



Supporting Information

for

Glucose-derived carbon materials with tailored properties as electrocatalysts for the oxygen reduction reaction

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Beilstein J. Nanotechnol. **2019**, *10*, 1089–1102. doi:10.3762/bjnano.10.109

Characterization of the carbon materials, electrochemical assessments and relations resulting from this work

Effect of microporosity

Table S1: Textural parameters of samples activated during different times.

Sample	S_{BET} ($\text{m}^2 \text{g}^{-1}$)	V_p ($\text{cm}^3 \text{g}^{-1}$)	V_{DR} ($\text{cm}^3 \text{g}^{-1}$)
AG _{1h}	936	0.38	0.35
AG _{3h}	1121	0.44	0.43
AG _{4h}	1314	0.54	0.52
AG _{6h}	1984	0.85	0.82

Table S2: Elemental analysis of samples activated during different times.

Sample	Carbon (wt %)	Oxygen (wt %)	Hydrogen (wt %)
AG _{1h}	96.3	3.3	0.4
AG _{3h}	96.6	3.0	0.4
AG _{4h}	96.8	2.8	0.4
AG _{6h}	97.3	2.4	0.3

Table S3: CO and CO₂ amounts obtained from TPD measurements of AG_{1h} and AG_{6h}

Sample	CO ($\mu\text{mol g}^{-1}$)	CO ₂ ($\mu\text{mol g}^{-1}$)	CO/CO ₂	Oxygen (%)
AG _{1h}	1111	172	6.5	2.3
AG _{6h}	971	136	7.2	2.0

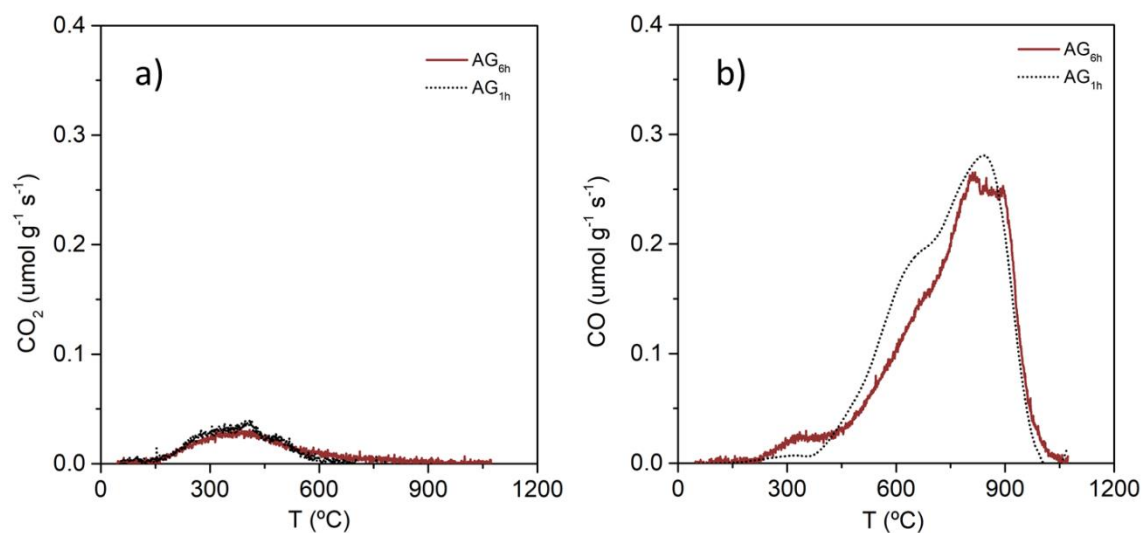


Figure S1: TPD profiles of CO_2 (a) and CO (b) for $\text{AG}_{6\text{h}}$ and $\text{AG}_{1\text{h}}$.

Table S4: Onset potential of samples activated during different times.

Sample	Onset Potential (V)
$\text{AG}_{1\text{h}}$	0.75
$\text{AG}_{3\text{h}}$	0.75
$\text{AG}_{4\text{h}}$	0.76
$\text{AG}_{6\text{h}}$	0.78

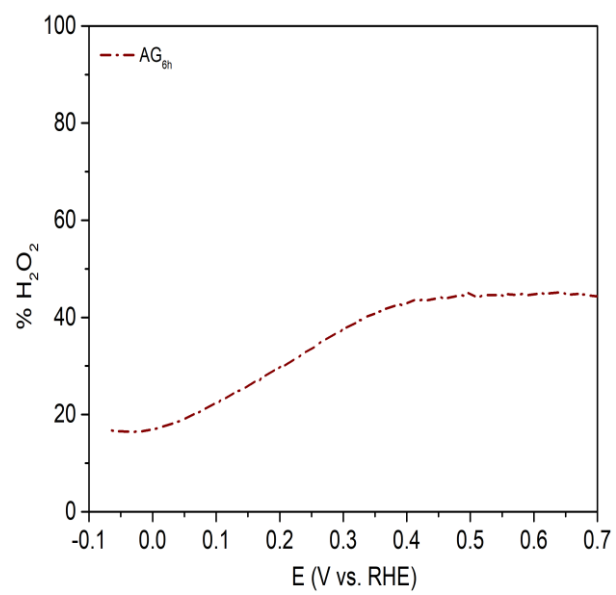


Figure S2: Hydrogen peroxide production for sample $\text{AG}_{6\text{h}}$.

Effect of the surface chemistry

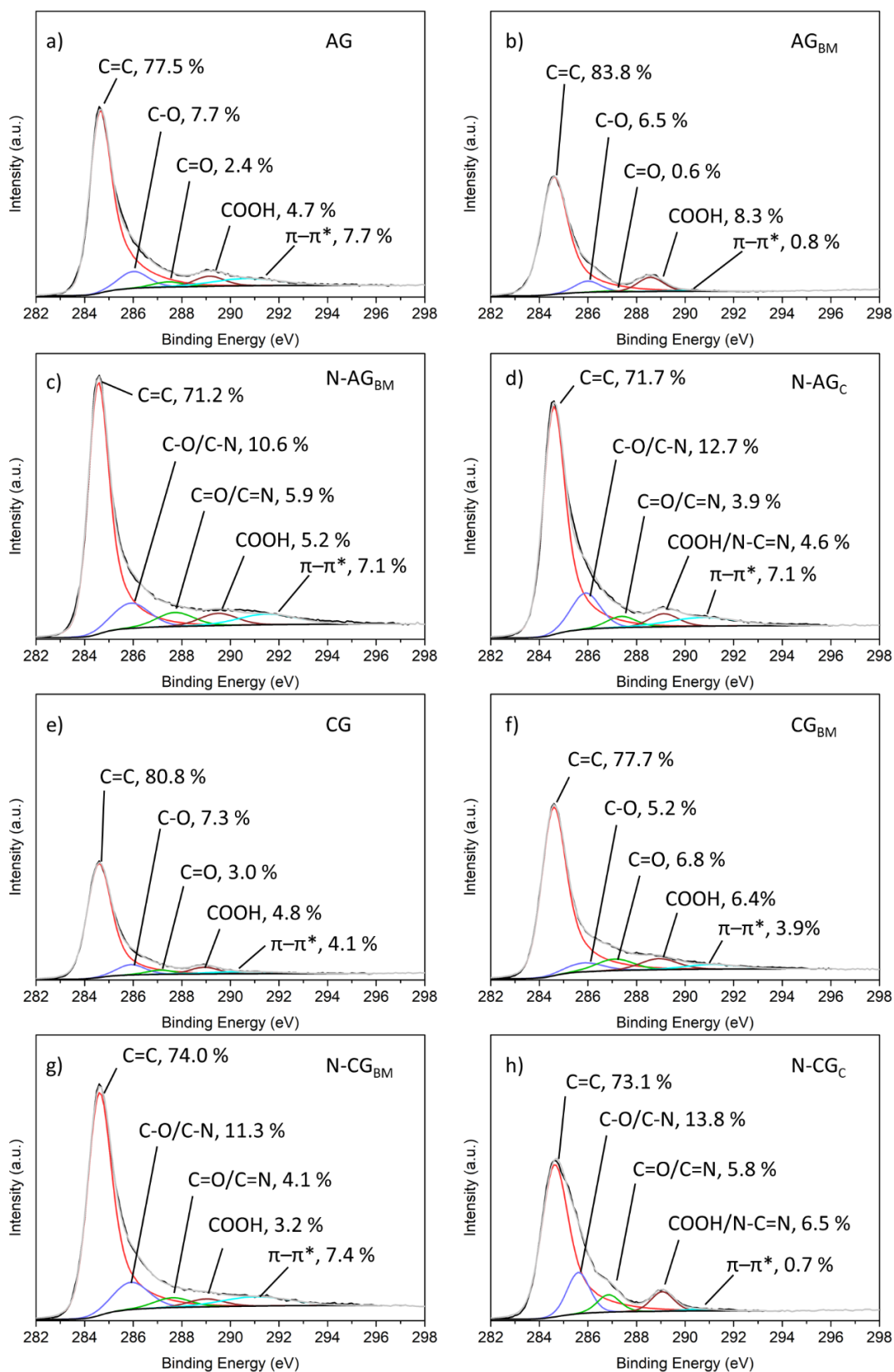


Figure S3: C 1s spectra deconvolution of AG (a), AG_{BM} (b), N-AG_{BM} (c), N-AG_C (d), CG (e), CG_{BM} (f), N-CG_{BM} (g), N-CG_C (h).

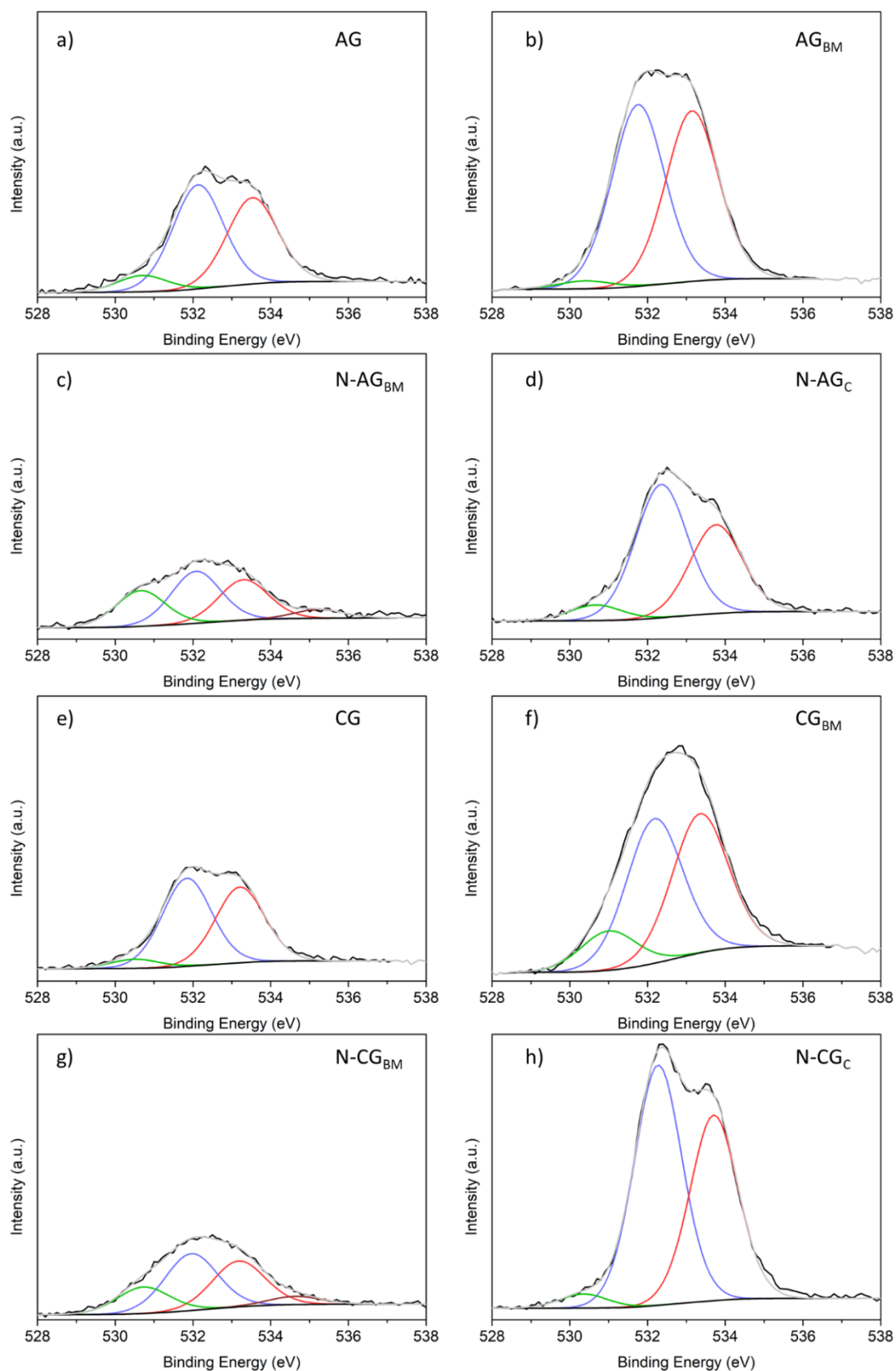


Figure S4: O 1s spectra deconvolution of AG (a), AG_{BM} (b), N-AG_{BM} (c), N-AG_C (d), CG (e), CG_{BM} (f), N-CG_{BM} (g), N-CG_C (h).

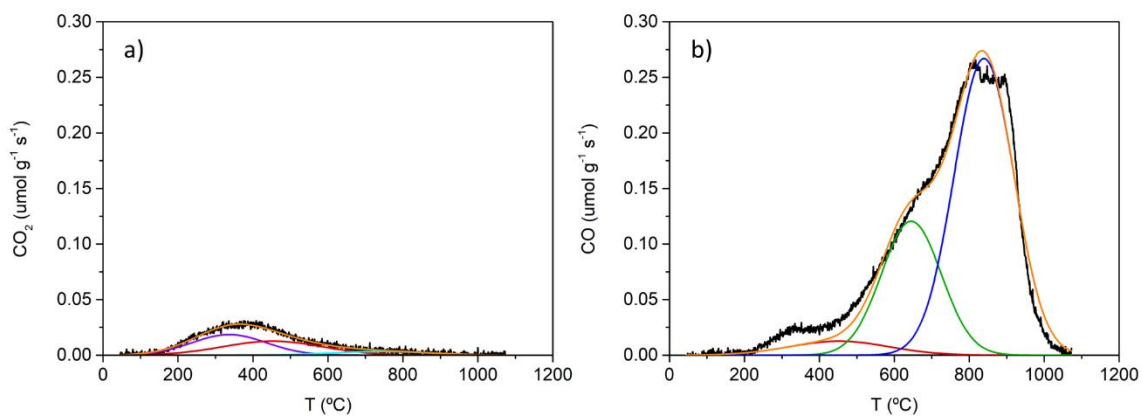


Figure S5: TPD profile deconvolution of AG for CO₂ (a) and CO (b).

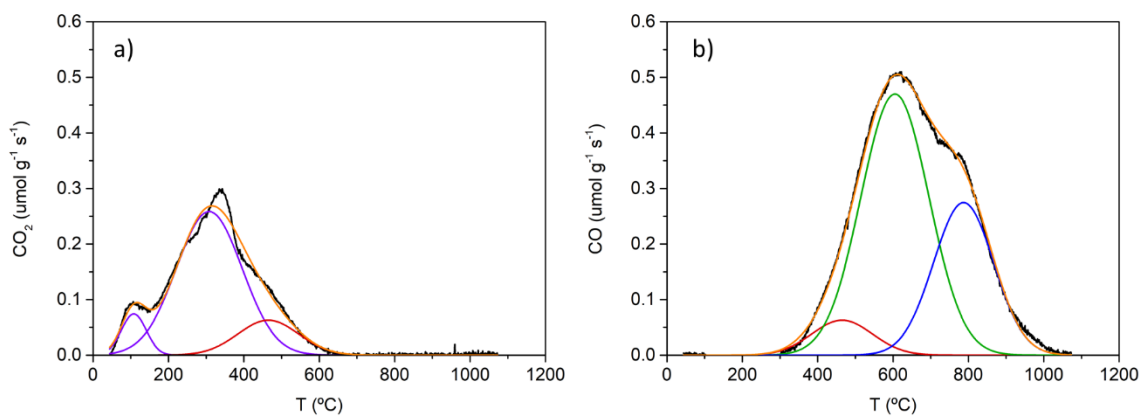


Figure S6: TPD profile deconvolution of AG_{BM} for CO₂ (a) and CO (b).

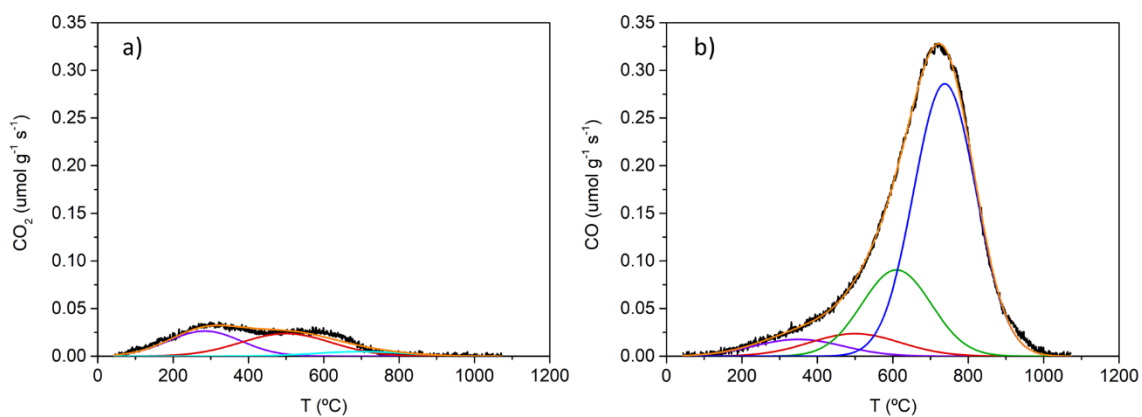


Figure S7: TPD profile deconvolution of CG for CO₂ (a) and CO (b).

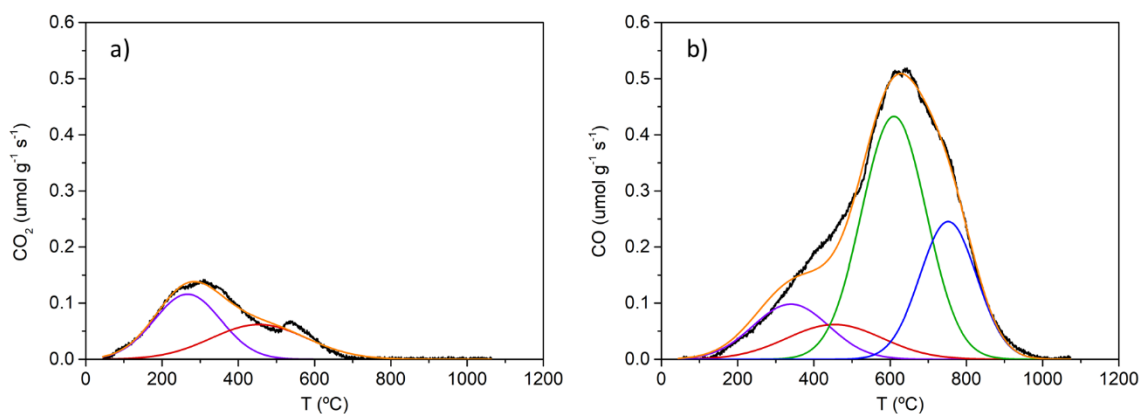


Figure S8: TPD profile deconvolution of CG_{BM} for CO₂ (a) and CO (b).

The activated sample (AG) presents three broad peaks in the CO₂ profile (Figure S5a), at 340 $^{\circ}\text{C}$, 450 $^{\circ}\text{C}$ and 740 $^{\circ}\text{C}$, which could be attributed to carboxylic acids, anhydride and lactone groups, respectively. AG_{BM} also presents the carboxylic acids and anhydride peaks (Figure S6a). However, its CO₂ profile closes at lower temperatures than the CO₂ profile of AG, thus almost not presenting the peak attributed of lactones. The CO₂ profile of sample CG (Figure S7a) also presents the same three peaks as the activated sample. Similar to activated samples, the modification of the structure by the ball milling method (Figure S8a) results in an almost depletion of the lactone group and in a significant increase of the carboxylic acids, once again corroborating the XPS results. Since lactones are cyclic esters of carboxylic acids, the defects created by ball milling may be opening the cycle creating the opportunity for the chain to react with air moisture and thus originating carboxylic functional groups. As a result of the decrease of lactones detected from CO₂ release, an increase of carboxylic acids occurs.

The CO profile of sample AG (Figure S5b) presents three main peaks corresponding to anhydrides, phenols and carbonyl/quinone groups at temperature ranges of 450-470 $^{\circ}\text{C}$, 600-650 $^{\circ}\text{C}$ and 790-840 $^{\circ}\text{C}$, respectively. These three groups are also present in sample AG_{BM} (Figure S6b) but with a much more significant contribution of phenols, corroborating the hypothesis of ball milling generating oxygen functional groups with weaker bonds due to reactions with air moisture. This

same effect is observed for carbonized samples (Figure S7b and Figure S8b for CG and CG_{BM}, respectively). Nevertheless, it should be noted that the temperature at which oxygen groups are released as CO₂ and CO is higher for sample AG than for sample CG, confirming that the activated sample is more chemically stable.

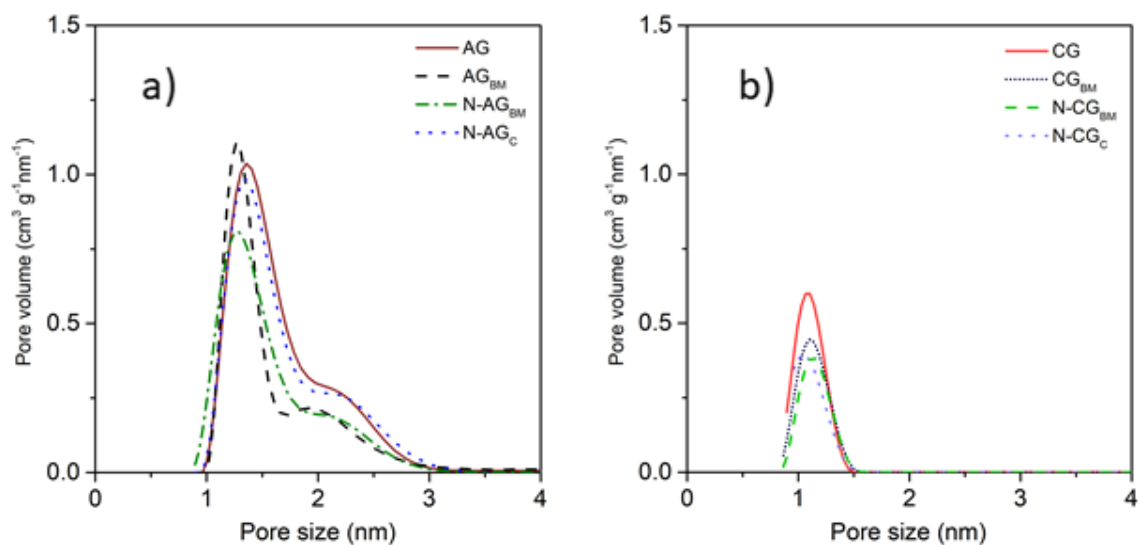


Figure S9: Pore size distributions for activated samples (a) and carbonized samples (b).

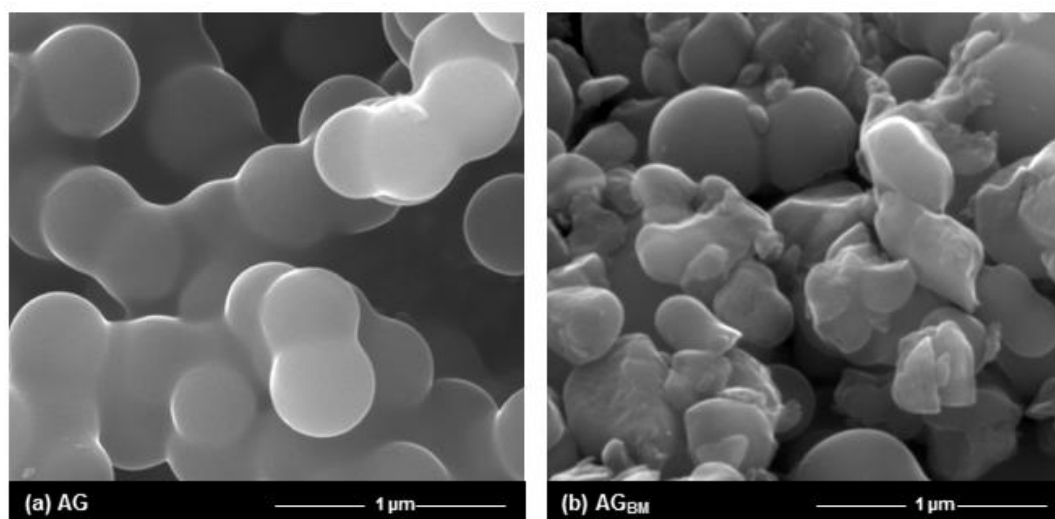


Figure S10: SEM images of activated glucose before (a) and after (b) ball-milling.

Table S5: Textural characterization of activated and carbonized samples.

Sample	S_{BET} ($\text{m}^2 \text{g}^{-1}$)	V_p ($\text{cm}^3 \text{g}^{-1}$)	V_{DR} ($\text{cm}^3 \text{g}^{-1}$)
AG	1984	0.85	0.82
AG _{BM}	1486	0.71	0.54
N-AG _C	1870	0.79	0.68
N-AG _{BM}	1528	0.75	0.55
CG	547	0.23	0.20
CG _{BM}	451	0.32	0.17
N-CG _C	445	0.20	0.17
N-CG _{BM}	375	0.20	0.15

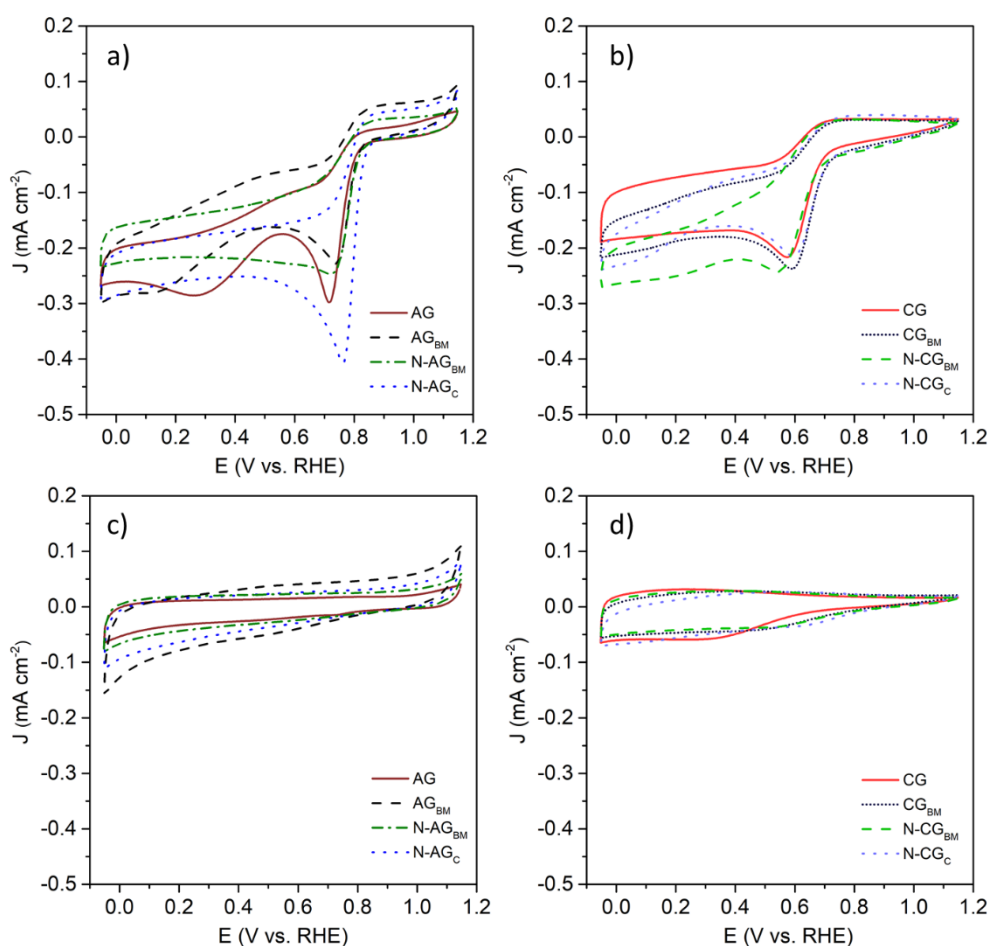


Figure S11: O₂-saturated cyclic voltammograms for activated samples (a) and carbonized samples (b) and N₂-saturated cyclic voltammograms for activated samples (c) and carbonized samples (d).

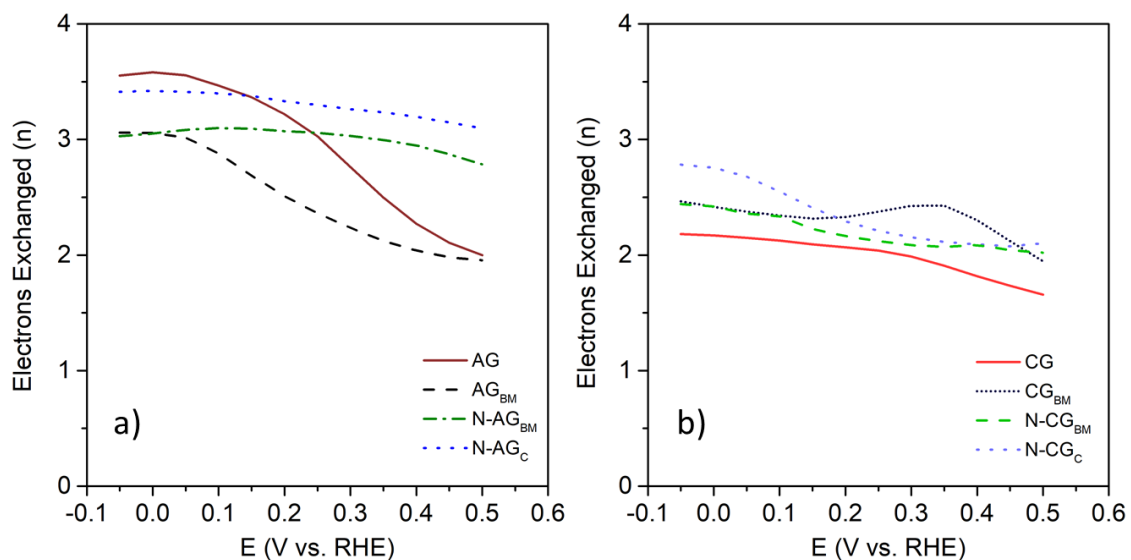


Figure S12: Number of electrons exchanged for activated samples (a) and carbonized samples (b) at different potentials.

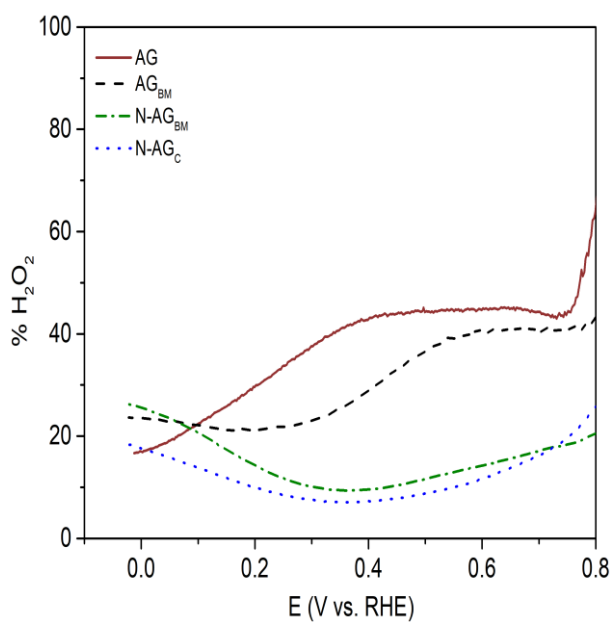


Figure S13: Hydrogen peroxide production for activated samples.