; pp 245–278. doi:10.1016/b978-0-12-818598-8.00013-4
- Vanichvattanadecha, C.; Jaroenworaluck, A.; Henpraserttae, P.; Wimuktiwan, P.; Manpetch, P.; Singhapong, W. J. Porous Mater. 2021, 28, 1137–1153. doi:10.1007/s10934-021-01065-5
- Degirmenci, V.; Erdem, Ö. F.; Ergun, O.; Yilmaz, A.; Michel, D.; Uner, D. *Top. Catal.* **2008**, *49*, 204–208. doi:10.1007/s11244-008-9078-z
- Zaccariello, G.; Moretti, E.; Storaro, L.; Riello, P.; Canton, P.; Gombac, V.; Montini, T.; Rodríguez-Castellón, E.; Benedetti, A. RSC Adv. 2014, 4, 37826–37837. doi:10.1039/c4ra06767c
- Zhu, L.; Nguyen, D. C. T.; Woo, J.-H.; Zhang, Q.; Cho, K. Y.; Oh, W.-C. Sci. Rep. 2018, 8, 12759. doi:10.1038/s41598-018-31188-w
- Fukahori, S.; Ichiura, H.; Kitaoka, T.; Tanaka, H. Appl. Catal., B 2003, 46, 453–462. doi:10.1016/s0926-3373(03)00270-4
- Diban, N.; Pacuła, A.; Kumakiri, I.; Barquín, C.; Rivero, M. J.; Urtiaga, A.; Ortiz, I. *Catalysts* 2021, 11, 1367. doi:10.3390/catal11111367
- Znad, H.; Abbas, K.; Hena, S.; Awual, M. R. J. Environ. Chem. Eng. 2018, 6, 218–227. doi:10.1016/j.jece.2017.11.077
- Fernández-Catalá, J.; Sánchez-Rubio, M.; Navlani-García, M.;
 Berenguer-Murcia, Á.; Cazorla-Amorós, D. ACS Omega 2020, 5, 31323–31331. doi:10.1021/acsomega.0c04793
- Shelyapina, M. G.; Mazur, A.; Yocupicio-Gaxiola, R. I.; Caudillo-Flores, U.; Urtaza, A.; Rodionov, I. A.; Zvereva, I. A.; Petranovskii, V. Appl. Magn. Reson. 2022, 53, 1609–1620. doi:10.1007/s00723-022-01498-4

- Turova, N. Ya.; Turevskaya, E. P.; Kessler, V. G.; Yanovskaya, M. I. The Chemistry of Metal Alkoxides; Kluwer Academic Publishers: Boston, 2002. doi:10.1007/b113856
- Zhang, H.; Chen, B.; Banfield, J. F.; Waychunas, G. A.
 Phys. Rev. B: Condens. Matter Mater. Phys. 2008, 78, 214106.
 doi:10.1103/physrevb.78.214106
- Deng, F.; Yue, Y.; Ye, C. J. Phys. Chem. B 1998, 102, 5252–5256.
 doi:10.1021/jp9801929
- Li, S.; Zheng, A.; Su, Y.; Fang, H.; Shen, W.; Yu, Z.; Chen, L.; Deng, F. Phys. Chem. Chem. Phys. 2010, 12, 3895–3903.
 doi:10.1039/b915401a
- 33. Zhukov, Y. M.; Efimov, A. Yu.; Shelyapina, M. G.; Petranovskii, V.; Zhizhin, E. V.; Burovikhina, A.; Zvereva, I. A. *Microporous Mesoporous Mater.* 2016, 224, 415–419. doi:10.1016/j.micromeso.2015.12.058
- 34. Shelyapina, M. G.; Krylova, E. A.; Zhukov, Y. M.; Zvereva, I. A.; Rodriguez-Iznaga, I.; Petranovskii, V.; Fuentes-Moyado, S. *Molecules* 2019, 24, 4216. doi:10.3390/molecules24234216
- Singh, B. K.; Kim, Y.; Kwon, S.; Na, K. Catalysts 2019, 9, 885. doi:10.3390/catal9110885
- 36. Lee, J. S.; Kim, H. S.; Park, N.-K.; Lee, T. J.; Kang, M. *Chem. Eng. J.* **2013**, *230*, 351–360. doi:10.1016/j.cej.2013.06.099
- Domoroshchina, E. N.; Chernyshev, V. V.; Kuz'micheva, G. M.;
 Dorokhov, A. V.; Pirutko, L. V.; Kravchenko, G. V.; Chumakov, R. B.
 Appl. Nanosci. 2018, 8, 19–31. doi:10.1007/s13204-018-0648-5
- 38. Galarneau, A.; Mehlhorn, D.; Guenneau, F.; Coasne, B.; Villemot, F.; Minoux, D.; Aquino, C.; Dath, J.-P. Langmuir 2018, 34, 14134–14142. doi:10.1021/acs.langmuir.8b02144
- Desmurs, L.; Galarneau, A.; Cammarano, C.; Hulea, V.; Vaulot, C.; Nouali, H.; Lebeau, B.; Daou, T. J.; Vieira Soares, C.; Maurin, G.; Haranczyk, M.; Batonneau-Gener, I.; Sachse, A. ChemNanoMat 2022, 8, e202200051. doi:10.1002/cnma.202200051
- Mergenbayeva, S.; Abitayev, Z.; Batyrbayeva, M.; Vakros, J.; Mantzavinos, D.; Atabaev, T. S.; Poulopoulos, S. G. Catalysts 2024, 14, 147. doi:10.3390/catal14020147
- 41. Shelyapina, M. G.; Nefedov, D. Y.; Antonenko, A. O.; Valkovskiy, G. A.; Yocupicio-Gaxiola, R. I.; Petranovskii, V. Int. J. Mol. Sci. 2023, 24, 15898. doi:10.3390/ijms242115898
- 42. Krylova, E. A.; Shelyapina, M. G.; Nowak, P.; Harańczyk, H.; Chislov, M.; Zvereva, I. A.; Privalov, A. F.; Becher, M.; Vogel, M.; Petranovskii, V. *Microporous Mesoporous Mater.* 2018, 265, 132–142. doi:10.1016/j.micromeso.2018.02.010
- Bolis, V.; Busco, C.; Ciarletta, M.; Distasi, C.; Erriquez, J.; Fenoglio, I.; Livraghi, S.; Morel, S. J. Colloid Interface Sci. 2012, 369, 28–39. doi:10.1016/j.jcis.2011.11.058
- Boffa, V.; Parmeggiani, L.; Nielsen, A. H.; Magnacca, G. Microporous Mesoporous Mater. 2016, 221, 81–90. doi:10.1016/j.micromeso.2015.09.017
- López, R.; Gómez, R. J. Sol-Gel Sci. Technol. 2012, 61, 1–7. doi:10.1007/s10971-011-2582-9
- Makuła, P.; Pacia, M.; Macyk, W. J. Phys. Chem. Lett. 2018, 9, 6814–6817. doi:10.1021/acs.jpclett.8b02892
- Alvarez, K. M.; Alvarado, J.; Soto, B. S.; Hernandez, M. A. *Optik (Munich, Ger.)* 2018, 169, 137–146. doi:10.1016/j.ijleo.2018.05.028
- Munguti, L. K.; Dejene, F. B.; Muthee, D. K. Mater. Sci. Eng., B 2023, 289, 116281. doi:10.1016/j.mseb.2023.116281
- Na, K.; Choi, M.; Park, W.; Sakamoto, Y.; Terasaki, O.; Ryoo, R.
 J. Am. Chem. Soc. 2010, 132, 4169–4177. doi:10.1021/ja908382n

 Yocupicio-Gaxiola, R. I.; Petranovskii, V.; Antúnez-García, J.; Fuentes Moyado, S. Appl. Nanosci. 2019, 9, 557–565. doi:10.1007/s13204-018-0935-1

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Zeolite materials with Ni and Co: synthesis and catalytic potential in the selective hydrogenation of citral

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Full Research Paper

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Abstract

Zeolitic materials incorporating mono- and bimetallic systems of nickel and cobalt were obtained from natural zeolite modified with Ni²⁺ and Co²⁺ chloride solutions through traditional ion exchange (IE) and impregnation (Imp) processes. Special attention was given to analyzing the cationic and anionic composition of the resulting materials. The catalytic potential was evaluated in the selective hydrogenation of citral, focused on the formation of unsaturated alcohols. The IE process replaced mainly Ca²⁺ and Na⁺ with Ni²⁺ and Co²⁺ cations in the zeolite phases (clinoptilolite and mordenite mix), while Imp resulted in higher metal content (2.0–2.7%) but retained significant amounts of chloride (1.9–3.8%), as confirmed by XRD and temperature-programmed reduction. The materials prepared by IE had negligible chloride content (0.02–0.07%), and their specific surface areas (138–146 m²/g) were greater than those of the materials obtained by Imp (54–67 m²/g). The bimetallic systems exhibited enhanced reducibility of the Co²⁺ and Ni²⁺ isolated cations, attributed to synergistic interactions that weakened the cation–framework binding. Catalytic activity tests showed that nickel species were primarily responsible for citronellal formation. Among all materials, the bimetallic CoNi_{IE} catalyst, prepared by IE, was the only one to produce unsaturated alcohols, suggesting that synergistic Ni–Co interactions played a role in their formation.

Introduction

Numerous publications in the literature highlight zeolites modified with metallic species for various applications, leading to the invention of new functional materials for sustainable development, such as catalysts [1-3]. Among the various methods used to modify zeolites, ion exchange is the most widely employed. Different ion exchange methods are known, such as ion exchange in conventional solutions, in the solid phase with molten salts, and with gaseous phases. Depending on the chosen modification method, particularities regarding the elemental composition and application of the resulting materials occur [4-6]. Cu-Y zeolites were obtained by contacting Na-Y zeolite and Cu(II) nitrate solution, using two different methods, namely, conventional solution ion exchange and incipient wetness impregnation, followed by calcinations at 600 °C in air [5]. The authors reported that, among both zeolites, the Cu-Y material obtained by impregnation followed by calcination exhibited a higher surface area and pore volume, which can positively influence its potential application as a material to reduce greenhouse gas emissions.

While most studies focus on monocationic exchange, multicationic exchange has raised significant interest. The synergy of properties in multicationic systems enables the creation of materials with enhanced properties compared to monocationic zeolites, which is crucial for the development of advanced catalysts and other materials [7-10]. Natural zeolites attract significant attention because of their abundance, low cost, non-toxic nature, and other valuable physical and chemical properties.

Modifying these materials with inexpensive metals leads to low-cost catalysts for various different processes. The selective hydrogenation of α,β -unsaturated aldehydes, such as citral, to unsaturated alcohols is a crucial reaction for producing fine chemicals, fragrances, and other high-value products [11]. The catalysts preferred for this reaction are currently based on noble metals because of their excellent performance [12,13]. However, their high cost and scarcity requires strategies to reduce their use or replace them with non-noble alternatives offering comparable catalytic properties.

Despite extensive research on zeolite-supported transition metal catalysts, their application to citral hydrogenation remains very little explored [11]. Most recent studies focus on the selective hydrogenation of related biomass-derived compounds, such as furfural and cinnamaldehyde [3,14,15]. Zeolites modified with nickel and cobalt have shown promising results in selective hydrogenation reactions, owing to their high dispersion of active sites and tunable acidity. For instance, a zeolite-supported Ni catalyst has demonstrated selectivity in furfural hydrogenation by leveraging controlled acidity to prevent over-

hydrogenation and optimize product yields [15]. Similarly, Co-modified zeolites have been effectively used in cinnamaldehyde hydrogenation, where the balance between acidity and metal dispersion facilitates the selective reduction of the carbonyl group while preserving other reactive sites [3].

Building on these insights, this work presents research on the selective hydrogenation of citral using both monometallic and bimetallic nickel and cobalt catalysts supported on natural zeolite, which was modified with Ni²⁺ and Co²⁺ chloride solutions through traditional ion exchange (IE) and impregnation (Imp) processes. Emphasis is put on analyzing the cationic and anionic composition of the materials resulting from both methods and the catalytic performance in citral hydrogenation with a focus on enhancing the formation of unsaturated alcohols.

Results and Discussion Composition and characterization of the materials

XRD patterns and a SEM micrograph of the starting zeolite mineral (ZSA) are shown in Figure 1 and Figure 2, respectively. The diffraction patterns are normalized and evidence the presence of mordenite and clinoptilolite—heulandite-type zeolites through their main diffraction peaks indicated on the ZSA graph. Other minor phases such as quartz are also present.

The SEM image shows a variety of crisscross crystals, which have the morphology expected for the zeolite types evidenced by XRD [16,17]. Very elongated crystals with acicular to fibrous characteristics, associated with mordenite, can be observed. Additionally, clinoptilolite—heulandite crystals with slats and tabular morphology are present. The amounts of clinoptilolite—heulandite and mordenite crystals displayed in the SEM image correspond to the intensity order of these zeolites shown in the XRD pattern (Figure 1). This indicates that zeolitic mineral from the San Andrés deposit is mainly composed of a mordenite and clinoptilolite—heulandite mixture. Furthermore, the SEM micrographs reveal free spaces between crystallites, contributing to the material's porosity and mesoporosity.

The XRD patterns of the materials obtained using both Imp and IE showed that the framework of both zeolites remains largely unchanged after the applied treatments. However, the materials obtained by Imp exhibit new peaks, such as those observed at 16.1° and 39.3° in CoZ_{Imp} , at 29.5° in NiZ_{Imp} , and at 28.5° in $CoNiZ_{Imp}$. These peaks are attributed to impurities, such as mixed metal chloride salts, deposited on the zeolite surface.

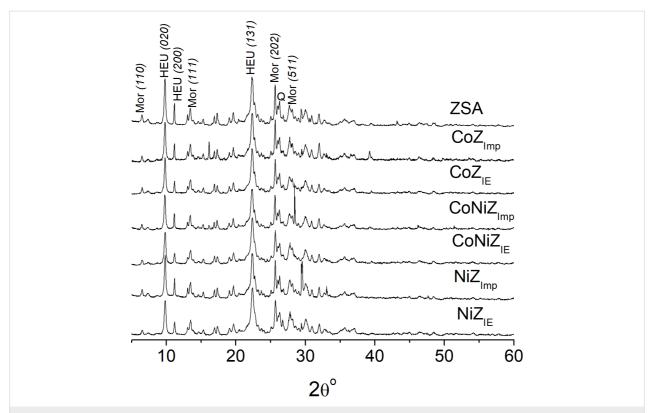


Figure 1: XRD pattern of the starting zeolite mineral (ZSA) and the materials obtained through traditional ion exchange (IE) and impregnation (Imp) methods. The Miller indexes (*hkl*) corresponding to the zeolite phases are shown in the pattern of the ZSA. Mor, HEU, and Q are associated to the phases of mordenite, clinoptilolite—heulandite, and quartz, respectively.

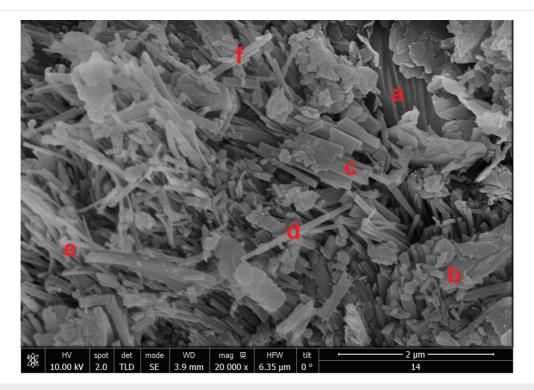


Figure 2: SEM micrograph of ZSA from the San Andrés deposit. Some HEU crystals are indicated by a, b, and c, where a and b exhibit tabular characteristics while c has slat morphology. Mordenite crystals, marked by d, e, and f, show acicular to fibrous shapes.

Specifically, the peaks at 16.1° and 39.3° correspond to both cobalt(II) chloride (CoCl₂·2H₂O, card 96-900-9874) and cobalt(II) hydroxychloride (Co₂Cl(OH)₃, card 96-231-0849). The diffraction peak at 29.5° is associated with NiCl₂ (card 96-900-9133), but it could also be assigned to FeCl₂ (lawrencite, card 96-901-9129) or CaCl₂·2H₂O (sinjarite, card 96-100-1836). The peak at 28.5° can be related to FeCl₂·2H₂O (card 96-231-0808), NaCl (halite, card 96-900-0630), or CaCl₂O₄ (calcium hypochlorite, card 96-220-7380). The low nickel and cobalt contents in the CoNiZ_{Imp} material (Table 1) may limit the detectability of diffraction peaks associated to cobalt and nickel chloride salts on the zeolite support.

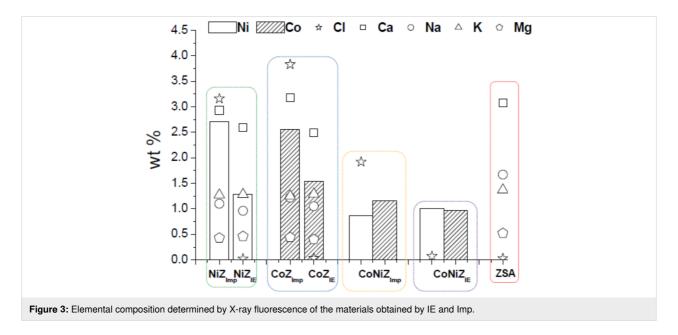
	It and nickel conten hrough IE and Imp.		values for the mate-
Sample	Co, wt %	Ni, wt %	Si/Al
CoZ _{IE}	1.54 ± 0.08	_	5.47
NiZ _{IE}	_	1.29 ± 0.1	5.20
CoNiZ _{IE}	0.97 ± 0.07	1.00 ± 0.08	5.47
CoZ _{Imp}	2.56 ± 0.20	_	5.35
NiZ _{Imp}	_	2.71 ± 0.21	5.22
CoNiZ _{Imp}	1.16 ± 0.08	0.87 ± 0.06	5.17
ZSA	_	_	5.20

The contents of cobalt and nickel and other elements, as well as the Si/Al ratios for the obtained materials are shown in Table 1 and/or Figure 3. The Si/Al ratio shows no significant change (Table 1), indicating a high degree of stability for the clinoptilolite—heulandite and mordenite phases after the applied hydrothermal treatments. Additionally, it can be observed that the Ni and Co concentrations are generally higher in materials ob-

tained through Imp compared to those produced via IE. However, it is important to note that materials obtained by Imp tend to have high chloride contents, whereas those produced by IE exhibit negligible chloride levels (Figure 3). The chloride originates from the NiCl2 and CoCl2 solutions used in the treatments, which remain as residues on the materials. The residual solution may be retained through surface adsorption within the porosity and mesoporosity formed between the zeolite crystallites, as shown in the SEM image (Figure 2). To remove excess solution, water washes are typically applied. The materials obtained via IE underwent extensive washing with distilled water, while those obtained via Imp were only lightly washed, leading to the observed differences in chloride content. According to this, there are color differences between the obtained materials and the starting zeolitic mineral (see Figure S1 of Supporting Information File 1), which are stronger in the materials obtained by Imp due to their higher content of residual salts.

Figure 3 also presents the Na, K, Ca, and Mg contents for the monometallic materials obtained from both treatments. Despite the previously mentioned observations, both types of materials (from IE and Imp) exhibit a reduction in Na, K, Ca, and Mg levels compared to ZSA, with this decrease being more pronounced in the materials obtained via IE. This suggests that in both treatments an ion exchange process occurred between the Ni²⁺ and Co²⁺ cations from solutions and the Ca²⁺, Na⁺, K⁺, and Mg²⁺ cations from the zeolite phases (mordenite and clinoptilolite-heulandite) contented within ZSA, as represented by Equation 1:

$$A_2^{n+} - Z_{(s)} + nM_{(aq)}^{2+} = M_n^{2+} - Z_{(s)} + 2A_{(aq)}^{n+},$$
 (1)



where $Z_{(s)}$ is the solid zeolite phase, and A^{n+} denotes the Ca^{2+} , Na^+ , K^+ , and Mg^{2+} cations within the zeolite. $M^{2+}_{(aq)}$ represents the Ni^{2+} and Co^{2+} cations from the solution.

Table 2 presents the quantities of Co²⁺, Ni²⁺, and Cl⁻ in equivalent moles per 100 g of each material, as well as the equivalent mole ratio of Cl⁻ anions relative to Co²⁺, Ni²⁺, and the Co²⁺-Ni²⁺ mixture. These values were determined from the elemental contents reported in Table 1 and Figure 3. It is evident that the equivalent mole ratios are different from unity, which suggests that these ions are not totally neutralizing their charges (i.e., 2 equivalents of Cl- per 1 equivalent of metal cation (see M²⁺ in Equation 1)). Moreover, in most cases they are less than unity, which indicates that a portion of the Co²⁺ and Ni²⁺ cations have not neutralized their positive charges with Cl⁻ anions. Instead, these cations likely neutralize their charge with negative charges from the zeolite framework, suggesting they occupy extra-framework cationic positions as compensation cations. This observation aligns with the earlier discussion that an ion exchange process occurred between these metal cations and the cations from the zeolite phases (as represented in Equation 1). For samples obtained by impregnation, the equivalent mole ratios are much higher mainly because of the large amount of chloride remaining on the surface. However, this does not mean that there are no Ni2+ and Co2+ cations occupying exchange positions. Note that chloride can also form salts with iron, sodium, and calcium cations leaving the zeolite, as proven by XRD.

In line with expectations for a natural zeolite, the studied materials exhibited hybrid type-I/IV adsorption isotherms (see

Figure S2 in Supporting Information File 1), indicating the presence of both microporous and mesoporous structures. The isotherms showed a hysteresis loop, which is associated with capillary condensation within the materials' pores and is expected to be more pronounced in HEU than in Mor. This can be attributed to the smaller channel diameter of HEU (maximum opening of $0.31 \times 0.75 \text{ nm}^2$) compared to Mor (maximum opening of $0.70 \times 0.65 \text{ nm}^2$). Additionally, the kinetic diameter of a nitrogen molecule (0.36 nm) is closer to the HEU channel diameter, further influencing the observed behavior.

Table 3 displays the specific surface area (SSA) and microporous volume ($V_{\rm micro}$) values determined for the obtained materials. It is evident that all materials acquired through Imp have lower SSA and $V_{\rm micro}$ values compared to those obtained via IE. This reduction is attributed to the presence of more adsorbed salts (chlorides) on the materials' surface, which is in agreement with the XRD results.

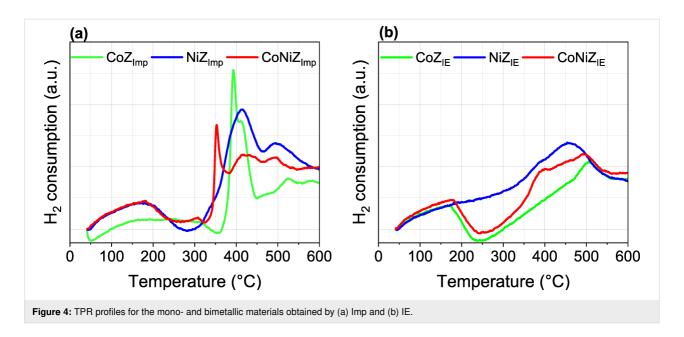
Conversely, the SSA and $V_{\rm micro}$ values for materials obtained by IE are higher than those of ZSA. This increase is consistent with expectations [17,18], as the ion exchange and water washing treatments effectively clean the materials' surface, thereby enhancing the available surface area and porosity (microporosity) for adsorption.

The temperature-programmed reduction (TPR) profiles of the materials obtained by both methods are shown in Figure 4. These profiles vary significantly depending on the used modification method. It has been reported that the cations supported on zeolites can be thermally reduced by hydrogen, which is

Table 2: Equivalent mole values of Co²⁺, Ni²⁺, and Cl⁻ per 100 g of material (equiv/100 g) and the equivalent mole ratio (Cl⁻/M²⁺) of Cl⁻ anions relative to metal cations (Co²⁺, Ni²⁺, and Co²⁺–Ni²⁺ mixture) of materials obtained by IE and Imp.

0.052 ± 0.003	_	$8.60 \pm 0.47 \times 10^{-4}$	0.0165
_	0.0438 ± 0.0033	$5.69 \pm 0.34 \times 10^{-4}$	0.0130
0.032 ± 0.0023	0.034 ± 0.0027	$1.90 \pm 0.13 \times 10^{-4}$	0.0287
0.086 ± 0.0067	_	0.1080 ± 0.0097	1.2558
_	0.092 ± 0.0071	0.0891 ± 0.0071	0.9684
0.039 ± 0.0026	0.029 ± 0.002	0.0541 ± 0.0037	0.7955
	0.032 ± 0.0023 0.086 ± 0.0067	- 0.0438 ± 0.0033 0.032 ± 0.0023 0.034 ± 0.0027 0.086 ± 0.0067 - 0.092 ± 0.0071	

Table 3: Specific surface area (SSA) and microporous volume (V_{micro}) of the ZSA and the materials obtained using IE and Imp. CoNiZ_{Imp} CoZ_{Imp} NiZ_{Imp} Sample ZSA CoZIE NiZ_{IE} CoNiZ_{IE} SSA (m²/g) 134 138 146 54 142 64 V_{micro} (cm³/g) 0.0316 0.0326 0.0015 0.0385 0.0067 0.0439 0.0139



influenced by the nature of co-cations present [19-22]. The TPR profiles display hydrogen consumption peaks at different temperatures, corresponding to the reduction of the nickel and cobalt cations arranged on the zeolite support. The TPR profiles of materials prepared via Imp show a greater number of hydrogen consumption peaks compared to those obtained by IE. Two peaks appeared centered at 170 °C and 500 °C for CoZ_{IE}, while CoZ_{Imp} exhibits hydrogen consumption in the 150-350 °C range and three additional peaks centered at 390 (intense), 410, and 515 °C. Similarly, NiZ_{IE} exhibits two peaks at 380 (low) and 450 °C, whereas NiZ $_{\mbox{\footnotesize Imp}}$ displays three peaks at 150, 410, and 500 °C. Regarding the bimetallic systems, CoNiZ_{IE} shows three peaks at 170, 380, and 500 $^{\circ}$ C, while CoNiZ_{Imp} shows five peaks at 170, 310, 350 (intense), 420, and 500 °C. In line with the elemental composition (Figure 3, Table 1 and Table 2), the textural parameters (Table 2) and previous discussions, hydrogen consumptions peaks can be attributed to the reduction of cobalt and nickel cations present as both isolated cations (compensation cations in extra-framework ionic positions) and chloride salts adsorbed on the zeolitic support. This distinction explains the differences between the TPR profiles. Thus, materials obtained by IE exhibit peaks associated only with isolated cations, while materials obtained by Imp show peaks linked to both isolated cations and chlorides.

Careful analysis of the profiles suggests that isolated $\mathrm{Co^{2+}}$ cations in extra-framework ionic positions undergo two hydrogen consumption events for their complete reduction, that is, one at 170 °C and another one in the 500–515 °C region. Similarly, isolated $\mathrm{Ni^{2+}}$ cations in these positions experience two reduction events, namely, one at 380 °C and another one in the 450–500 °C region. This behavior is consistent across both

modification methods and is observed in both monometallic and bimetallic systems. Beside this, the reduction of isolated cations is thermally favored in the bimetallic systems. Note that the intensity of the 380 °C peak, associated with isolated Ni²⁺ cation reduction, increases in bimetallic CoNiZ_{IE} compared to monometallic NiZ_{IE}. Furthermore, the reduction of isolated Co²⁺ cations that takes place at 510 °C in the monometallic CoZ_{IE} is shifted to a lower temperature (500 °C) in the bimetallic CoNiZ_{IE}. This suggests a mutual synergistic influence between Ni²⁺ and Co²⁺ cations with each facilitating the reduction of the other, likely due to a weakening of the interaction between isolated cations and the zeolite framework.

In bimetallic systems, Ni²⁺ and Co²⁺ cations compete for extraframework cationic positions in the zeolite phases. As a result, these cations occupy positions where their interaction with the zeolite framework is diminished compared to monometallic system, facilitating their reduction at lower temperature.

Catalytic test in citral hydrogenation

The main pathway of citral hydrogenation is illustrated in Figure 5, and the corresponding results are presented in Figure 6. Catalysts were tested as obtained, that is, without prior thermal activation. Although the conversion of citral after 3 h of reaction is relatively low for all catalysts, it indicates that the active sites are accessible to citral molecules. Overall, citral hydrogenation and the formation of citronellal are higher for catalysts containing nickel compared to those containing cobalt (Figure 6). This suggests that the most active catalytic sites for the conversion of citral to citronellal are associated with nickel species. Notably, the bimetallic CoNiZ_{IE} catalyst is the only one to show activity for the hydrogenation of citral to the unsatu-

rated alcohols geraniol and nerol (Figure 5), albeit in small quantities. In contrast, the bimetallic $CoNiZ_{Imp}$ shows lower citral conversion, with no formation of unsaturated alcohols detected. This discrepancy between the bimetallic catalysts may be attributed to differences in ionic species, as well as variation in specific surface area (Table 3) and nickel content (Table 1 and Figure 3). The $CoNiZ_{IE}$ catalyst has higher values for both parameters compared to $CoNiZ_{Imp}$. Additional factors contributing to these differences will be discussed later.

CHO
$$H_{2}$$
citronellal
$$CH_{2}OH$$

$$CH_{2}OH$$

$$CH_{2}OH$$
citronellol
$$CH_{2}OH$$

$$CH_{2}$$

Figure 5: Main pathways of citral hydrogenation.

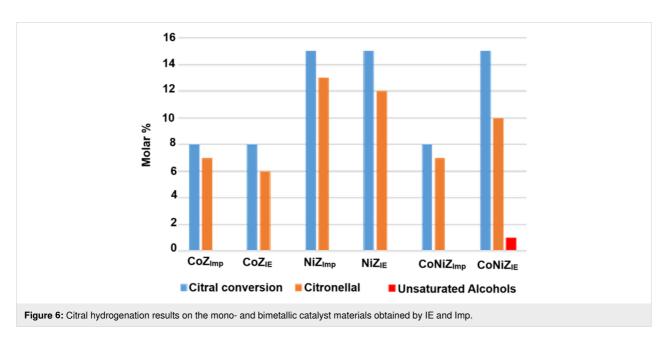
The catalytic activity of the $CoNiZ_{IE}$ catalyst in the selective hydrogenation of citral to unsaturated alcohols (geraniol and nerol) can be attributed to a synergistic interaction between cobalt and nickel species. These active species are likely associated with isolated cations or those formed during the hydrogenation process. For this reaction to proceed effectively, citral

and hydrogen must interact directly with the active catalytic centers, such as isolated $\rm Ni^{2+}$ cations located in extra-framework cationic positions in the zeolite channels. Given that citral has a molecular diameter of around 0.3 nm, it can enter the channels of clinoptilolite (maximum opening of 0.31 \times 0.75 nm) and mordenite (maximum opening of 0.70 \times 0.65 nm), particularly the latter because of the larger channel diameter. The diffusion of reactants (citral and hydrogen) through the zeolite channels is expected to be favored by a higher specific surface area and unobstructed channel entrances. This is consistent with the superior surface area of CoNiZ $_{\rm IE}$ and the presence of impurities of mixed metal chloride salts on the surface of CoNiZ $_{\rm Imp}$, as evidenced by elemental analysis, XRD, and TPR profiles.

In order to improve these results, the $CoNiZ_{IE}$ catalyst was subjected to in situ reduction at 500 °C for 2 h under H_2 flow prior to the catalytic test. However, this high reduction temperature resulted to be deleterious to the catalytic performance. While the catalytic performance of $CoNiZ_{IE}$ in the selective hydrogenation of citral to unsaturated alcohols remains low, the ability of this material to produce such alcohols indicates its potential. Future work will focus on optimizing the reduction temperature as lower reduction temperatures are expected to enhance performance. These findings will be addressed in subsequent studies.

Conclusion

A detailed investigation was conducted on zeolitic materials featuring mono- and bimetallic systems of nickel and cobalt, derived from natural zeolite rich in clinoptilolite and mordenite. These materials were prepared using Ni²⁺ and Co²⁺ chloride solutions through traditional ion exchange (IE) and impregna-



tion (Imp) methods. The Imp-prepared materials exhibited higher nickel and cobalt contents but also contained significant amounts of chlorides. Conversely, the IE-prepared materials showed negligible chloride content and larger specific surface areas. These differences were attributed to the substantial presence of chloride salts adsorbed on the surface of the Imp-prepared materials, as evidenced by XRD analysis. This also impacted the TPR profiles, with the Imp-prepared materials displaying hydrogen consumption patterns different from those of the IE-prepared materials. The TPR profiles further revealed that the thermal reduction of isolated Co²⁺ and Ni²⁺ ions (compensation cations in extra-framework ionic positions) was facilitated in the bimetallic systems, likely because of the synergetic interaction of multiple species and a reduced interaction between the cations and the zeolite framework. In line with these findings, the catalytic performance of the materials in the selective hydrogenation of citral showed marked differences. The most active catalytic sites for converting citral to citronellal were associated with nickel species. Among the catalysts, the bimetallic CoNi_{IE} material, prepared by IE, emerged as the most promising. It was the only catalyst to exhibit activity for the hydrogenation of citral to form unsaturated alcohols, suggesting a synergistic interaction between cobalt and nickel species. The active species are likely associated with isolated cations or those formed during the hydrogenation process.

This work highlights the potential of bimetallic $CoNiZ_{IE}$ materials as efficient catalysts for selective hydrogenation reactions, paving the way for further optimization and exploration of similar catalytic systems.

Experimental

Material and methods

Natural zeolite from the San Andrés deposit in Cuba, with a particle size range of 40–160 μm, was used. This zeolitic material consists primarily of mordenite and clinoptilolite-type zeolites (around 80%), along with minor accompanying phases (quartz, montmorillonite, feldspar) [23]. It underwent a purification process following a method similar to one described previously [18]. Its elemental chemical composition in oxide form is 65.7% SiO₂, 11.4% Al₂O₃, 3.4% CaO, 2.4% Na₂O, 1.3% K₂O, 1.1% MgO, and 2.6% Fe₂O₃. For simplicity, this purified zeolite is referred to as ZSA.

Monometallic (CoZ and NiZ) and bimetallic (CoNiZ) systems of nickel and cobalt were prepared from ZSA with NiCl₂, CoCl₂, and mixed NiCl₂/CoCl₂ solutions through both IE and Imp. The mixed NiCl₂/CoCl₂ solution was prepared by combining equal volumes of NiCl₂ and CoCl₂ solutions. The NiCl₂ and CoCl₂ used were reagent-grade, supplied by Sigma-Aldrich, St. Louis, MO, USA.

The IE processes were performed at 80 °C with reflux for 24 h, using a total of 2 milliequivalents of the corresponding cations (Ni $^{2+}$, Co $^{2+}$, and Ni $^{2+}$ /Co $^{2+}$) in solution per gram of ZSA and 1 g/10 mL solid/solution ratio. The solid phases were washed with distilled water to remove chloride ions and oven-dried at 110 °C.

The Imp processes were conducted using a total of 6% of corresponding metals (Ni, Co, and Ni/Co) per gram of ZSA and solutions with total metal contents of 0.2 mol/L. Both the ZSA and the solutions were heated to 80 $^{\circ}$ C before being mixed. After 24 h, the solid phases were separated, lightly washed with distilled water, and oven-dried at 110 $^{\circ}$ C.

Characterization

The elemental composition of ZSA and the modified materials from both treatments was determined using X-ray fluorescence analysis, performed with a ZETIUM PANalytical system. To characterize the specific surface area and pore structure of the materials, they were analyzed using Micro-Active software for TriStar II Plus instruments. Approximately 200 mg of each sample was degassed at 30 °C for 30 min and then at 250 °C for 4 h before surface area measurements via nitrogen adsorption at 77 K. The initial natural zeolite samples were also examined via powder X-ray diffraction (XRD) and scanning electron microscopy (SEM).

XRD patterns were recorded using a PW 1218 diffractometer (Philips, Almelo, Netherlands) equipped with a curved graphite monochromator and Cu K α radiation ($\lambda=1.5406$ Å). Data were collected at a scan speed of $2^{\circ}/\text{min}$ with a step size of 0.05° . SEM images were acquired using a FEI Nova NanoSEM 450 electron microscope. For this purpose, samples were mounted on holders and coated with a thin layer of gold prior to observation.

Temperature-programmed reduction (TPR) analyses were performed on an AutoChem 2910 instrument (Micromeritics, USA) equipped with a thermal conductivity detector (TCD). The procedure for TPR involved heating the sample in a 1.0 vol % $\rm H_2/Ar$ gas mixture at a flow rate of 30 mL/min, from room temperature to 600 °C, at a ramp rate of 5 °C/min. Hydrogen uptake was monitored using the TCD.

Catalytic test in citral hydrogenation

In a manner analogous to [24], the hydrogenation of citral was conducted in a 250 mL autoclave equipped with a magnetic stirrer and a temperature control unit. The catalysts (400 mg) were immersed in 90 mL of isopropanol and transferred into the autoclave. The reactor was first purged with nitrogen and then with hydrogen before raising the temperature to 130 °C. A mix-

ture of 3 mL citral and 10 mL isopropanol was then introduced into the reactor via a cylinder under 75 bar of hydrogen pressure. Time zero was considered at this point. During the catalytic test, the reaction was carried out under constant pressure using a pressure control system. After various reaction times, liquid samples were manually collected and analyzed by gas chromatography to determine conversion and selectivity values.

It is opportune to outline that preliminary experiments [25] conducted under various stirring conditions, catalyst loadings, and grain sizes confirmed the absence of both external and internal diffusion limitations.

Supporting Information

Supporting Information File 1

Additional figures.

[https://www.beilstein-journals.org/bjnano/content/supplementary/2190-4286-16-40-S1.pdf]

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Data Availability Statement

Data generated and analyzed during this study is available from the corresponding author upon reasonable request.

References

- Roth, W. J.; Opanasenko, M.; Mazur, M.; Gil, B.; Čejka, J.; Sasaki, T. Adv. Mater. (Weinheim, Ger.) 2024, 36, 2307341. doi:10.1002/adma.202307341
- Liu, M.; Miao, C.; Wu, Z. Ind. Chem. Mater. 2024, 2, 57–84. doi:10.1039/d3im00074e
- Sánchez-López, P.; Kotolevich, Y.; Yocupicio-Gaxiola, R. I.; Antúnez-García, J.; Chowdari, R. K.; Petranovskii, V.; Fuentes-Moyado, S. Front. Chem. (Lausanne, Switz.) 2021, 9, 716745. doi:10.3389/fchem.2021.716745
- Rodríguez-Iznaga, I.; Shelyapina, M. G.; Petranovskii, V. Minerals (Basel, Switz.) 2022, 12, 1628. doi:10.3390/min12121628
- Worathanakul, P.; Rakpasert, N. Int. J. Environ. Sci. Dev. 2016, 7, 885–888. doi:10.18178/jjesd.2016.7.12.899
- Zamaro, J. M.; Miró, E. E.; Boix, A. V.; Martínez-Hernández, A.; Fuentes, G. A. Microporous Mesoporous Mater. 2010, 129, 74–81. doi:10.1016/j.micromeso.2009.08.035
- Ramírez-Garza, R. E.; Rodríguez-Iznaga, I.; Simakov, A.; Farías, M. H.; Castillón-Barraza, F. F. Mater. Res. Bull. 2018, 97, 369–378. doi:10.1016/j.materresbull.2017.09.001
- Jiang, B.; Zhao, S.; Wang, Y.; Wenren, Y.; Zhu, Z.; Harding, J.; Zhang, X.; Tu, X.; Zhang, X. Appl. Catal., B 2021, 286, 119886. doi:10.1016/j.apcatb.2021.119886
- Rodríguez-Iznaga, I.; Petranovskii, V.; Chávez-Rivas, F.; Shelyapina, M. G. *Inorganics* 2022, 10, 34. doi:10.3390/inorganics10030034
- Rodríguez-Iznaga, I.; Petranovskii, V.; Castillón-Barraza, F. F.; Fuentes-Moyado, S.; Chávez-Rivas, F.; Pestryakov, A. *Materials* 2023, 16, 221. doi:10.3390/ma16010221
- Stolle, A.; Gallert, T.; Schmöger, C.; Ondruschka, B. RSC Adv. 2013, 3, 2112–2153. doi:10.1039/c2ra21498a
- Chung, S.-H.; Park, Y.-M.; Kim, M.-S.; Lee, K.-Y. Catal. Today 2012, 185, 205–210. doi:10.1016/j.cattod.2011.08.011
- Hong, U. G.; Kim, J. K.; Lee, J.; Lee, J. K.; Song, J. H.; Yi, J.; Song, I. K. Appl. Catal., A 2014, 469, 466–471. doi:10.1016/j.apcata.2013.10.029
- Popova, M.; Dimitrov, M.; Boycheva, S.; Dimitrov, I.; Ublekov, F.; Koseva, N.; Atanasova, G.; Karashanova, D.; Szegedi, Á. *Molecules* 2024, 29, 99. doi:10.3390/molecules29010099
- Taghavi, S.; Mäki-Arvela, P.; Vajglová, Z.; Peurla, M.; Angervo, I.; Eränen, K.; Ghedini, E.; Menegazzo, F.; Zendehdel, M.; Signoretto, M.; Murzin, D. Y. Catal. Lett. 2023, 153, 2674–2692. doi:10.1007/s10562-022-04178-x
- 16. Giordani, M.; Ballirano, P.; Pacella, A.; Meli, M. A.; Roselli, C.; Di Lorenzo, F.; Fagiolino, I.; Mattioli, M. Minerals (Basel, Switz.) 2022, 12, 627. doi:10.3390/min12050627
- 17. de la Nuez Pantoja, E. Y.; Rodríguez Iznaga, I.; Rodríguez Fuentes, G.; Petranovskii, V.; Martínez García, A.; Calvino Gámez, J. J.; Goma Jiménez, D.; Cauqui, M. Á.; Rivero González, L. A.; Collazo García, O. *J. Inorg. Organomet. Polym. Mater.* 2024, 34, 2522–2542. doi:10.1007/s10904-023-02991-4
- Chávez Rivas, F.; Rodríguez-Iznaga, I.; Berlier, G.; Tito Ferro, D.; Concepción-Rosabal, B.; Petranovskii, V. Catalysts 2019, 9, 866. doi:10.3390/catal9100866

- Herd, A. C.; Pope, C. G. J. Chem. Soc., Faraday Trans. 1 1973, 69, 833–838. doi:10.1039/f19736900833
- Suzuki, M.; Tsutsumi, K.; Takahashi, H.; Saito, Y. Zeolites 1989, 9, 98–103. doi:10.1016/0144-2449(89)90056-0
- 21. Suzuki, M.; Tsutsumi, K.; Takahashi, H.; Saito, Y. Zeolites 1988, 8, 381–386. doi:10.1016/s0144-2449(88)80175-1
- Peron, D. V.; Zholobenko, V. L.; Rodrigues de la Rocha, M.;
 Oberson de Souza, M.; Feris, L. A.; Marcilio, N. R.; Ordomsky, V. V.;
 Khodakov, A. Y. J. Mater. Sci. 2019, 54, 5399–5411.
 doi:10.1007/s10853-018-03250-5
- Selvam, T.; Schwieger, W.; Dathe, W. Clay Miner. 2014, 49, 501–512. doi:10.1180/claymin.2014.049.4.01
- Di, X.; Lafaye, G.; Especel, C.; Epron, F.; Qi, J.; Li, C.; Liang, C.
 ChemSusChem 2019, 12, 807–823. doi:10.1002/cssc.201802744
- Lafaye, G.; Micheaud-Especel, C.; Montassier, C.; Marecot, P.
 Appl. Catal., A 2002, 230, 19–30. doi:10.1016/s0926-860x(01)00980-2

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Mechanical stability of individual bacterial cells under different osmotic pressure conditions: a nanoindentation study of *Pseudomonas aeruginosa*

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Full Research Paper

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Abstract

Nanomechanical maps to test the mechanical response of the outer envelope of *Pseudomonas aeruginosa* were obtained utilizing atomic force microscopy in force–volume mode in the low range of loading forces when exposed to hypotonic (Milli-Q water), isotonic (PBS), and hypertonic (0.5 M NaCl) solutions. Imaging and mechanical testing showed that bacteria are highly resilient to deformation and can withstand repetitive indentations in the range of 500 pN. Analysis of force spectra revealed that although there are differences in the mechanical response within the first stages of nanoindentation, similar values in the slopes of the curves reflected a stable stiffness of about $k_B = 20$ mN/m and turgor pressures of $P_t = 12.1$ kPa. Interestingly, a change in the nonlinear regime of the force curves and a gradual increase in maximal deformation by the AFM tip from hypotonic to hypertonic solutions suggest a softening of the outer envelope, which we associate with intense dehydration and membrane separation between inner and outer envelopes. Application of a contact mechanics model to account for the minute differences in mechanical behavior upon deformation provided Young's moduli in the range of 0.7–1.1 kPa. Implications of the presented results with previously reported data in the literature are discussed.

Introduction

Pseudomonas aeruginosa (PA) is a Gram-negative bacterium belonging to the *Pseudomonas* genus. It is well known for its versatility and adaptability in various environments, as it can be found in multiple habitats, including soil, water, plants, and animals. It can also prosper in artificial environments such as metal or plastic pipes and medical devices [1-5]. Although PA is part of the normal microbiota of the skin and mucous membranes of many healthy individuals, it can cause serious opportunistic infections in the respiratory and urinary tracts and during wound healing in people with weakened immune systems or in hospitalized patients [6-8]. As a Gram-negative bacterium, PA is characterized by a distinctive cell wall structure constituted by a thin peptidoglycan layer enclosed by an outer membrane which contains lipopolysaccharides (LPS) [9-11]. The outer membrane also contains numerous proteins, lipoproteins, and channels contributing to its selective permeability [12,13]. Also integrated into the outer membrane, specific tension-activated channel proteins are responsible for the osmoregulation of the membrane envelope and its protection from threatening conditions such as severe osmotic downshocks, which can lead to an excessive increase in the membrane tension resulting in rupture [14,15]. The mechanosensitive (MS) family of channel proteins have been identified as the main efflux pumps required by PA to regulate the exit of osmolytes and reduce the membrane tension to acceptable life-compatible values. Therefore, the type and density of these MS channels triggered at different threshold values of membrane tension determine the survival capacity of the bacteria under drastic changes in osmotic pressure [16-20]. PA is also known to use other channel proteins to overcome the attack by antibiotics via rapid extrusion of the uptaken drugs, which confers remarkable resistance to this pathogen [21-34]. To hinder this difficulty, natural and synthetic molecular inhibitors with high binding affinity towards these channel proteins have been proposed [25-32]. Likewise, the application of silver and copper nanoparticles to block these molecular pores has been reported for PA and other bacteria as well [33-37]. In the latter cases, a direct consequence of this highly controlled membrane permeability is that membrane tension and rigidity are two intertwined physical parameters with a dynamic behavior dictated by the internal turgor pressure of the bacteria during swelling or plasmolysis. Therefore, understanding the dynamics of their mechanical response due to changes in external conditions or exposure to specific molecular agents is critical in generating strategies to control their undesired propagation.

Atomic force microscopy (AFM) is a powerful, sensitive technique that scans the surface topography of a sample with an

ultra-sharp tip while monitoring the interaction forces between this tip and the sample at the nanoscale. The force applied by the AFM tip on the sample is controlled by monitoring the deflection of an extremity of a micrometer mechanical lever onto which the tip is attached. In the study of pathogens, AFM excels in providing high-resolution topographic images while measurements are performed in solution in a fluid chamber under controlled environmental conditions. Thus, critical structural changes on the lifestyle of the pathogen can be investigated [38-42]. Beyond imaging, AFM force spectroscopy capabilities are essential to extract material properties of the investigated sample [11,43,44]. The force-volume (FV) mode allows for mapping surface physical parameters of the analyzed sample via controlled nanoindentations on a pre-programmed grid on the sample surface [44-47]. In conventional scanning modes (e.g., contact and tapping) the changes in displacement by the piezoelectric element are gathered in order to maintain a constant cantilever deflection or amplitude while a surface is scanned line by line Conversely, in FV quantitative information is extracted after analysis of a force-separation curve obtained from a performed nanoindentation. Therefore, the preprogramed grid in FV defines the amount of information (number of nanoindentations per scanned line) taken from the sample surface and also its resolution. Typical parameters obtained using the FV mode are the height, stiffness, adhesion, elasticity modulus, and dissipation of the sample. The mentioned parameters derived from force-separation curves are analyzed and mapped in real time as shown below in Figure 1. In the present study, we investigated the mechanical response of the outer membrane of PA at the single bacterium level in FV mode when exposed to relevant osmolarity conditions. Of particular interest was the extraction and comparison of nanomechanical maps obtained in the low range of loading forces to quantify its morphology, membrane stiffness, Young's modulus of elasticity and adhesion when PA was tested in hypotonic (Milli-Q water), isotonic (0.1 M phosphate buffered solution), and hypertonic (0.5 M NaCl) solutions.

Materials and Methods

Bacterial culture

The strain of *Pseudomonas aeruginosa* (ATCC[®] 27853 [™]) was reactivated by transferring it to Mueller–Hinton broth and incubating it for 24 h at a temperature of 37 °C. Subsequently, it was cultured on Mueller–Hinton agar and again incubated for 24 h under the same conditions. Isolated colonies of PA were collected using a calibrated loop and suspended in 5 mL of phosphate buffer supplemented with 50 µL of Mueller–Hinton broth. The suspension was vortexed for 10 s before its preparation for AFM analysis.

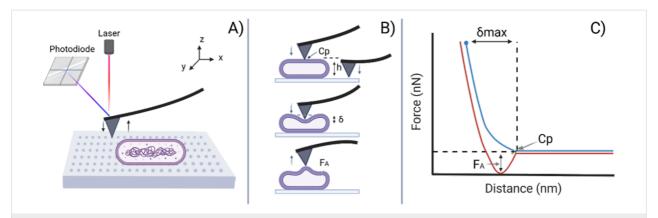


Figure 1: Schematic representation of the force—volume approach to quantify nanomechanical interactions with the atomic force microscope. A) A pre-selected grid that indicates the multiple specific points the AFM tip follows to perform individual indentations with a predefined maximal loading force. B) Drawing that shows how the tip indents on the sample surface in approach/retraction cycles, from which quantitative information such as height, stiffness, Young's modulus, and adhesion are extracted. In C), a schematic illustration of a representative force—separation curve taken on a deformable sample shows the profile of the interaction of the AFM tip with the sample surface and its contact point, maximal deformation, and adhesion force. Created in BioRender. García-torres, L. (2025) https://BioRender.com/zpchrmj. This content is not subject to CC BY 4.0.

Sample preparation for atomic force microscopy

For AFM, the sample was initially firmly adhered to a substrate for subsequent scanning, with the tip first used to identify the objects of study on the surface. In this case, PA in suspension was adhered to a solid mica substrate, previously cleaved with regular adhesive tape, and coated with 5 µL of the cationic polymer poly-L-lysine (PLL, MW 40 kDa, Sigma-Aldrich). The drop of PLL was not allowed to dry but incubated for 10 min, and then the surfaces were repeatedly rinsed with Milli-Q water to allow the formation of a thin film. This coating promotes bacterial adhesion through short-range electrostatic and hydrophobic interactions. Before deposition on the PLL-coated substrate, the bacterial suspension was centrifuged at 2500 rpm for 3 min, and the resulting supernatant was removed. The bacteria were then resuspended in 150 µL of PBS to increase their concentration and before the previous deposition of 5 µL on the PLL-coated substrate. A liquid cell was assembled to measure changes in bacterial membrane rigidity response when transitioning between different solutions. Once inside the fluid cell chamber, bacteria were imaged and tested in PBS solution and then exposed to the desired osmolarity conditions for investigation. The applied solutions were ultrapure water (Milli-Q), phosphate-buffered saline solution (PBS), and 0.5 M sodium chloride. Bacteria were first analyzed in tapping mode to determine their location and observe their morphology. Following this, the FV mode was used to create maps, and indentations at higher force loads were also performed using the point-andshoot tool.

Atomic force microscopy measurements

The surface analysis and mechanical characterization of living bacteria was performed with an AFM Multimode 8 from Bruker with a NanoScope V controller operated in fluid conditions throughout all experiments, using a pre-assembled fluid chamber with the appropriate solutions within a sealed O-ring. The instrument was operated in contact mode using MLCT probes from Bruker, cantilever D with a nominal spring constant of 0.03 N/m, and tip radius of 20-60 nm as provided by the manufacturer (Bruker), to study the morphological features of bacteria under different tested conditions. The deflection set point was adjusted during the measurement to optimize imaging conditions. Nanoindentations were performed using a maximum loading force of 500 pN for each nanoindentation applied in a pre-programmed grid of 48 points per line to quantify surface interactions and the mechanical response of bacteria. This allowed us to simultaneously acquire maps with enough resolution on the sample surface to identify individual bacteria and obtain the mechanical response from the membrane rigidity within a reversible regime. The advantage of mapping bacteria at low loading forces is that surface interactions between the tip and cell membrane can be analyzed, and information related to its adhesion or surface charge could be revealed with minimum damage to the mechanical integrity of the cell membrane. Before each experiment in FV mode, the cantilever was calibrated using the thermal-noise method which is integrated in the multimode AFM software and is briefly described next. First, the optical-lever sensitivity (OLS) of the cantilever was extracted. A force-distance curve was taken on a clean (previously cleaved) mica surface and a linear fit to the approach trace within the region following contact was performed to obtain the OLS from its slope. Afterwards, the cantilever was withdrawn above the surface and a thermal tune was performed to obtain the spectrum of the cantilever. A Lorentzian fit was then performed to the obtained peak and finally the cantilever constant was calculated.

To take into account any potential change in the dimensions of PA at different osmolarities, the cross section tool from the NanoScope software was used to measure the long (length) and short (width) axes of the bacteria after being tested in FV mode. At least 20 bacteria were measured for each investigated condition. In AFM, the half-width distance is usually reported to take into account the influence of the AFM tip due its curvature close to the apex. A single measurement was taken along the length of each bacteria, while three measurements were performed and averaged to take its width. This is presented below in Figure 2A and Figure 2B.

All obtained images, FV maps, and force-distance curves were analyzed with the NanoScope analysis software 1.7 from Bruker. Once data for nanomechanical mapping at low loading forces was extracted, it was also attempted to produce maps at high loading forces of about 3 nN, as described below in the results section.

Results

Before any nanoindentation measurements were performed, bacteria were first localized on the surface of poly-L-lysine-

coated mica. Supporting Information File 1, Figure S1 shows some 3D images of PA taken in contact mode when in PBS. Bacteria and the substrate can be observed. For a straightforward interpretation using quantitative nanomechanics in FV mode, the object to be mechanically tested must be clearly distinguished from a rigid substrate used as background. In this case, the thin layer of PLL (1-2 nm) on mica is expected to provide such background. Regions especially crowded with bacteria were intentionally avoided when applying FV. The capacity of AFM as a quantitative analysis tool when operated in FV mode is illustrated in Figure 1. Once a region has been selected with conventional AFM imaging, a grid is defined with a special resolution of points per line and tip velocity upon approach/retraction (see Figure 1A). Each approach/retraction cycle to a maximum load generates a force-separation curve analyzed in real time by the software. A color-coded map is obtained according to the physical parameters extracted from the curve, as shown in Figure 1B,C.

Figure 2 shows an FV image of the topography or height channel for several individual PA bacteria on the substrate. An estimation of the size of PA by AFM was made using the cross-

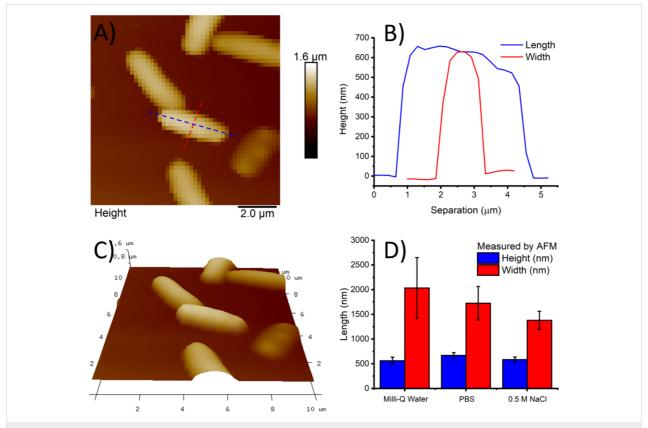


Figure 2: Mapping living bacteria with force—volume AFM. A) 2D height image of *Pseudomonas aeruginosa* obtained in FV mode where pixels indicating the location of single nanoindentations can be easily discerned. B) Cross-section profiles taken from both main axes of the bacteria shown in A) with dashed lines for length (blue) and width (red). C) 3D reconstruction of the image shown in A) shows how FV can also accurately represent the topography of the sample. D) Dimensions of PA obtained from AFM in different measured osmolarity conditions.

section tool along both axes to obtain its length, width, and height, as shown in Figure 2A and 2B. With a resolution of 48 points or pixels per line, FV can also provide accurate morphological measurements for bacteria, as shown in the 3D reconstruction in Figure 2C (see also Supporting Information File 1, Figure S2). These measurements were taken for bacteria in the three investigated conditions (i.e., Milli-Q water, PBS, and 0.5 M NaCl) and the results are shown in Figure 2D. Larger values for the length and width of the bacteria are expected in AFM measurements when compared to other experimental techniques such as optical microscopy or electron microscopy. This is due to the fact that the finite size of the tip apex intro-

duces an overestimation in the lateral dimensions of the imaged object but not of its height, where AFM excels in accuracy. From the plot in Figure 2D, we can observe that while its height remains around the same value of 500–700 nm, the change of its width shows a slight trend towards shrinkage of PA from hypotonic to hypertonic conditions. This might be a consequence or a first sign of readjustment of the cell volume when the adhered bacteria are abruptly exposed to different osmotic pressures.

In Figure 3, maps for three of the main physical parameters, height (h), stiffness (k), and Young's modulus (Y) obtained in

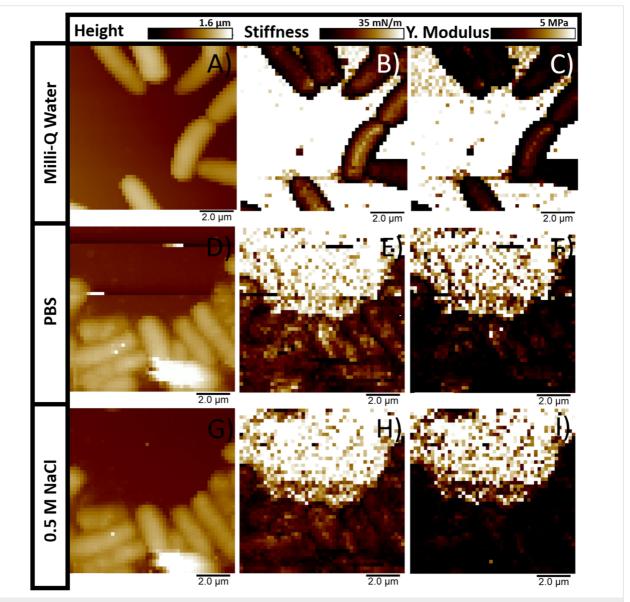


Figure 3: Nanomechanical maps of PA were obtained in FV mode for the three investigated conditions with a maximal loading force of 500 pN. The physical parameters shown are height (A, D, G), stiffness (B, E, H), and Young's modulus (C, F, I). As observed, PA is identified on the surface in contrast with the background due to its different interaction with the AFM tip under the same loading force. Measuring conditions are given as rows, while colored scales for each extracted parameter are shown as columns.

FV, are shown as columns, while the measuring conditions are given as rows. For the height channel in the first column, the identification of individual bacterium is possible even when they are aggregated. As expected, PA appears brighter than the substrate plane since the offset of the color-coded scale has been set for the best contrast and clarity for both. Bacteria appear dark to a brighter background for stiffness and Young's modulus. In these cases, the offset in the color-coded scales has been set to 0-35 mN/m and 0-5 MPa, respectively, for an easier and more direct interpretation of the obtained values. The stiffness is obtained from the slope of the curve during compression by the AFM tip from the point of contact (C_p) until it reaches its maximum predefined loading force. For this reason, the rigidity of the underlying substrate can be assumed to behave as an impenetrable substrate for the range of the applied loads. In the present case, the rigidity of muscovite mica represents a good approximation for the last assumption. However, the thin polymer layer of PLL applied to enhance the adhesion of PA is expected to introduce a certain but rather small degree of mechanical resistance. Visually, from the maps in the column for stiffness, it is difficult to discern a difference in the brightness on the surface of PA from the hypotonic (Milli-Q water) to isotonic (PBS) and finally to the hypertonic solution (0.5 M NaCl). Nonetheless, in the case of Young's modulus, bacteria in the maps seem to become slightly darker with the increase in ionic strength.

Contrary to stiffness, the map for Young's modulus is extracted after a mechanics model has been chosen in the analysis software and the appropriate parameters between the AFM tip and sample have been given. Although the apex of the tip used in this case is not a perfect cone, and the radius of the tip is about 10% of that of PA, analysis of the force response obtained in the force curves suggests that the Sneddon model provides a better description for the induced deformation than that of the Hertz model for the case when a spherical tip indents on an elastic planar half-space. Under the frame of the Sneddon model, a rigid conical indenter pushes a planar elastic semi-space characterized by a particular elastic modulus Y. The Sneddon model then provides a relation between the applied force load (F), the induced deformation (δ) , and the modulus of elasticity (Y) of the tested material in the following way:

$$F(\delta) = \frac{2Y \tan(\alpha)\delta^2}{\pi(1-\upsilon^2)},$$

with α as the half angle distended by the apex of the AFM tip, and ν is a unitless factor known as the Poisson ratio, which takes values between 0 and 0.5. In the present study, we use $\nu = 0.49$, as usually reported for biological samples [48,49].

Figure 4A shows the approach component of representative force-separation curves obtained on PLL and PA for each investigated condition for a maximal loading force of 500 pN. These curves show the mechanical response from the polymer layer on the mica surface and the outer membrane of PA. From the black line, it is easy to observe how the tip, upon contact, almost immediately attains full compression on the PLL layer, and no further deformation can be observed as the slope of the curve almost reaches verticality. The necessary deformation depth required to fully compress the PLL layer was 1-3 nm. An obvious difference in the force response is obtained on PA (blue, red, and green lines). For these low regimes of loading forces, it can be observed that the force response is nonlinear within the first 70 pN for Milli-Q water and about 100 pN for PBS. For 0.5 M NaCl, the curve remains nonlinear up to about 250 pN. For the maximal applied loading force of 500 pN, a maximal attained membrane deformation δ_{max} occurred at 55, 80, and 145 nm for Milli-Q water, PBS, and 0.5 M NaCl, respectively. Consequently, we observed a clear increase in the maximal induced deformation with an increase in the osmolarity. For this extent of induced deformation, the outer envelope of PA showed a reversible behavior in the mechanical response. Figure 4B shows a schematic representation of the AFM tip indenting on the outer membrane of PA and its maximum deformation δ_{max} . A fit to the obtained force response using the Sneddon model in the NanoScope software for the curves shown in A) is given in Supporting Information File 1, Figure S3. These representative force curves have been selected from nanoindentations captured along the middle region of the longer axis of the bacteria in FV maps. This is of central importance since it was observed that the force response drastically changes according to the direction of the scanning in relation to the orientation of the bacteria. Especially at the edges, the influence of the finite 3D size and angle of the AFM tip became evident because indentation can take place perpendicularly, along the longer axis of PA, or in an intermediate diagonal direction of the bacterial body, which ultimately significantly alters the profile of the obtained force response. In Figure 4C,D, histograms from FV maps for the obtained values for k and Y are given for the case of Milli-Q water. The rest of the data is given in Supporting Information File 1, Figure S4. Although expected, a main feature in the obtained histograms for k and Y for all measured samples was the presence of bimodal distributions, which reflects the mechanical response from a homogeneous substrate coated at some percentage with bacteria. Therefore, one distribution corresponded to the mechanical response from the PLL-coated mica, while the other to PA. In the case of stiffness, both distributions appear Gaussian, with the substrate stiffness (k_s) at higher values than that for the bacteria (k). Nevertheless, for Young's modulus, only the distribution arising from the substrate (Y_s) is Gaussian, while the one

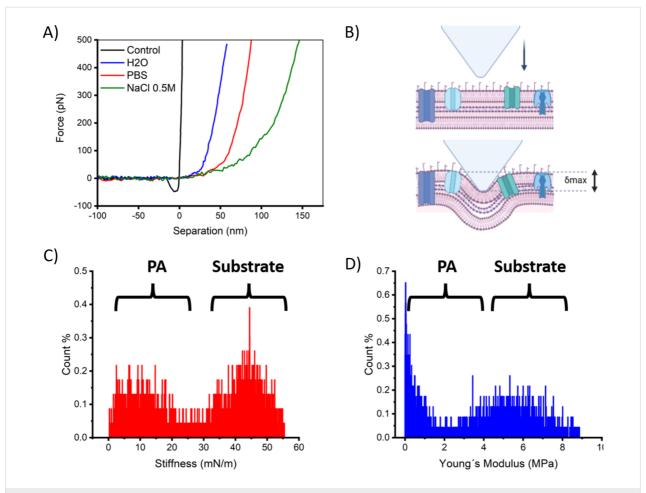


Figure 4: Degree of compression by nanoindentations in FV mode at low loading forces of 500 pN. In A) representative force—separation curves were obtained from the force response of PA indented under different tested conditions. Force curves were taken from the middle region of the bacteria to minimize variations arising from the finite size of the AFM tip interacting at the edges of the bacterial body. A control curve next to the bacteria is also shown as a reference for deformation (black line). In B), a drawing illustrates the outer membrane compression by the AFM tip (Created in BioRender. García-torres, L. (2025) https://BioRender.com/qynn0nv. This content is not subject to CC BY 4.0.). In C) and D), histograms for the stiffness and Young's modulus values obtained from a map taken in FV mode in Milli-Q are shown, respectively.

for the bacteria (Y) at lower values resembles a lognormal distribution. Supporting Information File 1, Figure S4 fits the obtained experimental data with Gaussian and lognormal distributions to obtain mean values. The obtained δ_{max} shows that in this low range of loading forces, the membrane rigidity is tested beyond the thickness of the bacterial outer cell wall, which, according to cryo-electron tomography, is about 25-30 nm [50]. In Table 1, the mean value obtained for the stiffness and Young's modulus from the distributions are given and compared with mean values of k_{PA} and Y_{PA} intentionally selected from the middle regions of PA. The corresponding mean values for the maximum deformation or penetration δ_{max} , attained by the AFM tip during compression on the central region of the bacteria, are also given. Table 1 shows that the mechanical resistance to deformation reflected in the stiffness remained almost unchanged when the mean values obtained from distributions for the different measuring conditions were compared. However, they differ from the mean values obtained from the middle region of bacteria. This accounts for the mechanical stability of the outer membrane of PA once a certain osmotic pressure is set. We can observe that mean values for k and Y obtained from the distributions were smaller than those obtained from the middle region of PA. This comparison also indicates how, in a nanoindentation map, the distribution overshadows the mean value taken where the geometrical conditions are usually preferred during mechanical testing since their number is just a small fraction of the entire set of indentations performed along a map. For mean values obtained along the central region of the bacteria, the stiffness also remained relatively unchanged for the different tested conditions. Still, we see a slight trend towards lower values from hypotonic to hypertonic solutions for the Young's modulus. This observation will be discussed below in the discussion section regarding the application of the mechanics model. Opposite to the Young's modulus,

Table 1: Mean v	ralues for stiffness and You	ung's modulus obtained fro	om distributions in FV map	os and from the central reg	ion of PA.
Solutions	k (mN/m) from distributions	Y (MPa) from distributions	<i>k</i> _{PA} (mN/m) from the middle region of PA	Y _{PA} (MPa) from the middle region of PA	δ_{max} (nm) from the middle region of PA
Milli-Q	8.83 ± 0.46	0.51 ± 0.07	21.3 ± 4.9	1.13 ± 0.38	51.74 ± 10.94
PBS	9.32 ± 3.69	0.5 ± 0.08	17.6 ± 5.28	0.93 ± 0.39	88.5 ± 17.47
0.5 M NaCl	7.4 ± 0.17	0.14 ± 0.01	23.2 ± 5.36	0.69 ± 0.44	92.15 ± 27.85

the mean maximal induced deformation upon compression δ_{max} increases in the presence of a higher salt concentration.

Supporting Information File 1, Figures S5 and S6 show maps for the adhesion force between the AFM tip interacting with the substrate and bacteria. In Supporting Information File 1, Figure S5A–F a clear contrast in the adhesion force between the bacteria and the substrate can be observed for the case of Milli-Q water, where it was possible to quantify the adhesion force to the AFM tip as shown in the cross-sections in C) and F). When salt is introduced into the system, the contrast in the map is almost entirely lost, strongly suggesting that the electrostatic surface forces have been screened by the presence of ions, as shown in Figure S6 of Supporting Information File 1.

Finally, in Figure 5A and 5B, it was demonstrated how applying repetitive compressions via nanoindentations at high loading forces in hypertonic solution led to the destruction of previously imaged bacteria. In Figure 5A, a region heavily coated with PA was mapped in 0.5 M NaCl with FV with 500 pN as the maximum loading force, and in Figure 5B, the same region was imaged under the same conditions but with a maximal loading force of 3000 pN. A complete loss of identifiable bacterial structures is obtained. These results demonstrate the incapacity of PA to maintain the structural integrity of the outer membrane when exposed to larger loads within the time scale of the present experiment, as shown in the force curve in Figure 5C. The maximal force that the membrane could withstand before rupture was $2.1 \pm 0.5 \, \text{nN}$.

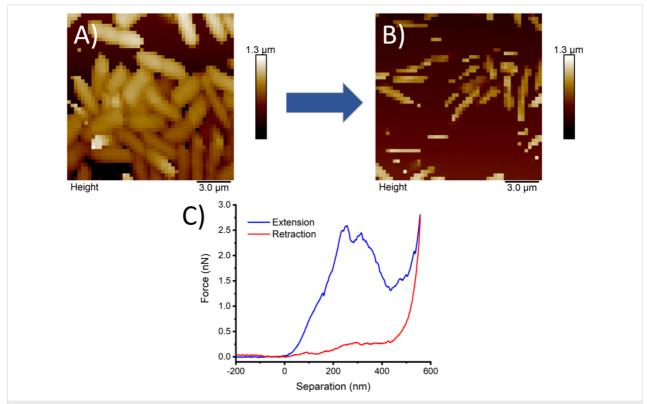


Figure 5: Force–volume maps were obtained at low A) and high B) loading forces in the same scanned region. The image clearly shows the entire destruction of previously mapped bacteria. C) A representative force–separation curve on PA where a drastic drop in the force is detected at about 2.5 nN.

Discussion

Based on the FV mode of AFM, the results presented here test the mechanical resilience of bacteria as a whole under different conditions of external osmotic pressure with repetitive nanoindentations. It was observed that histograms for the obtained physical parameters from nanoindentation maps provide a full picture of the landscape being imaged. However, they can obscure relevant mean values when there is a geometry-dependent interaction between the AFM tip and the sample. From the many individual nanoindentations across the bacteria surfaces along each scanned line, only a few test the central region of interest, where mechanics models can better estimate the rigidity or elastic modulus. The different orientations of the rodshaped morphology of PA on the substrate while being scanned by the quasi-conical AFM tip with a particular apex half-angle provide a broader spectrum of values on the nanoindentation map. Consequently, meaningful data is present but hidden within the statistical set. For this reason, mean values extracted from histograms were compared with data taken by point nanoindentations from the central region of the long axes of bacteria. Further analysis of these force curves is the focus of the following discussion. Upon compression by the AFM tip, PA showed two main regimes directly after contact. A first nonlinear regime following the contact point is commonly associated with the elastic response of the outer envelope. At the same time, for deeper deformations, a linear behavior in the force curve reflects the magnitude of internal turgor pressure P_t [51]. It is worth mentioning that for our case, only a handful of force curves, particularly in Milli-Q, presented a long tail upon approach, which frequently makes it difficult to identify the position of the contact point. This has been previously reported in other nanoindentation studies on bacteria, and its origin has been related to the action of repulsive electrostatic forces with long Debye screening lengths [44,52]. Our results obtained for the changes in the mechanical response of PA in the low deformation range under different external osmotic pressures are summarized in the parameters of stiffness (k), Young's modulus (Y), and maximal induced deformation (δ_{max}) (see Table 1). These results suggest that the mean values for stiffness remain almost unchanged for the tested conditions. However, mean values obtained for Young's modulus show a shallow trend to decrease with higher osmolarities, particularly when point nanoindentations were used. On the other hand, a clear increase in the penetration depth δ_{max} is observed for higher osmolarities. This behavior of k and Y can be understood, considering the observed change in curvature in the force curves at the beginning of the indentation process and before the curve approaches linearity. The Sneddon model used here to extract the elasticity modulus is sensitive to these changes in curvature even when the slope within the linear regime at the final stage of the trace remains almost unchanged. The increase in δ_{max}

(Figure 4a) from 55 to 145 nm accounts not only for this susceptibility of the outer envelope to deform at high osmolarities but also shows how the slope barely changes before it reaches the force threshold of 500 pN. These results are interpreted as follows. For the three investigated osmolarity levels, bacteria do not seem to be disrupted or damaged, at least from a morphological point of view when imaged in the low regime of loading forces or from its mechanical response to external deformation as given in the force curves. Consequently, these results suggest that in each tested solution, the tension exhibited by the outer envelope reflects the differences in the internal osmotic pressure built within the bacterial cell wall once the corresponding protein channels mitigate any threatening conditions to the integrity of the membrane (e.g., exceeding internal pressures due to drastic osmotic downshocks or shriveling caused by sudden shrinkage of the membrane in hypertonic solution). During exposure to a hypotonic solution, membrane lysis by osmotic shock is prevented by the rapid reaction of the MS channels, which act as safety valves which open as soon as the internal P_t causes an increase in the tension of the envelope above a specific threshold value. Then, it rapidly releases contained osmolytes to deflate the internal pressure and avoid membrane rupture. Using forward light scattering experiments, Çetiner et al. reported that the time required for PA to reach an equilibrium state after osmolyte release when exposed to drastic osmotic downshocks was about 150 ms [14]. In our experiments, PA was tested in different conditions after the solution was exchanged in situ in the liquid chamber of the AFM and allowed to settle. This time was about 5 min; therefore, is quite above the reaction time of PA to complete the osmolyte release. In our study, measurements were performed once these tensiontriggered channels had already dissipated the exceeding internal pressure and reduced membrane tension below the activation threshold. On the other hand, a pronounced efflux of water leaves the bacterial body in hypertonic solutions. It drastically reduces the internal P_t , resulting in severe dehydration of macromolecules inside the bacteria and the inner membrane envelopes. Lewenza et al. reported the treatment of PA with high salt concentrations and showed that PA develops plasmolysis bays due to shriveling in hypertonic solutions [53]. These bays are generated by a separation between the inner and outer membrane envelopes due to water depletion. As a protective countermeasure of PA against osmotic upshock to avoid collapse by shrinkage, it is known that cationic ions such as K⁺ are imported by special protein transporters and accumulated by bacteria to maintain homeostasis against hazardous external concentrations of sodium ions [54]. Also, other accumulated or internally synthesized osmoprotectants such as N-acetylglutaminylglutamine (NAGGN) and glutamate contribute to its survival under critical hypertonic solutions, while hydrophilins are also believed to confer protection [55]. Analysis of our force

curves with maximal loading forces of 500 pN in hypotonic and hypertonic solutions yielded mean values for the maximum penetration depths (δ_{max}) of about 52 and 92 nm, respectively. In contrast, for PBS, an intermediate value was obtained. Considering the above information regarding the states of swelling or plasmolysis, our results for the stiffening and softening of the outer envelope can be explained in terms of the internal pressure P_t . Although membrane rigidity is an intrinsic property of the two-dimensional material, membrane tension is not. Instead, membrane tension depends on the forces acting on the membrane, which is directly influenced by the internal turgor pressure [56]. As depicted in the force curves for the case of hypotonic solution, the combination of a high curvature with shorter δ_{max} strongly indicates a higher membrane tension as a result of osmolyte release in the final equilibrium phase by the MS channels, which is consistent with values similar or slightly above to those obtained in PBS. Meanwhile, in the hypertonic solution, the curvature at the beginning of the curve is lower, and further deformation is required to attain the linear behavior. Based on our force spectroscopy results and reported fluorescence microscopy images showing plasmolysis bays in PA under hypertonic conditions, we speculate that this mechanical response is explained by the detachment between the outer and inner bacterial envelope caused by dehydration [53]. A loose outer envelope that has lost its tight connection with the inner envelope due to severe water depletion could provide a consistent explanation for the lower mechanical resistance or softening observed during the first steps of deformation. Consequently, additional deformation would be necessary before reaching the mechanical resistance transmitted from the inner envelope.

As stated above, the contribution to the steepness in the linear regime in the force curves reflects the P_t . In our study, PA seems to keep its mechanical response within the same values in hypotonic and hypertonic solutions, which shows its mechanical fitness after the equilibration process to some extent. The elastic deformation of PA in the frame of AFM nanoindentation can be modeled as the simultaneous compression of two spring elements in series with different stiffness, where the system cantilever+tip with stiffness k_{cant} compresses PA with stiffness k_{PA} . Thus, in a force curve, the slope in the linear region represents the effective action of both springs acting together, which can be expressed as k_{eff} . Applying Hooke's law for a system of two springs being compressed in series and solving for k_{PA} , we obtain $k_{PA} = (K_{cant} \cdot K_{eff}) \cdot (k_{cant} - k_{eff})^{-1}$ [57]. Table 1 shows the mean values obtained from this calculation once the effective stiffness values were extracted from the force curves. Measurements and a detailed analysis of membrane deformation by AFM nanoindentations on the Gramnegative bacterium Magnetospirillum gryphiswaldense were

first published by Arnoldi and Boulbitch [51,58]. Their study revealed a stiffness value for the membrane in the 0.04-0.07 N/m range. Their theoretical analysis allowed them to use this value to estimate turgor pressures within the 85-150 kPa range. To avoid the complex mathematical analysis of that first study on membrane elasticity, Yao and coworkers presented a tension-dominated model to derive the internal turgor pressure [52] of the Gram-positive and negative bacterium from the linear region of the slope obtained from force curves during deformation. For Gram-negative bacteria, they found that P_t increases an order of magnitude when bacteria are tested in distilled water (≈1.9 × 10⁵ Pa) compared with growth medium (0.1–0.12 \times 10⁵ Pa). Under the framework of the tension-dominated model developed by Yao and coworkers for rod-shaped bacteria such as PA, we can estimate P_t in our present study. Considering a mean height for PA of about 600 nm, an inner radius of 250 nm, which considers the presence of the double membrane in Gram-negative bacteria, and a mean stiffness of $k_B = 20$ mN/m, we obtained a turgor pressure of $P_t = 12.1$ kPa. This obtained value for P_t is in the same order as that reported by Yao for the growth medium. The higher value they reported for the stiffness in distilled water directly impacted the estimation of P_t , as all other parameters were practically unchanged. We do not observe this substantial difference in stiffness between PBS and Milli-Q water in our measurements. Aware that both studies were performed with a similar experimental setup using a fluid chamber in the AFM, we can argue that time scales between measurements and between solutions were similar. We cannot explain the steeper slope reported in distilled water in that study. Furthermore, according to Cetiner et al., a rapid osmolyte release takes place within the first 150 ms following dilution, which should not only decelerate swelling but also return membrane tension below a threshold value during cell equilibration [14]. On the other hand, our reported values for Young's modulus seem to capture the combined features in the force curve, meaning the curved region and the linear response. Altogether, our study suggests that the induced nanoindentations reflect different states of tension of the outer envelope at shallow deformations for the different tested solutions. Still, a similar stiffness arising from the remaining contents dominates for larger deformations. We associate this membrane tension and mechanical response with the equilibration time of the bacteria once MS channels have dissipated the osmotic gradient.

Care must be taken when quantifying the strength of adhesion forces via repetitive nanoindentations with the AFM because the risk of tip contamination is always present, and also because the sensitive nature of the intermolecular forces during the measuring conditions (pH, ionic strength, temperature, etc.) can easily affect their magnitude. This was reflected in the obtained

adhesion maps (see Figures S5 and S6 of Supporting Information File 1). In Milli-Q water, it is well known that a silicon nitride AFM tip is negatively charged. Consequently, upon close contact, a negatively charged AFM tip will experience on one side an attractive force towards a positively charged surface given by the cationic PLL and on the other side a repulsive force arising from the negatively charged bacterial membrane. In Milli-Q water, electrostatic double-layer forces strongly enhance the magnitude of adhesion forces at close range. This effect can be observed in the force curve shown as the control in Figure 4A taken in Milli-Q water on the substrate coated with PLL. The black line shows the presence of a jump-in, indicating the existence of a pulling force ≈15–20 nm before contact. The obvious contrast in the map of the adhesion channels together with the cross-sections shown in Supporting Information File 1, Figure S5 indicate the magnitude of F_A . On PA, a negative FA or repulsion of about 25 pN was obtained, while a positive adhesion FA of about 50 pN on the PLL coat was measured. These results are consistent with the information gained by ξ-potential measurements of PA when measured in Milli-Q water, where a negative surface charge for PA of about -25 mV was obtained (data not shown). Introducing salts into the solution resulted in the electrostatic screening of the double-layer forces, which resulted in the cancelation of long-range forces upon approach. The implication of this is shown in the adhesion maps of Supporting Information File 1, Figure S6, where a loss in the strength of adhesion forces started to occur in PBS while in 0.5 M NaCl it was even more pronounced.

Conclusion

We performed shallow (500 pN) nanoindentations to quantify the changes in rigidity of the outer cell wall of Pseudomonas aeruginosa in hypotonic, isotonic, and hypertonic conditions. Force-volume AFM demonstrated its capacity for testing the mechanical properties of multiple bacteria at once. This mechanical nanoscale mapping allowed us to successfully discriminate the minute variations in the surface topography of bacteria (h), their mechanical resistance to external deformation (k), an estimation of their elasticity modulus (Y), and their adhesive behavior towards the AFM tip to the substrate (F_A) . Generated maps from nanoindentations indicated that PA tolerates drastic changes in osmotic pressure from isotonic (0.35 Osm) to hypotonic (<0.05 Osm) or hypertonic (1 Osm) solutions without showing serious structural or morphological damage of the bacterial bodies. Deformation of PA by a pseudo-conical tip resulted in force curves with two main features, mainly a first nonlinear region we associated with the magnitude of membrane tension followed by a linear region related to the internal turgor pressure. Our results showed that for the time scale of our experiments, the stiffness given by the linear regime of the

curves did not significantly change the range of tested osmolarities. Application of previously developed models for AFM nanoindentation that relate the stiffness of a rod-shaped bacteria under deformation to its internal turgor pressure was used. The obtained values were consistent with those reported for Gramnegative bacteria in growth medium but inconsistent with those reported in hypotonic solutions. We argue that osmotic shock and potential cell lysis are prevented by the rapid reaction of MS channels to low osmolarities, which also decreases membrane tension. Decompression of exceeding internal pressures within PA is then reduced, and obtained values for k_{PA} and P_t represent the equilibration phase of PA which counteracts exceeding swelling. This demonstrates the remarkable dynamic capacity of the MS channels to rapidly mitigate hazardous internal pressures in highly diluted solutions. Therefore, we expect high membrane tension values to occur but in a very narrow period (within ≈100 ms after exposure), making it challenging to monitor AFM experiments. However, it was observed that the nonlinear region was affected by changes in osmolarity, providing a soft mechanical response for hypertonic solutions. We conclude that this initial weak mechanical response to deformation originates from a loose outer membrane due to dehydration, known as plasmolysis bays. Despite this, PA still provided a considerable mechanical response consistent with an internal turgor pressure for larger deformations. These investigations using nanomechanical mapping with FV AFM unravel fine mechanical parameters of very resilient bacteria with high adaptability to diverse environments. They also highlight the potential of the experimental approach to study the activity of rationally designed molecular channel inhibitors that can specifically bind channel proteins.

Supporting Information

Supporting Information File 1

Additional figures.

[https://www.beilstein-journals.org/bjnano/content/supplementary/2190-4286-16-86-S1.pdf]

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The graphical abstract was created in BioRender. García-Torres, L. (2025) https://BioRender.com/ide97j2. This content is not subject to CC BY 4.0.

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Data Availability Statement

Data generated and analyzed during this study is available from the corresponding author upon reasonable request.

References

- Hall-Stoodley, L.; Costerton, J. W.; Stoodley, P. Nat. Rev. Microbiol. 2004, 2, 95–108. doi:10.1038/nrmicro821
- Wiehlmann, L.; Wagner, G.; Cramer, N.; Siebert, B.; Gudowius, P.; Morales, G.; Köhler, T.; van Delden, C.; Weinel, C.; Slickers, P.; Tümmler, B. Proc. Natl. Acad. Sci. U. S. A. 2007, 104, 8101–8106. doi:10.1073/pnas.0609213104
- Mena, K. D.; Gerba, C. P. Risk Assessment of Pseudomonas aeruginosa in Water. Reviews of Environmental Contamination and Toxicology; Springer: New York, NY, USA, 2009; Vol. 201, pp 71–115. doi:10.1007/978-1-4419-0032-6_3
- Diggle, S. P.; Whiteley, M. Microbiology (London, U. K.) 2020, 166, 30–33. doi:10.1099/mic.0.000860
- Tuon, F. F.; Dantas, L. R.; Suss, P. H.; Tasca Ribeiro, V. S. *Pathogens* 2022, 11, 300. doi:10.3390/pathogens11030300
- Reynolds, D.; Kollef, M. Drugs 2021, 81, 2117–2131. doi:10.1007/s40265-021-01635-6
- Restrepo, M. I.; Babu, B. L.; Reyes, L. F.; Chalmers, J. D.; Soni, N. J.; Sibila, O.; Faverio, P.; Cilloniz, C.; Rodriguez-Cintron, W.; Aliberti, S. Eur. Respir. J. 2018, 52, 1701190. doi:10.1183/13993003.01190-2017
- Wood, S. J.; Kuzel, T. M.; Shafikhani, S. H. Cells 2023, 12, 199. doi:10.3390/cells12010199
- López, C. A.; Zgurskaya, H.; Gnanakaran, S. Biochim. Biophys. Acta, Biomembr. 2020, 1862, 183151. doi:10.1016/j.bbamem.2019.183151
- Matias, V. R. F.; Al-Amoudi, A.; Dubochet, J.; Beveridge, T. J. J. Bacteriol. 2003, 185, 6112–6118. doi:10.1128/jb.185.20.6112-6118.2003
- Sun, J.; Rutherford, S. T.; Silhavy, T. J.; Huang, K. C. Nat. Rev. Microbiol. 2022, 20, 236–248.
 doi:10.1038/s41579-021-00638-0
- Chevalier, S.; Bouffartigues, E.; Bodilis, J.; Maillot, O.; Lesouhaitier, O.; Feuilloley, M. G. J.; Orange, N.; Dufour, A.; Cornelis, P. FEMS Microbiol. Rev. 2017, 41, 698–722. doi:10.1093/femsre/fux020
- Ude, J.; Tripathi, V.; Buyck, J. M.; Söderholm, S.; Cunrath, O.; Fanous, J.; Claudi, B.; Egli, A.; Schleberger, C.; Hiller, S.; Bumann, D. *Proc. Natl. Acad. Sci. U. S. A.* 2021, 118, e2107644118. doi:10.1073/pnas.2107644118
- Çetiner, U.; Rowe, I.; Schams, A.; Mayhew, C.; Rubin, D.; Anishkin, A.;
 Sukharev, S. J. Gen. Physiol. 2017, 149, 595–609.
 doi:10.1085/jgp.201611699

- Zhang, Y.; Daday, C.; Gu, R.-X.; Cox, C. D.; Martinac, B.; de Groot, B. L.; Walz, T. *Nature* **2021**, *590*, 509–514. doi:10.1038/s41586-021-03196-w
- Blount, P.; Iscla, I. Microbiol. Mol. Biol. Rev. 2020, 84, e00055-19. doi:10.1128/mmbr.00055-19
- Poolman, B.; Spitzer, J. J.; Wood, J. M. Biochim. Biophys. Acta, Biomembr. 2004, 1666, 88–104. doi:10.1016/j.bbamem.2004.06.013
- Haswell, E. S.; Phillips, R.; Rees, D. C. Structure 2011, 19, 1356–1369. doi:10.1016/j.str.2011.09.005
- 19. Levina, N. *EMBO J.* **1999**, *18*, 1730–1737. doi:10.1093/emboi/18.7.1730
- Reuter, M.; Hayward, N. J.; Black, S. S.; Miller, S.; Dryden, D. T. F.; Booth, I. R. J. R. Soc., Interface 2014, 11, 20130850. doi:10.1098/rsif.2013.0850
- Lorusso, A. B.; Carrara, J. A.; Barroso, C. D. N.; Tuon, F. F.; Faoro, H. Int. J. Mol. Sci. 2022, 23, 15779. doi:10.3390/ijms232415779
- Compagne, N.; Vieira Da Cruz, A.; Müller, R. T.; Hartkoorn, R. C.; Flipo, M.; Pos, K. M. Antibiotics (Basel, Switz.) 2023, 12, 180. doi:10.3390/antibiotics12010180
- Amaral, L.; Martins, A.; Spengler, G.; Molnar, J. Front. Pharmacol. 2014, 4, 168. doi:10.3389/fphar.2013.00168
- 24. Li, X.-Z.; Plésiat, P. Antimicrobial Drug Efflux Pumps in Pseudomonas Aeruginosa. Efflux-Mediated Antimicrobial Resistance in Bacteria; Springer International Publishing: Cham, Switzerland, 2016; pp 359–400. doi:10.1007/978-3-319-39658-3 14
- Tambat, R.; Mahey, N.; Chandal, N.; Verma, D. K.; Jangra, M.; Thakur, K. G.; Nandanwar, H. ACS Infect. Dis. 2022, 8, 255–270. doi:10.1021/acsinfecdis.1c00281
- Ferrer-Espada, R.; Shahrour, H.; Pitts, B.; Stewart, P. S.;
 Sánchez-Gómez, S.; Martínez-de-Tejada, G. Sci. Rep. 2019, 9, 3452.
 doi:10.1038/s41598-019-39659-4
- Laborda, P.; Alcalde-Rico, M.; Chini, A.; Martínez, J. L.;
 Hernando-Amado, S. *Environ. Microbiol.* **2021**, *23*, 7396–7411.
 doi:10.1111/1462-2920.15511
- Alenazy, R. *Biology (Basel, Switz.)* 2022, 11, 1328. doi:10.3390/biology11091328
- Rampioni, G.; Pillai, C. R.; Longo, F.; Bondì, R.; Baldelli, V.;
 Messina, M.; Imperi, F.; Visca, P.; Leoni, L. Sci. Rep. 2017, 7, 11392.
 doi:10.1038/s41598-017-11892-9
- Sharma, A.; Gupta, V. K.; Pathania, R. Indian J. Med. Res. 2019, 149, 129–145. doi:10.4103/jjmr.ijmr_2079_17
- Fujiwara, M.; Yamasaki, S.; Morita, Y.; Nishino, K. J. Infect. Chemother. 2022, 28, 595–601. doi:10.1016/j.jiac.2022.01.003
- Shriram, V.; Khare, T.; Bhagwat, R.; Shukla, R.; Kumar, V.
 Front. Microbiol. 2018, 9, 2990. doi:10.3389/fmicb.2018.02990
- Dey, N.; Kamatchi, C.; Vickram, A. S.; Anbarasu, K.; Thanigaivel, S.; Palanivelu, J.; Pugazhendhi, A.; Ponnusamy, V. K. Environ. Res. 2022, 204, 111968. doi:10.1016/j.envres.2021.111968
- 34. Madhi, M.; Hasani, A.; Mojarrad, J. S.; Rezaee, M. A.; Zarrini, G.; Davaran, S. *Gene Rep.* **2020**, *21*, 100915. doi:10.1016/j.genrep.2020.100915
- Musiol, R. Expert Opin. Ther. Targets 2023, 27, 953–963. doi:10.1080/14728222.2023.2263910
- 36. Christena, L. R.; Mangalagowri, V.; Pradheeba, P.; Ahmed, K. B. A.; Shalini, B. I. S.; Vidyalakshmi, M.; Anbazhagan, V.; Sai subramanian, N. RSC Adv. 2015, 5, 12899–12909. doi:10.1039/c4ra15382k

- Nefedova, E.; Shkil, N.; Luna Vazquez-Gomez, R.; Garibo, D.;
 Pestryakov, A.; Bogdanchikova, N. Pharmaceutics 2022, 14, 763.
 doi:10.3390/pharmaceutics14040763
- 38. Dufrêne, Y. F. mBio 2014, 5, e01363-14. doi:10.1128/mbio.01363-14
- Dufrêne, Y. F. Nat. Rev. Microbiol. 2008, 6, 674–680. doi:10.1038/nrmicro1948
- Benn, G.; Pyne, A. L. B.; Ryadnov, M. G.; Hoogenboom, B. W. Analyst 2019, 144, 6944–6952. doi:10.1039/c9an01185d
- Jarosławski, S.; Duquesne, K.; Sturgis, J. N.; Scheuring, S. Mol. Microbiol. 2009, 74, 1211–1222. doi:10.1111/j.1365-2958.2009.06926.x
- Dorobantu, L. S.; Gray, M. R. Scanning 2010, 32, 74–96.
 doi:10.1002/sca.20177
- 43. Olubowale, O. H.; Biswas, S.; Azom, G.; Prather, B. L.; Owoso, S. D.; Rinee, K. C.; Marroquin, K.; Gates, K. A.; Chambers, M. B.; Xu, A.; Garno, J. C. ACS Omega 2021, 6, 25860–25875. doi:10.1021/acsomega.1c03829
- 44. Longo, G.; Rio, L. M.; Roduit, C.; Trampuz, A.; Bizzini, A.; Dietler, G.; Kasas, S. J. Mol. Recognit. 2012, 25, 278–284. doi:10.1002/jmr.2171
- Polyakov, P.; Soussen, C.; Duan, J.; Duval, J. F. L.; Brie, D.; Francius, G. *PLoS One* **2011**, *6*, e18887. doi:10.1371/journal.pone.0018887
- 46. Xu, X.; Feng, H.; Zhao, Y.; Shi, Y.; Feng, W.; Loh, X. J.; Vancso, G. J.; Guo, S. Cell Rep. Phys. Sci. 2024, 5, 101902. doi:10.1016/j.xcrp.2024.101902
- Offroy, M.; Razafitianamaharavo, A.; Beaussart, A.; Pagnout, C.;
 Duval, J. F. L. RSC Adv. 2020, 10, 19258–19275.
 doi:10.1039/d0ra00669f
- 48. Han, R.; Vollmer, W.; Perry, J. D.; Stoodley, P.; Chen, J. *Nanoscale* **2022**, *14*, 12060–12068. doi:10.1039/d2nr02577a
- Zhang, H.; Wang, H.; Wilksch, J. J.; Strugnell, R. A.; Gee, M. L.; Feng, X.-Q. Soft Matter 2021, 17, 2042–2049. doi:10.1039/d0sm02075c
- Valentová, L.; Füzik, T.; Nováček, J.; Hlavenková, Z.; Pospíšil, J.; Plevka, P. *EMBO J.* 2024, 43, 4384–4405. doi:10.1038/s44318-024-00195-1
- 51. Arnoldi, M.; Fritz, M.; Bäuerlein, E.; Radmacher, M.; Sackmann, E.; Boulbitch, A. *Phys. Rev. E* **2000**, *62*, 1034–1044. doi:10.1103/physreve.62.1034
- Yao, X.; Walter, J.; Burke, S.; Stewart, S.; Jericho, M. H.; Pink, D.; Hunter, R.; Beveridge, T. J. Colloids Surf., B 2002, 23, 213–230. doi:10.1016/s0927-7765(01)00249-1
- Lewenza, S.; Mhlanga, M. M.; Pugsley, A. P. J. Bacteriol. 2008, 190, 6119–6125. doi:10.1128/jb.00603-08
- Elabed, H.; González-Tortuero, E.; Ibacache-Quiroga, C.; Bakhrouf, A.; Johnston, P.; Gaddour, K.; Blázquez, J.; Rodríguez-Rojas, A. BMC Microbiol. 2019, 19, 142. doi:10.1186/s12866-019-1499-2
- Aspedon, A.; Palmer, K.; Whiteley, M. J. Bacteriol. 2006, 188, 2721–2725. doi:10.1128/jb.188.7.2721-2725.2006
- Steinkühler, J.; Sezgin, E.; Urbančič, I.; Eggeling, C.; Dimova, R.
 Commun. Biol. 2019, 2, 337. doi:10.1038/s42003-019-0583-3
- Michel, J. P.; Ivanovska, I. L.; Gibbons, M. M.; Klug, W. S.; Knobler, C. M.; Wuite, G. J. L.; Schmidt, C. F. *Proc. Natl. Acad. Sci. U. S. A.* 2006, *103*, 6184–6189. doi:10.1073/pnas.0601744103
- Boulbitch, A. J. Electron Microsc. 2000, 49, 459–462. doi:10.1093/oxfordjournals.jmicro.a023829

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