

# **Supporting Information**

for

# Sequential two-step, one-pot microwave-assisted Urech synthesis of 5-monosubstituted hydantoins from L-amino acids in water

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# NMR and MS spectra of synthesized compounds

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## **Supporting Information**

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## 1. General information

All chemicals were acquired from commercial sources (Merck or Thermo Fischer) at reagent grade and were used without further purification. The syntheses were carried out in an Anton-Parr Monowave 400 microwave reactor with the reaction temperature controlled using the on-broad infra-red sensor. The NMR spectra were acquired in a Bruker DMX 400 NMR instrument at a resolution of 400 MHz and 100 MHz, for <sup>1</sup>H and <sup>13</sup>C NMR respectively, using the residual solvent peak of DMSO-*d*<sub>6</sub> at 2.50 ppm (<sup>1</sup>H NMR) and 39.5 ppm (<sup>13</sup>C NMR) as reference. All chemical shifts ( $\delta$ ) were recorded in ppm unit and coupling frequency (*J*) in Hertz (Hz). Shorthand notations were used for the NMR characterization: s (singlet); d (doublet); q (quartet); dd (doublet of doublets); td (triplet of doublets); m (multiplet). The high resolution *m/z* values were measured in Dalton units (Da) using an Agilent 1290 Infinity LC system coupled to an Agilent 6520 Accurate-Mass Q-TOF mass spectrometer with the ESI source in positive mode. Shorthand notation: calcd (calculated). HPLC analyses were performed using a mobile phase of acetonitrile/ultrapure water (9:1) in an Agilent 1260 with a Zorbax C<sub>18</sub> column (5 µm) and detected by diode array at 210 nm and 254 nm. All hydantoin compounds were reported to be >95% pure.

### 2. Synthesis and characterization of products

**General procedure** for the synthesis of (*S*)-5-benzylimidazolidine-2,4-dione (**H2a**):

A 30 mL microwave reactor vial was charged with L-phenylalanine (5 mmol), distilled water (7 mL), and potassium cyanate (25 mmol) and irradiated in an Anton–Paar Monowave 400 microwave reactor at 80 °C for 1 hour. Upon completion of the *N*-carbamylation reaction, as confirmed by TLC analysis, the reaction mixture was treated with concentrated hydrochloric acid (7 mL) and microwave irradiated at 80 °C for 15 min. The precipitates in the reaction mixture were filtered, washed with 1 M HCl solution (2 × 7 mL), distilled water (2 × 10 mL), and dried to afford **H2a** as a white solid. Yield 89%.  $[\alpha]_D^{20}$ -100.9 (*c* 11, CH<sub>3</sub>CN). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  10.44 (s, 1H), 7.93 (s, 1H), 7.17 – 7.30 (m, 5H), 4.33 (td, *J* = 5, 1 Hz, 1H), 2.92 (m, 2H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>)  $\delta$  175.2, 157.2, 135.6, 129.8, 128.1, 126.7, 58.4, 36.4. LC-MS (ESI) m/z: 191 [M+H]<sup>+</sup>, 163, 120. HRMS (ESI) m/z: [M+H]<sup>+</sup> Calcd for C<sub>10</sub>H<sub>11</sub>N<sub>2</sub>O<sub>2</sub> 191.0815; Found: 191.0815. The procedure was repeated by substituting L-phenylalanine with the corresponding amino acid to obtain **H2b–c** and **H2e–f**.

**General procedure** for the synthesis of (*S*)-5-((2,5-dioxoimidazolidin-4-yl)methyl)-1*H*-imidazol-3-ium chloride (**H2d**):

A 30 mL microwave reactor vial was charged with L-histidine (5 mmol), distilled water (7 mL), and potassium cyanate (25 mmol) and irradiated in an Anton-Paar Monowave 400 microwave reactor at 80 °C for 1 hour. Upon completion of the *N*-carbamylation reaction, as confirmed by TLC analysis, the

reaction mixture was treated with concentrated hydrochloric acid (7 mL) and microwave irradiated at 80 °C for 15 min. The reaction mixture was neutralized to pH 7 using saturated sodium bicarbonate solution and extracted with ethyl acetate multiple times. The organic extracts were combined, dried with anhydrous sodium sulfate, and concentrated under reduced pressure to afford **H2d** as a white solid. Yield 70%. <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  10.59 (s, 1H), 7.88 (s, 1H), 7.74 (s, 1H), 6.88 (s, 1H), 5.52 (s, 2H), 4.25 (t, *J* = 6 Hz, 1H), 2.95 (dd, *J* = 15, 4 Hz, 1H), 2.81 (dd, *J* = 15, 6 Hz, 1H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>)  $\delta$  175.5, 157.4, 134.6, 131.6, 117.0, 57.6, 28.8. LC-MS (ESI) m/z: 181 [M+H]<sup>+</sup>, 136. The procedure was repeated by substituting L-histidine with the corresponding amino acid to obtain **H2g**–**j**.

## 2.1 Characterization data

## Carbamoyl-L-phenylalanine (H1a):

White solid. Yield 89%.  $[\alpha]_D^{20}$ -33.4 (*c* 10, CH<sub>3</sub>CN). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  7.28 (t, *J* = 7 Hz, 2H), 7.20 (m, 3H), 6.18 (d, *J* = 8 Hz, 1H), 5.62 (s, 2H), 4.33 (dd, *J* = 13, 8 Hz, 1H), 2.99 (dd, *J* = 13, 5 Hz, 1H), 2.84 (dd, *J* = 13, 8 Hz, 1H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>)  $\delta$  174.1, 158.4, 137.6, 129.4, 128.3, 126.6, 53.9, 37.7. LC-MS (ESI) m/z: 209 [M+H]<sup>+</sup>, 166, 120. Data in accordance with literature [1].<sup>20</sup>

(S)-5-(4-Hydroxybenzyl)imidazolidine-2,4-dione (H2b):

White solid. Yield 84%.  $[\alpha]_D^{20}$ -90.5 (*c* 10, acetone). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  10.39 (1H, s), 9.27 (s, 1H), 7.87 (s, 1H), 6.96 (d, *J* = 8 Hz, 2H), 6.64 (d, *J* = 8 Hz, 2H), 4.23 (q, *J* = 5 Hz, 1H), 2.80 (d, *J* = 5 Hz, 2H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>)  $\delta$  175.4, 157.3, 156.1, 130.8, 125.5, 114.9, 58.7, 35.6. LC-MS (ESI) m/z: 207 [M+H]<sup>+</sup>, 179, 136, 107. HRMS (ESI) m/z: [M+H]<sup>+</sup> Calcd for C<sub>10</sub>H<sub>11</sub>N<sub>2</sub>O<sub>3</sub> 207.0764; Found: 207.0759.

(S)-5-((1H-Indol-2-yl)methyl)imidazolidine-2,4-dione (H2c):

White solid. Yield 71%.  $[\alpha]_{D}^{20}$ -97.1 (*c* 10, CH<sub>3</sub>CN). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  10.90 (s, 1H), 10.37 (s, 1H), 7.91 (s, 1H), 7.55 (d, *J* = 8 Hz, 1H), 7.32 (d, *J* = 8 Hz, 1H), 7.12 (d, *J* = 2 Hz, 1H), 7.05 (td, *J* = 7, 1 Hz, 1H), 6.97 (td, *J* = 7, 1 Hz, 1H), 4.30 (dd, *J* = 4, 1 Hz, 1H), 3.06 (d, *J* = 4 Hz, 2H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>)  $\delta$  175.8, 157.5, 135.9, 127.5, 124.2, 120.9, 118.7, 118.4, 111.3, 108.0, 58.4, 26.5. LC-MS (ESI) m/z: 230 [M+H]<sup>+</sup>, 130. HRMS (ESI) m/z: [M+H]<sup>+</sup> Calcd for C<sub>12</sub>H<sub>12</sub>N<sub>3</sub>O<sub>2</sub> 230.0924; Found: 230.0926.

(S)-5-Phenylimidazolidine-2,4-dione (H2e):

White solid. Yield 48%.  $[\alpha]_{D}^{20}$ -136.0 (*c* 10, CH<sub>3</sub>CN). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  10.81 (s, 1H), 8.41 (s, 1H), 7.32 – 7.43 (m, 5H), 5.16 (s, 1H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>)  $\delta$  174.3, 157.6, 136.1, 128.7, 128.3, 126.8, 61.3. LC-MS (ESI) m/z: 177 [M+H]<sup>+</sup>, 149, 106. Data in accordance with literature [2].<sup>21</sup>

(S)-5-Isobutylimidazolidine-2,4-dione (H2f):

White solid. Yield 34%.  $[\alpha]_{D}^{20}$ -78.0 (*c* 10, CH<sub>3</sub>CN). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  10.59 (s, 1H), 8.01 (s, 1H), 3.99 (m, 1H), 1.75 (m, 1H), 1.34 – 1.51 (m, 2H), 0.87 (t, *J* = 6 Hz, 6H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>)  $\delta$  176.6, 157.5, 56.2, 40.8, 24.2, 23.2, 21.5. LC-MS (ESI) m/z: 157 [M+H]<sup>+</sup>, 129. Data in accordance with literature [2].<sup>21</sup>

(S)-5-((R)-1-Hydroxyethyl)imidazolidine-2,4-dione (H2g):

White solid. Yield 78%.  $[\alpha]_{D}^{20}$ -103.4 (*c* 10, CH<sub>3</sub>CN). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  7.89 (s, 1H), 5.48 (s, 1H), 4.93 (d, *J* = 6 Hz, 1H), 3.85 – 3.92 (m, 1H), 3.82 (dd, *J* = 3, 1 Hz, 1H), 1.12 (d, *J* = 6 Hz, 3H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>)  $\delta$  174.8, 158.2, 65.2, 63.9, 20.3. LC-MS (ESI) m/z: 145 [M+H]<sup>+</sup>, 101. Data in accordance with literature [3].<sup>22</sup>

(S)-5-Isopropylimidazolidine-2,4-dione (H2h):

White solid. Yield 86%.  $[\alpha]_D^{20}$ -85.9 (*c* 10, CH<sub>3</sub>CN). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  10.56 (s, 1H), 7.89 (s, 1H), 3.89 (dd, *J* = 4, 1 Hz, 1H), 1.94 – 2.02 (m, 1H), 0.93 (d, *J* = 7 Hz, 3H), 0.79 (d, *J* = 7 Hz, 3H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>)  $\delta$  175.5, 157.9, 62.8, 29.6, 18.6, 15.9. LC-MS (ESI) m/z: 143 [M+H]<sup>+</sup>, 115. HRMS (ESI) m/z: [M+H]<sup>+</sup> Calcd for C<sub>6</sub>H<sub>11</sub>N<sub>2</sub>O<sub>2</sub> 143.0815; Found: 143.0814.

(S)-5-Methylimidazolidine-2,4-dione (H2i):

White solid. Yield 83%.  $[\alpha]_D^{20}$ -23.2 (*c* 10, CH<sub>3</sub>CN). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  10.58 (s, 1H), 7.88 (s, 1H), 4.02 (q, *J* = 7 Hz, 1H), 1.21 (d, *J* = 7 Hz, 3H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>)  $\delta$  176.9, 157.2, 53.4, 17.3. LC-MS (ESI) m/z: 115 [M+H]<sup>+</sup>. Data in accordance with literature [2].<sup>21</sup>

(S)-5-(2-(Methylthio)ethyl)imidazolidine-2,4-dione (H2j):

White solid. Yield 59%.  $[\alpha]_{D}^{20}$ -49.9 (*c* 10, CH<sub>3</sub>CN). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  10.66 (s, 1H), 8.02 (s, 1H), 4.10 (qd, *J* = 4, 1 Hz, 1H), 2.53 (m, 2H), 2.05 (s, 3H), 1.87 – 1.97 (m, 1H), 1.71 – 1.80 (m, 1H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>)  $\delta$  175.9, 157.4, 56.6, 31.0, 28.8, 14.5. LC-MS (ESI) m/z: 175 [M+H]<sup>+</sup>, 147, 127. Data in accordance with literature [2].<sup>21</sup>

### 3. NMR and HRMS Spectra



<sup>1</sup>H NMR spectrum of carbamoyl-L-phenylalanine (**H1a**)



<sup>13</sup>C NMR spectrum of carbamoyl-L-phenylalanine (H1a)



Mass spectrum of carbamoyl-L-phenylalanine (H1a)



<sup>1</sup>H NMR spectrum of (*S*)-5-benzylimidazolidine-2,4-dione (**H2a**)



<sup>13</sup>C NMR spectrum of (S)-5-benzylimidazolidine-2,4-dione (H2a)







Mass spectrum of (S)-5-benzylimidazolidine-2,4-dione (H2a)



<sup>1</sup>H NMR spectrum of (*S*)-5-(4-hydroxybenzyl)imidazolidine-2,4-dione (**H2b**)



<sup>13</sup>C NMR spectrum of (S)-5-(4-hydroxybenzyl)imidazolidine-2,4-dione (H2b)



HRMS spectrum of (S)-5-(4-hydroxybenzyl)imidazolidine-2,4-dione (H2b)



Mass spectrum of (S)-5-(4-hydroxybenzyl)imidazolidine-2,4-dione (H2b)



<sup>1</sup>H NMR spectrum of (*S*)-5-((1*H*-indol-2-yl)methyl)imidazolidine-2,4-dione (**H2c**)



<sup>13</sup>C NMR spectrum of (*S*)-5-((1*H*-indol-2-yl)methyl)imidazolidine-2,4-dione (**H2c**)



HRMS spectrum of (S)-5-((1H-indol-2-yl)methyl)imidazolidine-2,4-dione (H2c)



150 200 250 300 350 400 450 500 550 600 650 700 750 800 850 900 950 Counts vs. Mass-to-Charge (m/z)

0 100

Mass spectrum of (S)-5-((1H-indol-2-yl)methyl)imidazolidine-2,4-dione (H2c)



<sup>1</sup>H NMR spectrum of (S)-5-((2,5-dioxoimidazolidin-4-yl)methyl)-1H-imidazol-3-ium chloride (H2d)



<sup>13</sup>C NMR spectrum of (S)-5-((2,5-dioxoimidazolidin-4-yl)methyl)-1H-imidazol-3-ium chloride (H2d)



Mass spectrum of (S)-5-((2,5-dioxoimidazolidin-4-yl)methyl)-1H-imidazol-3-ium chloride (H2d)



<sup>1</sup>H NMR spectrum of (*S*)-5-phenylimidazolidine-2,4-dione (**H2e**)



<sup>13</sup>C NMR spectrum of (S)-5-phenylimidazolidine-2,4-dione (H2e)



Mass spectrum of (S)-5-phenylimidazolidine-2,4-dione (H2e)



<sup>1</sup>H NMR spectrum of (*S*)-5-isobutylimidazolidine-2,4-dione (**H2f**)



<sup>13</sup>C NMR spectrum of (*S*)-5-isobutylimidazolidine-2,4-dione (**H2f**)









<sup>1</sup>H NMR spectrum of (*S*)-5-((*R*)-1-hydroxyethyl)imidazolidine-2,4-dione (**H2g**)



<sup>13</sup>C NMR spectrum of (S)-5-((R)-1-hydroxyethyl)imidazolidine-2,4-dione (H2g)



Mass spectrum of (S)-5-((R)-1-hydroxyethyl)imidazolidine-2,4-dione (H2g)



<sup>1</sup>H NMR spectrum of (S)-5-isopropylimidazolidine-2,4-dione (**H2h**)



<sup>13</sup>C NMR spectrum of (S)-5-isopropylimidazolidine-2,4-dione (H2h)



HRMS spectrum of (S)-5-isopropylimidazolidine-2,4-dione (H2h)



Mass spectrum of (S)-5-isopropylimidazolidine-2,4-dione (H2h)



<sup>1</sup>H NMR spectrum of (*S*)-5-methylimidazolidine-2,4-dione (**H2i**)



<sup>13</sup>C NMR spectrum of (S)-5-methylimidazolidine-2,4-dione (H2i)







<sup>1</sup>H NMR spectrum of (S)-5-(2-(methylthio)ethyl)imidazolidine-2,4-dione (H2j)







Mass spectrum of (S)-5-(2-(methylthio)ethyl)imidazolidine-2,4-dione (H2j)

# 4. Chromatograms of compounds

Aca Openator		
Acq. Operator	•	STSTEM Seq. Line : 1
Acq. Instrument	:	HPLC 1260 Location : Vial 1
Injection Date	:	4/7/2024 3:32:06 PM Inj: 1
		Inj Volume : 10.000 µl
Method	:	C:\CHEM32\1\DATA\CHANG WEI JIN H2A 2024-07-04 15-30-36\HYDANTOIN.M (
		Sequence Method)
Last changed	:	4/7/2024 3:30:45 PM by SYSTEM
DAD1 A, Si	ig=	210,4 Ref=off (CHANG WEI JIN H2A 2024-07-04 15-30-36\001-0101.D)
mAU	326	
1750	di.	
1500		
1250 -		
1000 -		
750 -		
500-	2	9
250	ŧ	2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2
0	F.	
		5 10 15 20 25 min

Signal 1: DAD1 A, Sig=210,4 Ref=off

Peak #	RetTime [min]	Туре	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	2.403	BV	0.0480	20.58212	6.67029	0.1715
2	2.526	VB	0.0913	1.19494e4	2133.56299	99.5478
3	5.847	BB	0.1119	7.91664	1.07548	0.0660
4	7.089	BB	0.1312	25.78545	3.03190	0.2148
Total	s :			1.20037e4	2144.34066	

# Chromatogram and HPLC integrations of H2a.



Peak #	RetTime [min]	Туре	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	2.358	BV	0.0581	5826.54150	1543.05322	95.9929
2	2.590	VB	0.0581	118.87553	31.47240	1.9585
3	2.734	BB	0.0524	5.85003	1.77786	0.0964
4	2.973	BV	0.0584	9.89534	2.59918	0.1630
5	3.044	VV	0.0557	5.55367	1.48331	0.0915
6	3.240	VB	0.1110	14.20711	1.95022	0.2341
7	3.833	BB	0.0822	37.24251	6.98077	0.6136
8	6.945	BV	0.1061	15.36413	2.18387	0.2531
9	7.096	VB	0.1371	36.23384	3.94834	0.5970

Totals :

6069.76367 1595.44917

Chromatogram and HPLC integrations of H2b.





Peak #	RetTime [min]	Туре	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	2.438	BV	0.0583	8261.82910	2176.68433	99.6956
2	2.709	VB	0.0679	9.51642	2.05883	0.1148
3	2.980	BB	0.0703	15.70722	3.37788	0.1895

#### Totals :

8287.05273 2182.12103

#### Chromatogram and HPLC integrations of H2c.



```
Signal 1: DAD1 A, Sig=210,4 Ref=off
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Peak	RetTime	Туре	Width	Area	Height	Area
# 	[min]		[min] 	[mAU*s]	[mAU]	% 
1	2.298	BB	0.0909	1.19004e4	2074.88818	99.1798
2	3.198	BV	0.0828	9.44985	1.65101	0.0788
3	3.420	VB	0.0741	65.08628	13.05972	0.5424
4	4.629	BB	0.0908	6.53019	1.10785	0.0544
5	6.885	BB	0.1238	17.34209	2.15695	0.1445

```
Totals :
```

1.19988e4 2092.86372

Chromatogram and HPLC integrations of **H2d**.



Signal 1: DAD1 A, Sig=210,4 Ref=off

Peak I #	RetTime [min]	Туре	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	2.462	BV	0.0866	1.15082e4	2145.84326	98.9389
2	2.749	VV	0.0870	36.11869	5.93891	0.3105
3	2.897	VV	0.0545	7.37558	1.93413	0.0634
4	2.999	VB	0.0673	49.22173	11.20341	0.4232
5	7.148	BB	0.1362	30.71308	3.43865	0.2640
Totals	5:			1.16317e4	2168.35836	

Chromatogram and HPLC integrations of **H2e**.

	Sig=	210 4 8	of-o	ff (001	1 0101	D)															
DADTA																					
mAU –	<u>8</u>																				
1750 -	di.																				
1500 -																					
1250 -																					
1000 -																					
750 -																					
500 -																					
250 -	433	573	927	846	221																
0	- qil	nini	4	ю,	1																
			5		1	1	10		i	1		15			20			25	1		min

Signal 1: DAD1 A, Sig=210,4 Ref=off

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	2.433	BB	0.0740	15.73734	2.96331	0.1829
2	2.594	BB	0.0675	8455.26465	1993.84448	98.2673
3	3.573	VB	0.1246	21.32673	2.33320	0.2479
4	3.935	BB	0.1093	22.54987	3.01346	0.2621
5	4.927	BB	0.1084	21.42057	2.82658	0.2490
6	5.846	BB	0.1085	16.58592	2.40312	0.1928
7	7.221	BB	0.1300	51.46523	6.12673	0.5981

Totals :

8604.35031 2013.51088

#### Chromatogram and HPLC integrations of H2f.



Signal 1: DAD1 A, Sig=210,4 Ref=off

Peak	RetTime	Туре	Width	Area	Height	Area	
#	[min]		[min]	[mAU*s]	[mAU]	%	
1	2.261	BV	0.0692	55.54348	11.03689	0.8953	
2	2.355	VV	0.0588	6015.70117	1639.38928	96.9619	
3	2.720	BV	0.0812	16.12285	2.97426	0.2599	
4	2.886	VB	0.0849	9.75303	1.65237	0.1572	
5	3.451	BB	0.1131	13.81607	1.69416	0.2227	
6	3.806	BB	0.1069	15.91802	2.13667	0.2566	
7	4.705	BB	0.1313	36.14483	3.65674	0.5826	
8	5.659	BB	0.1050	16.47449	2.43191	0.2655	
9	7.001	BB	0.1359	24.71371	2.77591	0.3983	

Totals :

6204.18764 1667.74819

#### Chromatogram and HPLC integrations of H2g.



Signal 1: DAD1 A, Sig=210,4 Ref=off

Peak	RetTime	Туре	Width	Area	Height	Area		
#	[min]		[min]	[mAU*s]	[mAU]	%		
1	2.422	BV	0.0646	7871.98828	1968.86609	98.3380		
2	2.851	VB	0.1019	23.49626	3.19237	0.2935		
3	3.308	BV	0.0706	8.47418	1.81192	0.1059		
4	3.405	VB	0.0874	11.54817	1.88816	0.1443		
5	3.764	BB	0.1185	15.21869	1.80299	0.1901		
6	4.635	BB	0.1022	61.07473	8.66549	0.7630		
7	5.590	BB	0.1048	13.23519	1.96022	0.1653		

Totals :

8005.03551 1988.18724

Chromatogram and HPLC integrations of H2h.



Signal 1: DAD1 A, Sig=210,4 Ref=off

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	2.400	BV	0.0699	8741.37305	2044.75537	97.5539
2	2.889	VV	0.1088	30.63241	3.76871	0.3419
3	3.148	VV	0.1633	28.19417	2.23343	0.3146
4	3.451	VB	0.1410	32.44205	3.07816	0.3621
5	3.832	BB	0.1322	16.16709	1.71199	0.1804
6	4.789	BB	0.1154	16.98713	2.07756	0.1896
7	5.693	BB	0.1082	14.48612	2.05597	0.1617
8	7.058	BB	0.1308	49.59040	5.97726	0.5534
9	22.902	BB	0.3420	30.68933	1.32613	0.3425

Totals :

8960.56175 2066.98460

Chromatogram and HPLC integrations of H2i.



Signal 1: DAD1 A, Sig=210,4 Ref=off

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
		-				
1	2.108	BV	0.2010	21.04398	1.64822	0.2402
2	2.441	VV	0.0691	8506.89258	2020.25354	97.0935
3	2.889	VV	0.1384	45.47044	4.26387	0.5190
4	3.145	VV	0.1424	23.84449	2.23623	0.2721
5	3.360	VB	0.1439	49.40802	4.57715	0.5639
6	3.835	BB	0.1245	15.77759	1.79521	0.1801
7	4.726	BB	0.1003	84.84641	12.33343	0.9684
8	5.708	BB	0.1068	14.26201	2.05921	0.1628
Total	s :			8761.54551	2049.16687	

Chromatogram and HPLC integrations of H2j.

### 5. References

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