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# The Cucurbit[8]uril effect on the properties of Oroxin A

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## Abstract

In this study, we investigated host–guest interactions between Oroxin A (OA) and cucurbit[8]uril (Q[8]) using  $^1\text{H}$  NMR, MS, UV-Vis and IR spectroscopy. The results showed that OA and Q[8] formed an inclusion compound (OA@Q[8]) with a molar ratio of 1:1 and a binding constant of  $1.299 \times 10^7 \text{ L}\cdot\text{mol}^{-1}$ . In addition, the effect of Q[8] on the properties of OA was investigated through comparative experiments. The solubility of OA in water increased 22.47-fold when the concentration of Q[8] was  $1 \times 10^{-4} \text{ mol}\cdot\text{L}^{-1}$ . Q[8] hardly affected the antioxidant capacity of OA, while the cumulative release of OA in gastric juice increased 2.3-fold after forming the inclusion compound with Q[8].

## Keywords

Oroxin A; Cucurbit[8]uril; Host-guest interaction; Inclusion complex; Properties

## Introduction

Cucurbit[n]urils (Q[n]s) are a family of macrocyclic cage compounds synthesized by the condensation of glycoluril and formaldehyde in a strong acid solution [1–3]. As a consequence of the specific structural features of Q[n]s, which have two hydrophilic portals decorated with partially negatively charged carbonyl groups and a hydrophobic cavity [4], cucurbit[n]urils are able to form host–guest complexes with a range of drugs [5–7]. These complexes involve three main intermolecular forces: a hydrophobic effect, hydrogen bonding and ion-dipole interactions at the carbonyl portals [7–9]. The high thermal stability [10], ease of synthesis [11], general absence of cytotoxicity or toxicity [12,13] and their good molecular recognition and binding constants [14] have shown that Q[n]s are ideal drug carriers [15,16]. Moreover, Q[n]s can enhance physical stability [17,18] and increase solubility [19,20] of drug molecules after complexation.

Oroxin A (OA, baicalein-7-O-glucoside, Figure 1(A)) is one of the active ingredients isolated from the traditional herbal medicine *Oroxylum indicum* (L.) Kurz of Asian countries [21–23]. Accumulating studies have shown the beneficial biological effects of OA, which include antioxidant, antidiabetic, anticancer, antibacterial, anti-inflammatory and anti-viral properties [24–29]. Herein, we selected Q[8] as a host molecule and investigated its host–guest interactions with OA, as well as its effect on the properties of OA. Our results provide an approach and theoretical basis for the development and utilization of Oroxin A.

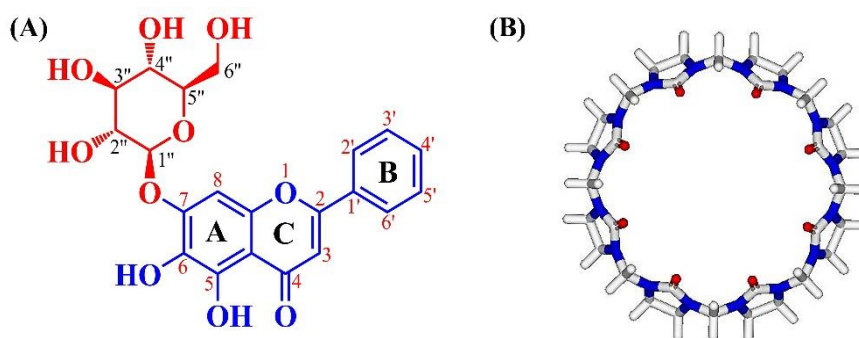
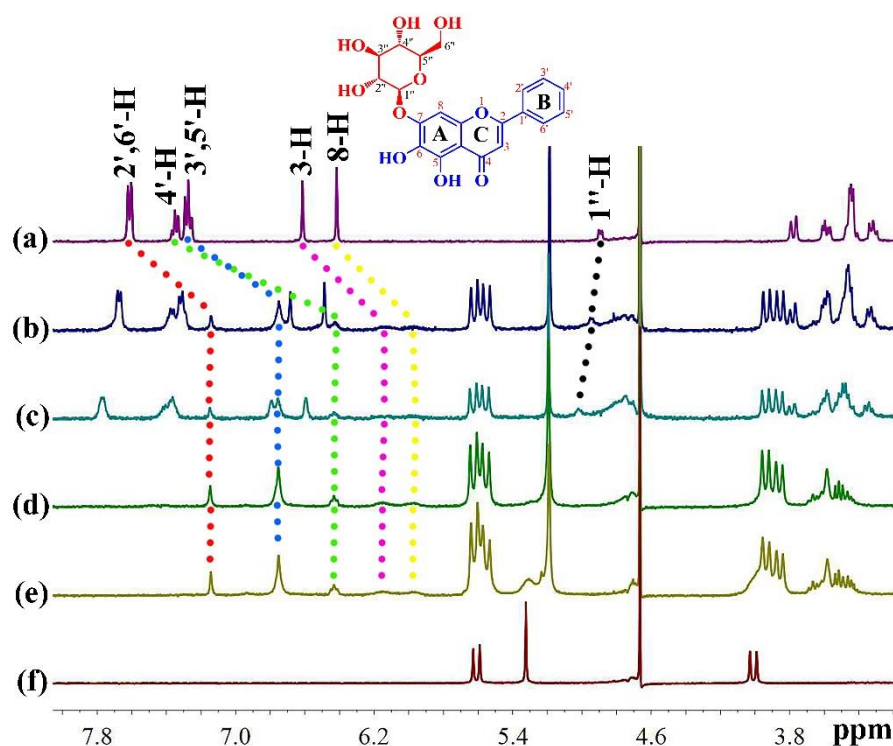


Figure 1 The molecular structures of OA and Q[8]

## Results and discussion

### 2.1 Host-guest interactions

The host-guest interaction can be effectively observed using  $^1\text{H}$  NMR spectroscopy, and the mode of action of the cucurbit[n]uril-guest can be inferred from the chemical shift changes of the guest proton resonance peaks.  $^1\text{H}$  NMR titration experiments were performed in  $\text{D}_2\text{O}$  containing 10% DMSO by volume at room temperature. As shown in Figure 2, upon addition of Q[8], some of the peaks due to protons on the OA aglycone shifted to higher field, while peaks due to the glycosidic proton shifted to lower field. At the same time, the proton peaks on Q[8] were split, indicating that OA interacted with Q[8]. When the host-guest molar ratio was 1:1, all of the OA aglycone proton peaks moved to higher field, indicating entry into the cavity of Q[8]. Proton peaks due to the glycosidic -H and glucose were shifted to lower field, indicating that they were located at the portal of Q[8].

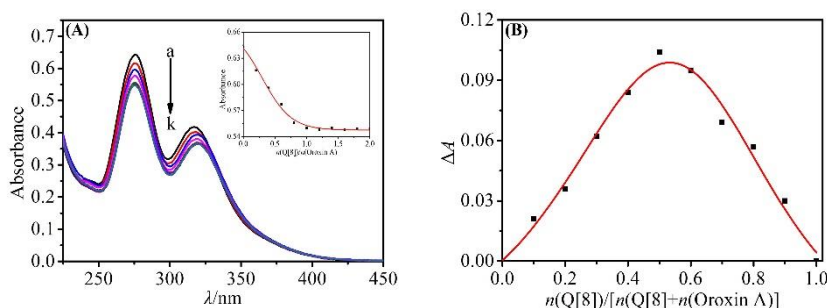


**Figure 2**  $^1\text{H}$  NMR titration of OA with Q[8] were performed in  $\text{D}_2\text{O}$  containing 10% DMSO by volume, OA ( $500 \mu\text{mol}\cdot\text{L}^{-1}$ ) upon the addition of different molar equivalents of Q[8]: (a) 0, ( b ) 0.35, (c) 0.44, (d) 1.03, (e) 1.60 and (f) neat Q[8]

**Table 1** Changes in the  $^1\text{H}$  NMR chemical shifts

$^1\text{H}$ nucleus	$\Delta\delta/\text{ppm}$
2',6'-H (cycle B)	0.47
4'-H (cycle B)	0.92
3',5'-H (cycle B)	0.52
3-H (cycle C)	0.46
8-H (cycle A)	0.45
1''-H (glycoside)	-0.13

To further determine the host–guest ratio of the inclusion complex formed by Q[8] and OA, their interaction was investigated using UV-visible absorption spectroscopy via a molar ratio method and Job's method. **Figure 3(A)** shows the UV-visible absorption spectra of the interaction between Q[8] and OA. It can be seen that the UV absorption of OA at 275 nm and 316 nm decreased significantly as the concentration of Q[8] was increased. When  $n(\text{Q}[8])/n(\text{OA}) = 1$ , there was a clear transition of absorbance of the system. Upon further addition of Q[8], the absorption value of the system tended to be constant, indicating formation of a 1:1 complex with a binding constant  $K = 1.299 \times 10^7 \text{ L}\cdot\text{mol}^{-1}$ . The result of the Job's plot also confirmed the combination of Q[8] and OA in a 1:1 mode (**Figure 3(B)**).



**Figure 3** (A) The UV-Vis absorption spectra recorded for OA in the presence of Q[8] ( $c(\text{Q}[8])$ : a–k, 0, 0.2, 0.4, 0.6, ...,  $2.0 \times 10^{-5} \text{ mol}\cdot\text{L}^{-1}$ ) and (B) Job's plot obtained for OA in the presence of Q[8](B).

Figure 4 shows the IR spectra recorded for Q[8] (a), OA (b), a physical mixture of Q[8] and OA ( $n(\text{Q}[8]):n(\text{OA}) = 1:1$ ) (c) and the OA@Q[8] inclusion complex (d). Curve (c) contains characteristic peaks of curves (a) and (b) without interaction in the physical mixture. Comparing spectra (c) and (d), the peaks at 1617.41, 1482.23 and 1451.06  $\text{cm}^{-1}$  due to stretching vibrations of the two benzene rings disappeared, and the peak at 1079.42  $\text{cm}^{-1}$  due to C–O stretching vibrations was obviously weakened in the inclusion complex, which were caused by Q[8].

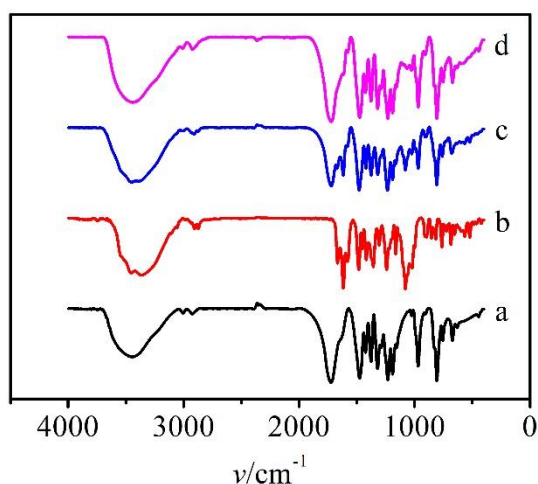


Figure 4 IR spectra recorded for (a) Q[8], (b) OA, (c) a physical mixture of Q[8] and OA, and (d) the OA@Q[8] inclusion complex

The mass spectrum of the OA@Q[8] inclusion complex features the parent ion peak at  $m/z$  1783.5716  $[\text{M}+\text{Na}]^+$  (calcd. 1783.4983  $[\text{M}+\text{Na}]^+$ ) (Figure S1), further supporting the formation of a 1:1 inclusion complex between OA and Q[8]. The possible host–guest mode is shown in Figure 5.

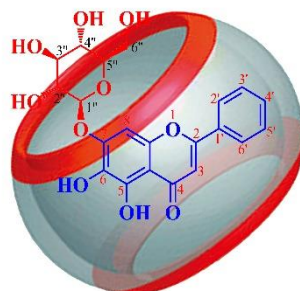


Figure 5 The possible interaction mode for OA@Q[8]

## 2.2 The effect of OA on the properties of cucurbit[8]uril

### Phase-solubility

Phase solubility studies were conducted to investigate the solubility of OA in the presence of Q[8]. As can be seen from Figure 6, the solubility of OA in water is very poor ( $4.62 \times 10^{-6} \text{ mol}\cdot\text{L}^{-1}$ ). The solubility of OA increased linearly in water with addition of Q[8]. When the concentration of Q[8] was  $1.0 \times 10^{-4} \text{ mol}\cdot\text{L}^{-1}$ , the solubility of OA was increased 22.47-fold. The solubility curve equation was  $S = 0.01c + 0.0575$ ,  $R^2 = 0.9986$ .

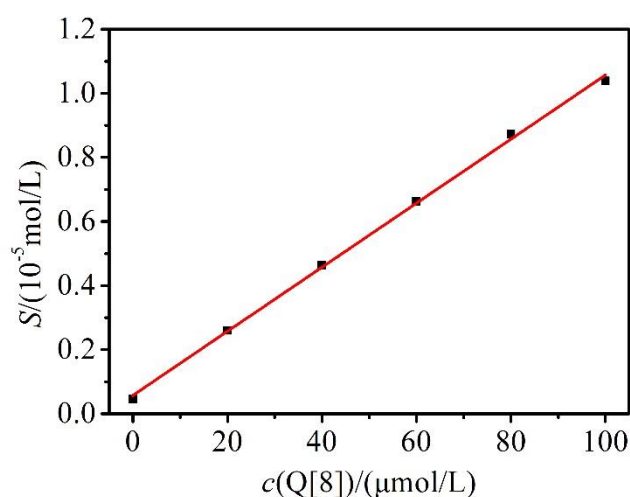


Figure 6 The phase solubility graph obtained for OA in Q[8] aqueous solution at  $\lambda = 275 \text{ nm}$

### Antioxidant activity

OA has strong antioxidant activity and effectively eliminates  $\text{ABTS}^+$  radicals. If the antioxidant activity of OA was decreased significantly by formation of the OA@Q[8] complex, the medicinal value of OA would be seriously affected. Figure 7 shows the results for OA and OA@Q[8] scavenging of  $\text{ABTS}^+$  radicals in the range of 1–20  $\mu\text{mol}\cdot\text{L}^{-1}$ . The  $\text{IC}_{50}$  values of OA and OA@Q[8] were  $4.65 \times 10^{-6} \text{ mol}\cdot\text{L}^{-1}$  and  $4.80 \times 10^{-6} \text{ mol}\cdot\text{L}^{-1}$ , respectively, which indicates that Q[8] does not affect the antioxidant activity of OA.

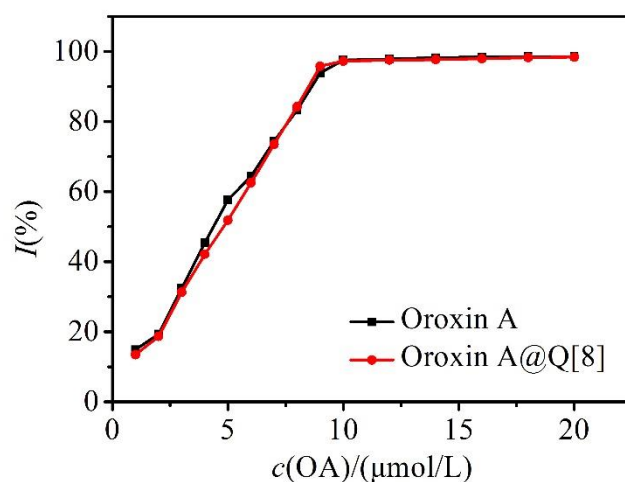


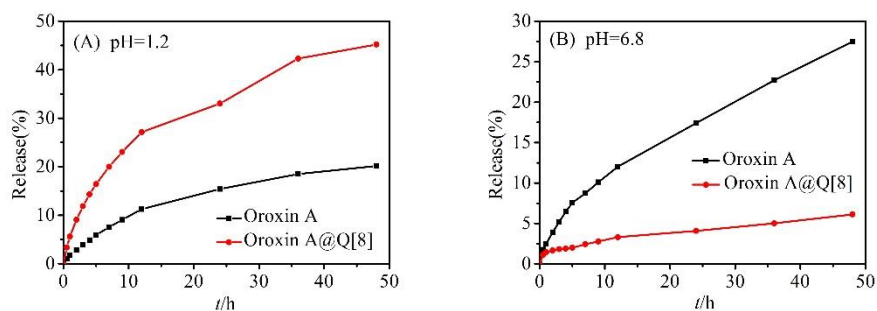
Figure 7 The clearance rate curve of  $\text{ABTS}^+$  upon increasing the concentration of OA and the OA@Q[8] inclusion complex

### Drug release *in vitro*

Figure 8 shows the cumulative release of OA and OA@Q[8] inclusion complex in artificial gastric juice (pH = 1.2) and artificial intestinal juice (pH = 6.8). It can be seen from Figure 8(A) that release of OA@Q[8] inclusion compound in artificial gastric juice was much higher than that of OA after 12 h. The cumulative release of OA and OA@Q[8] reached 11.25% and 27.15%, respectively, after 12 h. After 48 h, Q[8] increased the measured cumulative release of OA in artificial gastric juice by 2.3-fold.

In artificial intestinal fluid (Figure 8(B)), the release rate of OA was faster than that of OA@Q[8]. After 12 h, the cumulative release of OA was 12.02%, while there was only 3.31% release of OA@Q[8].





**Figure 8** The release curves of OA and OA@Q[8]

## Conclusions

In summary, the experimental results showed that OA and Q[8] formed a host–guest complex in a ratio of 1:1. The aglycone of OA enters the cavity of Q[8] and the glucose is located at the portal of Q[8], with a binding constant of  $1.299 \times 10^7 \text{ L}\cdot\text{mol}^{-1}$ . The solubility of Oroxin A was increased 22.47-fold when the concentration of added Q[8] was  $1.0 \times 10^{-4} \text{ mol}\cdot\text{L}^{-1}$ . The results of the UV absorption spectrum analysis showed that Q[8] enhanced the cumulative release of OA in artificial gastric juice by 2.3-fold, but had no effect on its antioxidant activity.

## Supporting Information

Supporting Information File 1:

Title: Apparatus, Materials and methods.

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