

Supporting Information

for

Antibiofilm and cytotoxic metabolites from the entomopathogenic fungus *Samsoniella aurantia*

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HRESIMS profiles and NMR spectroscopic data of compounds 1–6

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Biofilm inhibition assay

Biofilm inhibition assays were conducted according to the methodology previously described (Soliga et al., 2021), targeting the inhibition of biofilm formation in *Staphylococcus aureus* (DSM 1104). *S. aureus* were cultured in CASO medium at 37 °C for 18 h, respectively, in a shaker at 120 rpm after being thawed from a -20 °C stock solution. The culture's optical density at 600 nm (OD600) was measured and adjusted to a 0.001 McFarland standard in CASO medium with 4% glucose for *S. aureus*, respectively. Microtiter plates were loaded with 150 µL of the bacterial suspension containing serially diluted test compounds (125–0.9 µg/mL) and incubated for 24 h at 37 °C. The evaluation of biofilm inhibition was performed by staining the biofilm with 0.1% crystal violet (CV). After staining, the plates were washed, and the biofilm dissolved in 95% ethanol. The absorbance of the dissolved biofilm was measured at 530 nm using a plate reader. The negative control was methanol (2.5%) while microporenic acid A at a concentration range of 7.8 to 125 µg/mL was used as the positive control for *S. aureus* (Chepkirui et al., 2018). All experiments were performed twice, and the standard deviation (SD) was maintained at or below 15% for *S. aureus*.

Differences between samples and the control group were determined by a two-tailed Student's *t*-test. Statistical significance was defined as p < 0.05. Analysis was carried out using GraphPad Prism 9[®] (GraphPad Software, San Diego, CA, USA).

References

Chepkirui, C.; Yuyama, KT.; Wanga, LA.; Decock, C.; Matasyoh, JC.; Abraham, WR.; Stadler, M. Microporenic acids A-G, biofilm inhibitors, and antimicrobial agents from the Basidiomycete *Microporus* Species. *J. Nat. Prod.* **2018**, 81, 778-784.

Soliga, KJ.; Bär, SI.; Oberhuber, N.; Zeng, H.; Schrey, H.; Schobert, R. Synthesis and bioactivity of ancorinoside B, A marine diglycosyl tetramic acid. *Mar. Drugs.* **2021**, 19, 583.



Figure S1. A. Effect of farinosone D (1) on growth of *S. aureus* at varying concentrations in CASO medium. B. Effect of farinosone A (2) on growth of *S. aureus* at varying concentrations in CASO medium. C. Effect of farinosones D (1) and A (2) relative percent adhesion of *S. aureus* to fibrinogen. Microporenic acid A (MAA) was used as positive control. Methanol was used as a solvent control and taken as 100%. Error bars indicate standard deviation of duplicates in two biological repeats; p values: * p < 0.05, ** p < 0.01, *** p < 0.001.



Figure S2. A. Effect of farinosone D (1) on growth of *S. aureus* at varying concentrations in Müller Hinton Broth (MHB) medium. B. Effect of farinosone A (2) on growth of *S. aureus* at varying concentrations in Müller Hinton Broth (MHB) medium.







Generic Display Report

Figure S4. HRESIMS spectrum of 1.



Figure S5. ¹H NMR spectrum of **1** in DMSO- d_6 at 500 MHz.



Figure S6. ¹³C NMR spectrum of **1** in DMSO- d_6 at 125 MHz.



Figure S7. ¹H-¹H COSY spectrum of **1** in DMSO- d_6 at 500 MHz.



Figure S8. HMBC spectrum of 1 in DMSO- d_6 at 500 MHz.



Figure S9. HSQC spectrum of $\mathbf{1}$ in DMSO- d_6 at 500 MHz.



Figure S10. ROESY spectrum of $\mathbf{1}$ in DMSO- d_6 at 500 MHz.



Figure S11. Comparison of experimental (black) and simulated Boltzmann-averaged ECD spectra for compound **1**.

Table S1. Similarity factor (Bruhn et al., 2013) for each calculated stereoisomer of compound 1.

Stereoisomer of 1	Similarity factor
(2'S,3'S,12S)	0.70
$(2^{S}, 3^{S}, 12R)$	0.69
$(2^{R}, 3^{S}, 12R)$	0.61
$(2^{R}, 3^{S}, 12S)$	0.60
$(2^{S}, 3^{R}, 12R)$	0.25
(2'S, 3'R, 12S)	0.24
$(2^{R}, 3^{R}, 12S)$	0.16
$(2^{R}, 3^{R}, 12R)$	0.16

Bruhn, T.; Schaumlöffel, A.; Hemberger, Y.; Bringmann, G. SpecDis: quantifying the comparison of calculated and experimental electronic circular dichroism spectra. *Chirality* **2013**, *25(4)*, 243–249. doi:10.1002/chir.22138







Figure S13. ¹H NMR spectrum of **2** in acetone- d_6 at 500 MHz.



Figure S14. DEPTQ spectrum of 2 in acetone- d_6 at 125 MHz.



Figure S15. ¹H-¹H COSY spectrum of **2** in acetone- d_6 at 500 MHz.



Figure S16. HMBC spectrum of $\mathbf{2}$ in acetone- d_6 at 500 MHz.



Figure S17. HSQC spectrum of 2 in acetone- d_6 at 500 MHz.







Figure S19. ¹H NMR spectrum of **3** in acetone- d_6 at 500 MHz.



Figure S20. ¹H-¹H COSY spectrum of **3** in acetone- d_6 at 500 MHz.



Figure S21. HMBC spectrum of **3** in acetone- d_6 at 500 MHz.



Figure S22. HSQC spectrum of **3** in acetone- d_6 at 500 MHz.



Figure S23. HRESIMS spectrum of 4.



Figure S24. ¹H NMR spectrum of **4** in DMSO- d_6 at 500 MHz.



Figure S25. ¹H-¹H COSY spectrum of **4** in DMSO-*d*₆ at 500 MHz.







Figure S27. HSQC spectrum of **4** in DMSO-*d*₆ at 500 MHz.



Figure S28. HRESIMS spectrum of 5.



Figure S29. ¹H NMR spectrum of **5** in DMSO- d_6 at 500 MHz.



Figure S30. ¹H-¹H COSY spectrum of **5** in DMSO- d_6 at 500 MHz.



Figure S31. HMBC spectrum of **5** in DMSO-*d*₆ at 500 MHz.



Figure S32. HSQC spectrum of $\mathbf{5}$ in DMSO- d_6 at 500 MHz.







Figure S34. ¹H NMR spectrum of **6** in DMSO- d_6 at 500 MHz.