**Electronic supporting information for “Mechanochemical bottom-up synthesis of phosphorus-linked, heptazine-based carbon nitrides using sodium phosphide”**

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# **1. General methods and materials**

**Caution: Sodium phosphide readily decomposes in the presence of moist air, and protic organic solvents to produce toxic and pyrophoric phosphane gas, all workups should be conducted in a glovebox or well ventilated fumehood.**

All manipulations were conducted in an argon-filled glovebox with O2 and H2O levels both at and below 0.5 ppm. Dimethoxyethane (Millipore-Sigma) was dried and distilled under argon over sodium metal with a benzophenone radical indicator. All other solvents were used as received. Sodium phosphide was made using a previously reported method[1] from red phosphorous (Millipore-Sigma), sodium metal (ThermoFischer) and naphthalene (Millipore-Sigma) using standard Schlenk techniques and an argon-filled glovebox. All nanomaterial workups can be conducted under normal, ambient conditions, unless otherwise stated. All chemicals were used as received unless otherwise stated.

Melem (1), potassium cyanmelurate (2), and tri-chloro tri-*s*-heptazine (3) were synthesised using methods previously reported in the literature.[2] To briefly summarize, melem (1) was synthesised by thermal treatment of melamine (Millipore-Sigma) in a covered alumina crucible at 550 °C for 4 h at a ramp rate of 5 °C/min.

Potassium cyanmelurate, (2)

Melem (3.4 g, 15.8 mmol) was then refluxed in 35 mL of 2.5 M aqueous solution of potassium hydroxide at 110 °C for 4 h, hot filtered to remove any insoluble material, cooled on ice to 0 °C before precipitating the product with 95% ethanol (5.3 g, 15.8 mmol).

1,5,9-tri-chloroheptazine (3)

Potassium cyanmelurate (5.3 g, 15.8 mmol) was ground together with phosphorus pentachloride (9.7 g, 46.9 mmol) in a mortar and pestle for ca. 30 s until homogeneous. This powder was added to a 100 mL Schlenk tube using a powder funnel, while under a positive flow of argon. The Schlenk tube was sealed with a glass stopper and heated in an oil bath at 130 °C for 10 h, lifting the stopper every hour under a strong active flow of argon to release overpressure. The resulting pale-yellow powder was washed via Soxhlet extraction in dry toluene for 24 h and the toluene fraction was filtered through cotton before drying on a RotoVap. The pale-yellow tri-chloroheptazine (3) (2.6 g, 9.4 mmol) was stored protected from light in an argon-filled glovebox until use.



**Figure S1:** Synthesis of 1,5,9-tri-chloroheptazine from melamine.

## **1.1. Powder X-ray diffraction measurements**

Powder X-ray diffraction patterns were acquired over a range of 5° to 50° 2θ with a step size of 0.02° 2θ and an integration time of 0.5 s using a Bruker D8 Advantage X-Ray Diffractometer equipped with a Cu-Kɑ (λ = 1.5418 Ǻ) source, LinxEye detector, and a Ni filter. Measurements were performed using a Si zero background holder. PXRD patterns were analysed using X’Pert HighScore Plus.

## **1.2. X-ray photoelectron spectroscopy measurements**

Samples were analysed on a Fischer Scientific Kα X-Ray spectrometer with an excitation source of Al Kα= 1486.6 eV. The binding energies were corrected by referencing the C 1s line to 284.80 eV. A spot size of 400 μm was used, running 5 survey scans at 200 mV for 50 ms residence times, and 10 scans for specific elements, also at residence times of 50 ms. Deconvolution and peak position were determined using Avantage processing software.

## *1.3.* Solution *13C NMR*

13C solution NMR was conducted using a Bruker AVIIIHD 500 MHz NMR spectrometer. Solid sample (~10 mg) was dissolved in 700 µL of DMSO-d6 (Millipore Sigma). Spectra were analysed using MestReNova (v14.2.1)

## **1.4. 31P MAS NMR analysis of g-h-PCN and g-h-PCN300**

Direct polarization 31P solid-state NMR was performed using a Varian 400 MHz VNMRS widebore spectrometer operating at a 1H frequency of 399.76 MHz with a 4 mm double-resonance probe spinning at 13 kHz. The 31P excitation pulse was 2.2 μs long and 1H decoupling was performed using SPINAL-64 decoupling at 110 kHz. For all phosphorylated materials, a recycle delay of 150 s was used and 128 transients were acquired of each sample, for a total experiment time of just under 5.5 h. Sample integration was compared to an external ammonium dihydrogen phosphate standard acquired in one single scan after 2.5 min in the magnet (to ensure full polarization).

**1.5.** **Prediction of NMR spectra through periodic DFT calculations**

Periodic density functional theory calculations were performed using the plane wave code, CASTEP (v20.11).[3] Input files were generated using the cif2cell program.[4] The plane wave kinetic energy cut off was converged at 700 eV, and the core regions of electron density were described by On-The-Fly ultrasoft pseudopotentials from the CASTEP library. Monkhorst-Pack grids[5] with a k-point spacing of 0.07 2π Å-1 were used to sample the Brillouin zone. Structural optimizations were performed suing the PBE[6] functional with the Grimme D3 dispersion correction.[7] Input structures for the paddlewheel monomer, a corrugated bulk, and a planar bulk structure of g-h-PCN were created by adapting the N-bridged equivalents published by Tragl *et al.*,[8] Garcia and Kroll,[9] and the phase 1 structure by Wang *et al.*,[10] respectively. All structures were adapted by replacing the bridging nitrogen atoms with phosphorus. Predictions of the 31P MAS NMR shielding tensors were calculated using the gauge-inducing projector augmented waves (GIPAW) method[11, 12] available in CASTEP. Again, core regions of electron density were described by pseudopotentials from the CASTEP library. The online tool MagresView v1.6.2[13] was used to analysis the magnetic shielding data. The isotropic chemical shifts, δiso, of the materials were obtained using, δiso = σref - σiso. Where σiso is the isotropic magnetic shielding of the molecule studied and σref is the shielding of a given reference species. Ammonium dihydrogen phosphate (ADHP) was chosen as the reference species with its experimental peak known to be 0.9 ppm. The magnetic shielding of ADHP was calculated using the outlined calculation strategy with the same plane wave energy cutoff and k-point spacing. The initial structure of ADHP was obtained from the Cambridge Structural Database, reference code VITPIA.[14]

**1.6 FTIR-ATR measurement**

Samples were analysed via FTIR using a Fourier Diagform-Infrared Attenuated Total Reflection PerkinElmer UATR Two Spectrometer through a range of 400 to 4000 cm-1, with a resolution of 1 cm-1. OPUS 7.5 was used to process and analyse the resulting data.

## **1.7 Scanning tunneling electron microscopy- electron energy loss spectroscopy (STEM-EELS)**

STEM-EELS measurements were performed using a ThermoFisher Talos F200X transmission electron microscope operated at 200 keV, equipped with a high brightness XFEG Schottky source and a Gatan Enfinium ER 970 EELS spectrometer. The core-loss spectra were corrected using the zero-loss peak (ZLP) shift correction from a Dual-EELS measurement. The dwell time for EELS acquisition on every spectrum image was 0.5 sec, with a drift correction at every row. Relative thickness was measured from an area of 100 nm × 100 nm

## **1.8. Photoluminescence measurements**

The photoluminescence (PL) emission spectra and time-resolved PL (TRPL) spectra were recorded in a PTI QuantaMaster 500 spectrofluorometer and TimeMaster Lifetime fluorometer (Horiba, France). PL excitation wavelength was fixed at 350 nm with a slit overture of 2.0 nm and the decay was obtained with a LED with a fixed excitation wavelength of 369 nm and an overture of 0.2 nm for 30 s.

## **1.9. Photoelectrochemical measurements**

A standard three-electrode system was used to determine the photo-electrochemical properties and band structure, using Pt wire and Ag/AgCl as counter and reference electrodes, respectively. The working electrode was prepared on fluorine-doped tin oxide (FTO) glass (2.5 x 2.5 cm), which was cleaned by sonication in acetone for 3 h and then dried. The FTO slide was spin-coated with 5 ml of slurry, which was obtained by the sonication of 20 mg of photocatalyst in ethanol for 3h. The transient photocurrent response was performed on an Autolab PGSTAT204 electrochemical workstation with a 0.5 M aqueous solution of Na2SO4 as the electrolyte. The photocurrent was measured at a bias voltage of 0.8 V under solar light irradiation (150 W xenon lamp) with a 10 s light on-off cycle.

# **2. Results and Discussion**

**2.1 13C NMR spectrum**

Chart

Description automatically generated

**Figure S2:** 13C NMR of 1,5,9-tri-chloroheptazine in DMSO-d6.

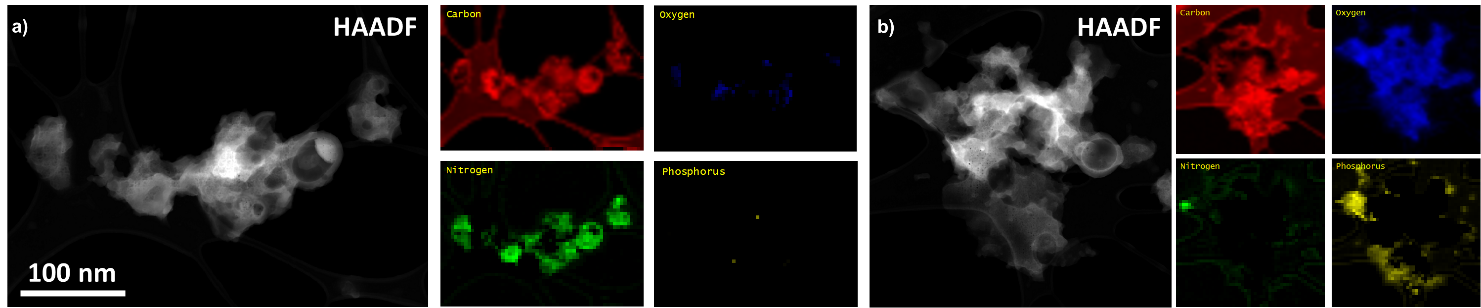
**2.2 FTIR-ATR analysis**

Chart, line chart

Description automatically generated

**Figure S3:** FTIR-ATR of g-h-PCN (green) and g-h-PCN300 (teal).

**2.3 STEM-EELS analysis**

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**Figure S4:** STEM-HAADF images (left) and EELS maps (right) for the characteristic elements presented on graphitic carbon networks. a) g-h-PCN (no annealing) and b) g-h-PCN300 (annealing at 300 °C under argon).

## **2.4 Photochemical analysis**

Graphical user interface, chart, histogram

Description automatically generated

**Figure S5**: a) Diffuse reflectance spectroscopy b) photoluminescence c) photocurrent and d) Nyquist measurements of g-C3N4 (purple), g-h-PCN (blue), , and g-h-PCN300 (green).

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